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ABSTRACT

Purpose. *B. cepacia* infection in cystic fibrosis (CF) patient is associated with increased morbidity and mortality. Temocillin (TMO), a semisynthetic 6-*o*-methoxy β -lactam, has already been successfully used in pilot studies for the treatment of pulmonary infections in CF patients infected with *B. cepacia*. We determined the susceptibility of well characterized panel strains and clinical isolates of *B. cepacia complex* to TMO in comparison with 3 other β -lactams used in CF patients: meropenem (MER), ceftazidime (CTZ), and piperacillin/tazobactam (PTZ).

Methods. The MICs were measured by microdilution in Mueller-Hinton broth using the CLSI method. *Pseudomonas aeruginosa* ATCC 27853 and *Escherichia coli* ATCC 25922 were included as control strains. CLSI breakpoints for Enterobacteriaceae were used for MER, CTZ, and PTZ and that of Fuchs *et al.* (1985 Eur J Clin Microbiol 4:30-33) for TMO.

Results. The table below shows the MIC50 and MIC90 obtained on 100 strains of *B. cepacia* from 9 genomovars of the complex (n = 30 and 35 for genomovars II and III, respectively, and 5 for genomovars I, and IV to IX).

β -lactam	MIC50 μ g/ml	MIC90 μ g/ml	Breakpoint μ g/ml	% susceptibility
TMO	8	32	16	81
MER	4	16	4	66
CTZ	4	> 128	8	70
PTZ	16	> 128	16	51

The susceptibility pattern was similar among the different genomovars. Interestingly, 7 strains were susceptible only to TMO, while 6/35 *B. multivorans* and 7/30 *B. cenocepacia* were resistant to all the antimicrobials tested.

Conclusion. TMO was active against more strains than any of the 3 other comparators. Combined with the results of the clinical pilot studies, our data suggest a potential therapeutic use for TMO in CF patients infected with *B. cepacia complex* strains.

METHODS

MICs were measured by broth microdilution using the CLSI method. *Pseudomonas aeruginosa* ATCC 27853 and *Escherichia coli* ATCC 25922 were included as control strains. All control values were within recommendations. CLSI breakpoints for Enterobacteriaceae were used for MER, CTZ and PTZ, and the value proposed by Fuchs *et al.* for TMO (**je donnerais les valeurs !!!**)

INTRODUCTION and OBJECTIVE

CF is caused by mutations in a gene encoding a protein named CFTR which functions as a chloride channel in epithelial membranes [Collins, 1992]. The most dramatic changes are observed in CF airways causing chronic pulmonary infections with surprisingly few bacterial pathogens: *Pseudomonas aeruginosa* (most common isolate), *Staphylococcus aureus*, *Stenotrophomonas maltophilia* and *Burkholderia cepacia* [CFF Annual Report, 2002].

The *B. cepacia complex* (*Bcc*) represents at least 9 distinct bacterial species or "genomovars". *Bcc* are found in soil and on plants. The identification of unique *Bcc* strains CF sputum isolates implies acquisition from unknown reservoirs. The global prevalence rate among CF patients is around 3% (with up to 8% in adults) (Saiman and Siegel, 2003). Infections with *Bcc* are regarded as crucial for CF patients because in about one third of patients it causes a rapid decline of lung function, with as consequence, a dramatic reduction of life expectancy (up to 50%) (Corey and Farewell, 1996).

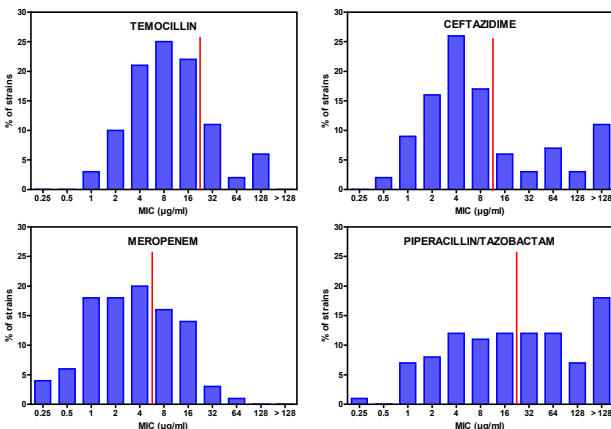
Although temocillin has already been used in a pilot clinical studies (Taylor *et al.*, 1992) with success for the treatment of *Bcc* infections in CF patients, only a few *in vitro* susceptibility data are available.

Our aim was, therefore, to determine the MICs of antibiotic used in CF patients (meropenem [MER], ceftazidime [CTZ], and piperacillin/tazobactam [PTZ]) in comparison with that of temocillin (TMO) towards a well characterized panel of laboratory strains and clinical isolates of *B. cepacia complex*.

RESULTS

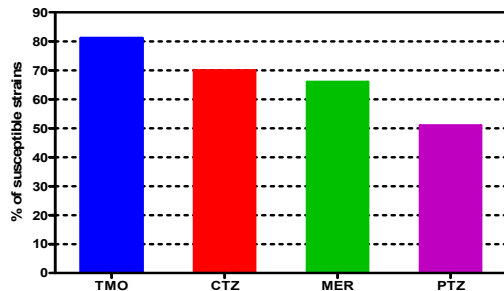
Figure 1 shows the MIC distributions of TMO, CTZ, MER, and PTZ against all the 100 strains of *B. cepacia complex*.

The red line correspond to the CLSI breakpoint for CTZ, MER, and PTZ and that of Fuchs *et al.* for TMO



- No major differences were observed among the different genomovars
- No significant differences between the susceptibility patterns of panel strains and clinical isolates were observed

Figure 2 shows the global susceptibility of all strains to the 4 different β -lactams investigated



Some strains were extremely resistant while 7 strains were susceptible only to temocillin. Table 1 describes these particular strains

strains	nbr of strains	Genomovars
resistant to all antibiotics	13	II (6) and III (7)
susceptible only to TMO	7	I (1), III (5), and VI (1)
CTZ	3	II
MER	1	V
PTZ	1	II

CONCLUSIONS

- TMO active against more *B. cepacia complex* strains compared to other clinically used β -lactams.
- These results, combined with those of pilot clinical studies, suggest a potential advantage of TMO in *B. cepacia* infected CF patients

REFERENCES

- Corey M, Farewell V. Determinants of mortality from cystic fibrosis in Canada. Am J Epidemiol 1996; 143:1007-17.
- Cystic Fibrosis Foundation Patient Registry Annual Report 2001, Cystic Fibrosis Foundation, Bethesda, Maryland, USA, 2002.
- Fuchs PC, Barry AL, Thomsberry C, Jones RN. Interpretive criteria for temocillin disk diffusion susceptibility testing. Eur J Clin Microbiol. 1985 4(1):30-3
- Saiman L, Siegel J. Cystic Fibrosis Foundation. Infection control recommendations for patients with cystic fibrosis: microbiology, important pathogens, and infection control practices to prevent patient-to-patient transmission. Infect Control Hosp Epidemiol. 2003 24:56-62.
- Taylor RFH, Gaya H, Hodson ME. Temocillin and cystic fibrosis: outcome of intravenous administration in patients infected with *Pseudomonas cepacia*. J Antimicrob Chemother 1992; 29:341-4.