Influence of Oxidant Stress on the Activity of Antibiotics against the Extracellular and Intracellular (THP-1 cells) Forms of *S. aureus* and its Small Colony Variants (SCV) Mutants

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Abstract

BACKGROUND. Intracellular survival of S. aureus (SA) is associated with persistent infections, especially by SCVs. As reactive oxidant species (ROS) may be critical in antibiotic-dependent bacterial killing, we have examined how activation of monocytes modulate antibiotic (AB) activity against wiid-type phenotype MRSA COL and its menD and hemB SCV mutants.

METHODS. Strains and antibiotics are shown in the Table. MICs: microdiution in MH Bat pH 55 (anticiding page) seconds, with preincubation with H₂O₂ when indicated). Cells: THP-1 monocytes activated with Photod Myristete Acetate (PMA). Intracellular activity: Change in CFU from post-phagocytosis inoculum after 24 h incubation with A8: Na-cetycysteine (NAC) used as a scavemegr of ROS; relative potency and efficacy calculated from the Hill equation of concressonse curve.

RESULTS.

Antibiotic	Strain	Ext	racellular		Intracellular		
		MIC (µg/mL)*	Reduction in MIC upon preincubation with H ₂ O ₂ (fold dilutions) ^b	Cs (X MIC) in THP-1 cells :	Gain in potency upon cell activation by PMA (fold reduction in Cs) ^d	Loss of efficacy upo scavenging oxidant species by NAC (Alog CFU)	
Gentamicin	WT	1	3	0.15	4.4	2.6	
(GEN)	menD	32	8	0.03	1.7	2.2	
	hemB	0.5	2	0.35	17.4	2	
Moxifloxacin	WT	0.125	6	0.50	6.6	2	
(MXF)	menD	0.25	7	0.05	1.2	1.9	
	hemB	0.125	7	0.24	4.7	2.2	
Oritavancin	WT	0.25	0	2.45	0.8	1.3	
(ORI)	menD	0.125	0	0.57	1.0	0.6	
	hemB	0.25	1	0.38	0.2	0.7	

*value measured at pH 5.5 to mimic the phagelysosomal environment (infected compartment of the cells) lerence (fold diution) between the MIC measured at pH 5.5 in control conditions and for bacteria that had been pre-exposed for 30 min to 10 mM H-O- before antibiotic addition

* static concentration (i.e. extracting in a drawn in trianch, and trianch, additional additional interaction and trianch interaction). The static rest in the rest in the static rest in the rest in the static rest in the rest in t

⁴ ratio between the Ca calculated as described above in undifferentiated THP-1 colis and in THP-1 colis that had been preincubated for 48 h with 200 gpL of Photota Mystatia Acatata (PMA) change in Intracellular incolum (xlog CPU) in THP-1 colis incubated for 24 h with antibiotics at their static concentrations in the response of 55 mM Aurahhvehem (MC), convent examined in extension as compared in control remotings (non-trace).

In broth, preincubation with H₂O, markedly decreased the MIC of GEN and MXF but not that of GRI. Call adviation by PMA increased the intracellular potency of GEN and MXF (wild-type strain and *hemB* mutant), with eNAC reduced their activity by 2 goin non-activated cells. In contrast, cell activation had no effect on intracellular ORI potency and NAC reduced their J tog only.

CONCLUSIONS. Activation of THP-1 cells by PMA may increase the intracellular relative potency of AB according to the dependence of their activity on ROS (cooperation with host defenses). Conversely, agents for which cell activation has minimal effect (like ORI), may remain as effective when host defense mechanisms are weakened.

Introduction

Small-Colony Variants (SCVs) of Staphylococcus aureus have a propensity to survive within eukaryotic cells, which has been associated with the persistent and/or recurrent character of the infections they cause.

Reactive oxidant species (ROS) are critical for antibiotics such as fluoroquinolones or aminoglycosides to express their bactericidal activity in broth [1].

We therefore wondered whether ROS produced by monocytes upon activation by PMA [2] could also enhance the activity of bactericidal antibiotics towards intra-phagocytic SCVs.

Aims and set-up

Our aim was to assess the influence of ROS on the extracellular and intracellular activity of 3 bactericidal antibiotics with distinct modes of action (gentamicin, moxifloxatin and oritavancin) against the extracellular and intracellular forms of the MRSA COL strain and its isogenic menadione- and hemin-dependent SCVs.

We used a model of infected THP-1 human monocytes in which a menadione-dependent mutant grows much more slowly than an hemindependent strain but remains as susceptible to antibiotics in terms of maximal reduction of the inoculum compared to the post-phagocytosis values [3].

For extracellular activity, ROS were generated by preexposure to H₂O₂. For intracellular activity, THP-1 cells were activated with phorbol myristyl acetate (PMA) and scavenged by addition of N-acetyl-cysteine (NAC) [4].



strain and its SCVs (supplemented or not in MSB or in hemin)

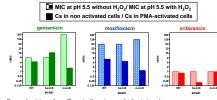
Antibiotic	pН	H ₂ 0 ₂	MIC (mg/L)					
			Wild-type -	menD mutant		hemB mutant		
				- MSB	+ MSB	- hemin	+ hemin	
Gentamicin	7.4	-	0.25	1	1	0.5	0.125	
		+	0.03	0.125	0.125	0.125	0.125	
	5.5	-	1	32	32	32	0.5	
		+	0.125	0.125	0.125	0.125	0.06	
Moxifloxacin	7.4	-	0.03	0.125	0.125	0.125	0.06	
		+	0.008	0.008	0.008	0.002	0.008	
	5.5	-	0.125	0.25	0.5	0.25	0.125	
		+	0.002	0.002	0.002	0.001	0.002	
Oritavancin	7.4	-	0.25	0.03	0.03	0.125	2	
		+	0.25	0.03	0.03	0.125	2	
	5.5	-	0.25	0.125	0.06	0.125	0.25	
		+	0.25	0.125	0.06	0.125	0.25	

MICs were measured in broth at pH 7.4 or 5.5 for bacteria pre-exposed or not to10 mM H₂O₂ for 30 min

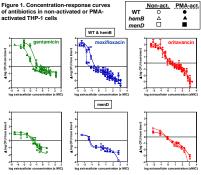
MICs of gentamicin and moxifloxacin were reduced if bacteria were preincubated with H₂O₂.
Those of oritavancin remained unchanged.

Figure 2. Comparison of the change in extracellular and intracellular potency induced by exposure to H₂O₂ (extracellular) or by PMA activation (intracellular)

MICs were measured for bacteria preexposed or not to 10 mM H₂O₂ for 30 min. Static conc. (Cs) were determined by graphical interpolation using the conc-effect curves



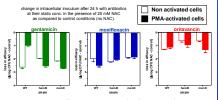
- · For gentamicin and moxifloxacin, there is a marked gain in potency
- extracelluarly, in the presence of H₂O₂ towards all strains
- intracellularly, upon PMA activation, towards the WT strain and the hemB mutant
- For oritavancin, neither H₂O₂ nor PMA activation were able to modify extracellular or intracellular potency, respectively



Infected cells were exposed during 24 h to antibiotics at increasing concentrations. Data are expressed as changes from the post-phagoctosis inoculum, with concentrations expressed in X MIC at pH 5.5 (+ hemin for hem/B strain that grows normally within the cells [1]).

- Gentamicin and moxifloxacin showed higher potency (lower Cs) against the WT and hemB mutant in PMA-activated cells than in non-activated cells.
- · Oritavancin has a similar potency in both cell types irrespective of the strain.

Figure 3. Loss of antibiotic intracellular efficacy (at their static conc.) induced by the addition of NAC in non-activated or PMA-activated cells



 Loss in intracellular efficacy in the presence of NAC was greater for gentamicin and moxifloxacin than for oritavancin in both cell types for all strains.

Methods

- Bacterial strains: strain COL (wild-type, HA-MRSA); its menD and hemB SCV mutants [5,6], supplemented or not with 2 µg/mL menadione sodium bisulfite (MSB) or hemin.
- Extracellular activity: MIC determination according to CLSI recommendations [7] in CA-MHB adjusted at pH 7.4 or 5.5, for bacteria preincubated or not for 30 min with 10 mM H₂O₂ in the dark.
- Cell activation: Incubation with 200 µg/L PMA for 48 h at 37°C.
- Intracellular activity: infection of cells by preopsonized bacteria, antibiotic exposure and assessment of activity determined after 24h incubation following published procedure [3]. Curve fitting of concentration-effects performed using sigmid or biphasic sigmidial regressions. When applicable, coincubation with 25 mM NAC and antibiotics at their static concentration during 24 h.

Conclusions

- For gentamicin or moxifloxacin, activation of cells increases their intracellular potency against normal phenotype and hemin-dependent strains (but not against the menadione-dependent strain because it is intrinsically more susceptible). These drugs may, therefore, depend on host defense mechanisms to fully exert their activity.
- Conversely, cell activation did not modify the intracellular activity of oritavancin, suggesting that this drug is not dependent on oxidant species for activity and that it may remain effective even when oxygen-dependent host defense mechanisms are weakened.

References

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Results