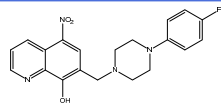


## INTRODUCTION AND OBJECTIVES

Because of the alarming increase of resistance to current antibiotics and of the lack of novel drugs acting on multiresistant strains, new therapeutic approaches need to be explored, among which the inhibition of virulence mechanisms. T3SS is a major virulence factor in *Pseudomonas aeruginosa* (*P.a*) that allows the translocation of effector proteins from the bacterial cytoplasm directly into the eukaryotic host cells, contributing to the establishment and dissemination of acute infections [1]. The 8-hydroxyquinolines have shown inhibitory activity against T3SS of *Yersinia* and *Chlamydia* spp [2]. In this study, we evaluated the activity of INP1855 against *P.a* in vitro and in vivo.

### INP1855 (8-hydroxyquinoline)



MW = 382.4  
pK<sub>a</sub> = 5.75 and 8.02 (zwitterionic at pH 7)  
log P = 3.59  
log D<sub>pH7</sub> = 2.23

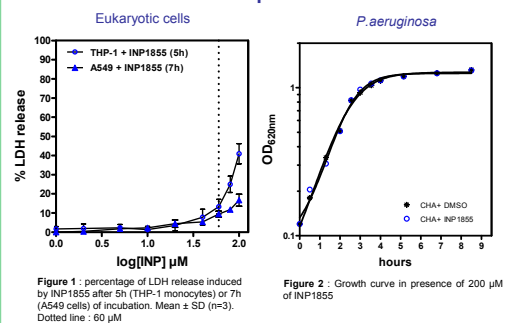
calculated with Reaxys

## MATERIAL AND METHODS

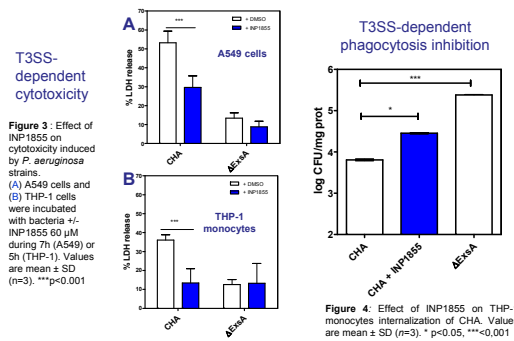
- Bacterial strains:** CHA (clinical isolate with high expression of T3SS) and CHAΔExsA (deletion of T3SS regulon).
- Eukaryotic cells:** human alveolar epithelial A549 cells and THP-1 monocytes. **Mice:** C57BL6. **Chemical:** INP1855
- Growth inhibition assay:** Optical density (620 nm) of the bacterial suspension
- Cell viability (eukaryotic cells):** lactate dehydrogenase (LDH) release
- Motility assay (bacteria):** swarming in LB agar 0.5% plate.
- Internalization assay:** Bacteria pre-treated with INP1855 (3 h) and opsonised with human serum (1 h). Phagocytosis initiated with 10 CFU/cell for 2 h (THP-1 cells) in medium containing 60 μM of INP1855 (elimination of non-phagocytosed bacteria by incubation with 100 μg/mL gentamicin (1 h). After washing, cells were lysed in water and aliquots plated for CFU counting [3].
- T3SS transcription assay:** (1) the bioluminescence of the reporter strain CHA pC lux (constructed by fusion of *luxCDABE* to the promoter of the regulator operon *exsCEBA* [4]) in the presence of THP-1 cells or A549 cells over time of incubation with 60 μM of INP1855, (2) the levels of T3SS genes expression by RT-PCR in CHA strain in the presence of INP1855 60 μM.
- T3SS functionality:** Detection of ExoS by Western Blot
- In vivo assay:** Mouse model of acute lung aggression induced by CHA with co-institution of bacteria and INP1855 200 μM. After 16 h, mice were sacrificed and different parameters were evaluated (weight gain, bacterial burden, lung lesion, bacterial dissemination and bronchoalveolar lavage cellularity).

## RESULTS

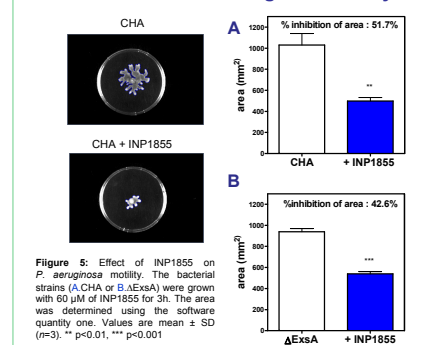
### INP1855 is not toxic for eukaryotic cells up to 60 μM, neither for bacteria at 200 μM



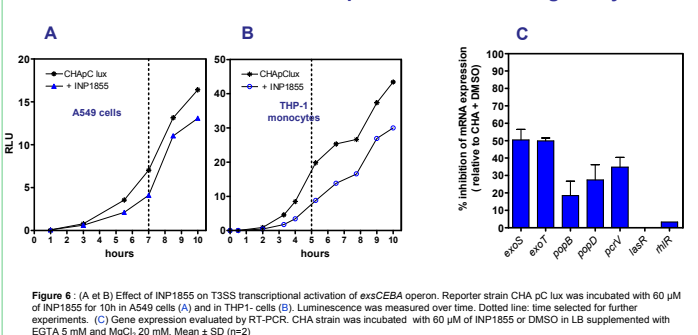
### INP1855 decreases T3SS-mediated virulence



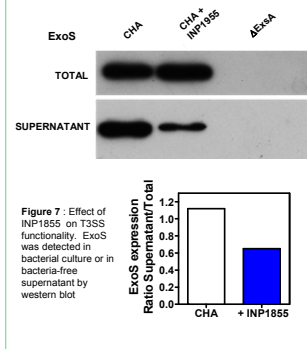
### INP1855 reduces flagellar motility



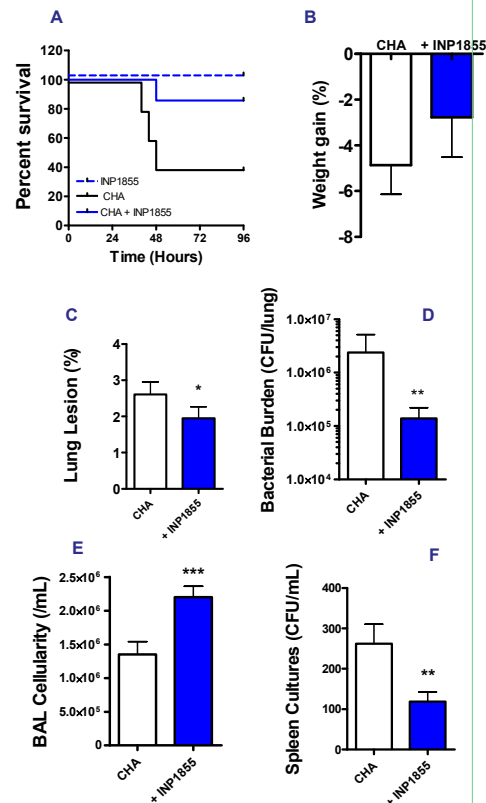
### INP1855 decreases T3SS transcription without affecting QS system



### INP1855 impairs ExoS secretion

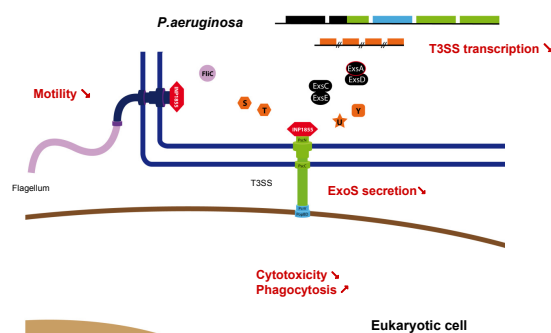


### INP1855 increases mice survival and bacterial clearance



## CONCLUSION

- INP1855 protects epithelial and phagocytic cells from T3SS-induced toxicity and is also effective in vivo.
- It inhibits both secretion of T3SS toxins and flagellar motility, suggesting a molecular target common to these two secretion systems.
- Although further studies are needed to better characterize its mode of action, our data highlights a potential interest for INP1855 as anti-pseudomonal agent.



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- Decreased functionality of T3SS as potential mechanism of action of INP1855:**
- direct or indirect inhibition of the T3SS export apparatus and of flagellar basal body (which show a high degree of similarity) causing
    - a concomitant decrease in motility, toxin secretion and T3SS transcription (ExsE, the repressor of the transcriptional factor ExsA, being translocated via the T3SS)
    - a reduction in toxicity of *P. aeruginosa* for the host cells and an increased phagocytosis (demonstrated in vitro).
    - a reduction in pulmonary lesions and infection dissemination observed in vivo