

Interactions of Ciprofloxacin with DPPC and DPPG: Fluorescence Anisotropy, ATR-FTIR and ^{31}P NMR Spectroscopies and Conformational Analysis

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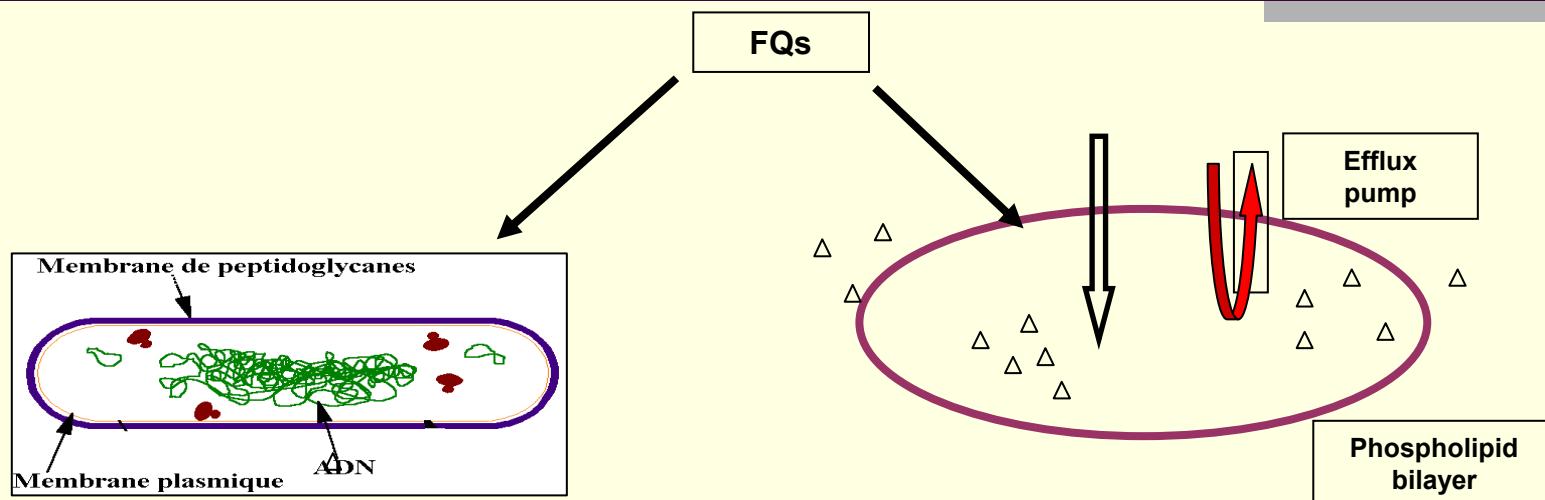
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Introduction

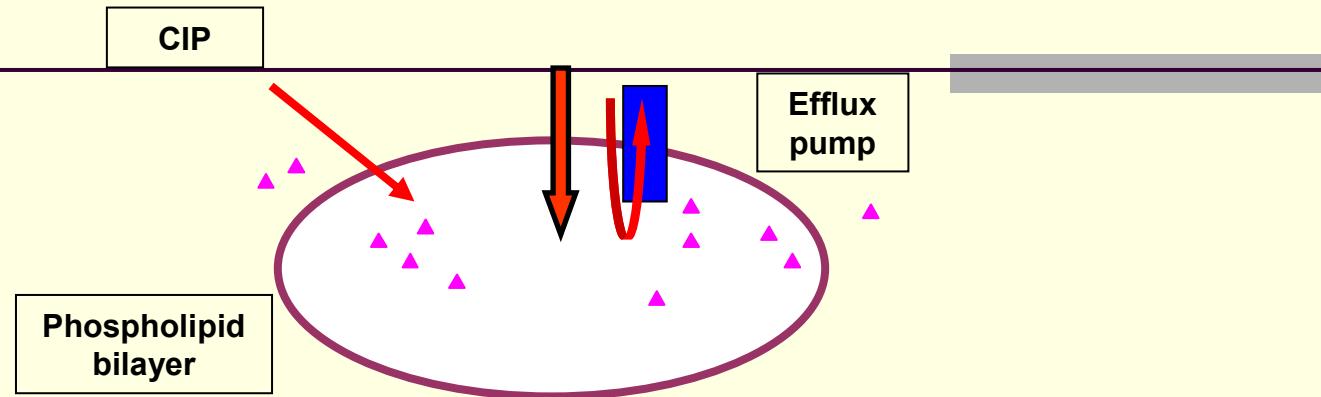
Fluoroquinolones (FQs)- lipids interaction



To kill intracellular bacteria and to reach bacterial target, the FQS :

- interact with cell membrane lipids bilayer : where they can be recognized by the membrane protein efflux pumps
- interact with bacterial membrane

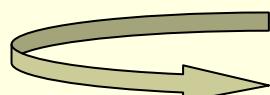
Contribution of efflux pumps and lipids composition to resistance of bacteria to Fluoroquinolones (CIP)



The mechanism of entrance of CIP involved:

- Diffusion process
- Recognition by efflux proteins in cell membranes and /or bacteria membrane

(Michot et al., 2004, Wang et al., 2007, Périchon et al., 2007)



CIP interact with phospholipids
Role of lipids in this phenomena ?

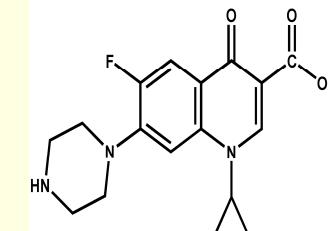
Aim of study

Characterization of the interactions between CIP
with eucaryotic and procaryotic model lipids
membrane (DPPC vs DPPG)

Materials

i) Antibiotics

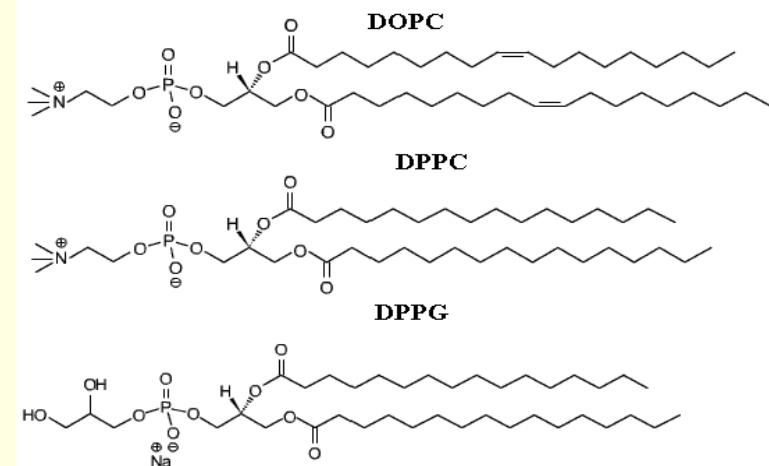
- Ciprofloxacin



ciprofloxacin

ii) Model membranes (Lipids)

- Zwitterionic lipid (DPPC, DOPC)
- Negative charged lipid (DPPG)
- Liposomes: LUVs and MLV



Methods

a) Binding parameters of Ciprofloxacin with lipids (DPPC, DOPC, DPPG)

Steady state anisotropy measurement

b) Effect of Ciprofloxacin on the mobility of phosphate heads of DPPC and DPPG

^{31}P Nuclear Magnetic Resonance (NMR)

c) Ciprofloxacin effect on the melting temperature of DPPC and DPPG

Attenuated Total-Reflection- Fourier Transform Infrared Spectroscopy (ATR-FTIR)

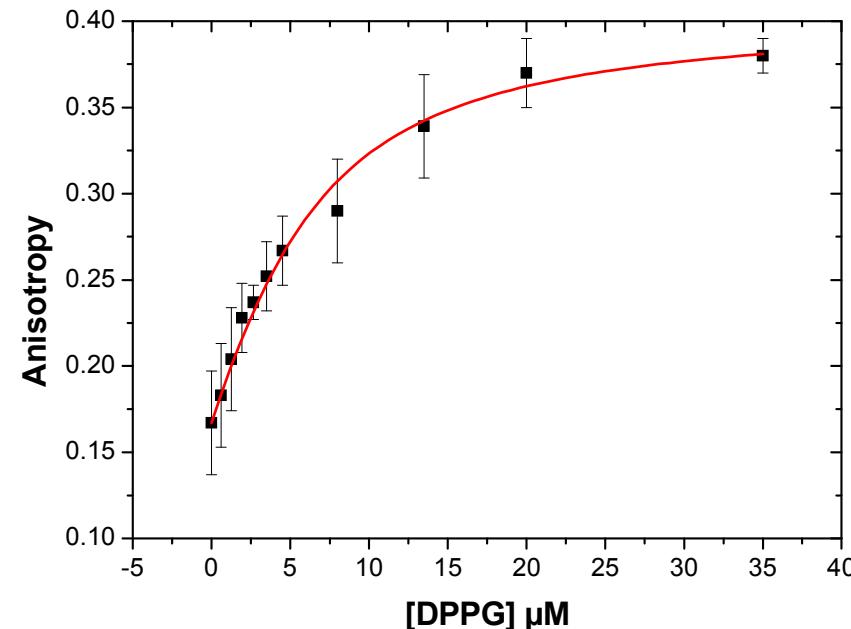
d) Assembly of CIP with phospholipids by molecular modeling

Hypermatrix procedure

Results

a) Binding of CIP to Lipids by steady state anisotropy titration

[CIP] = 5 μM



$$K_{\text{app}} = (8.6 \pm 0.5) \times 10^5 \text{ M}^{-1}$$

a) Binding parameters of CIP with lipid vesicles

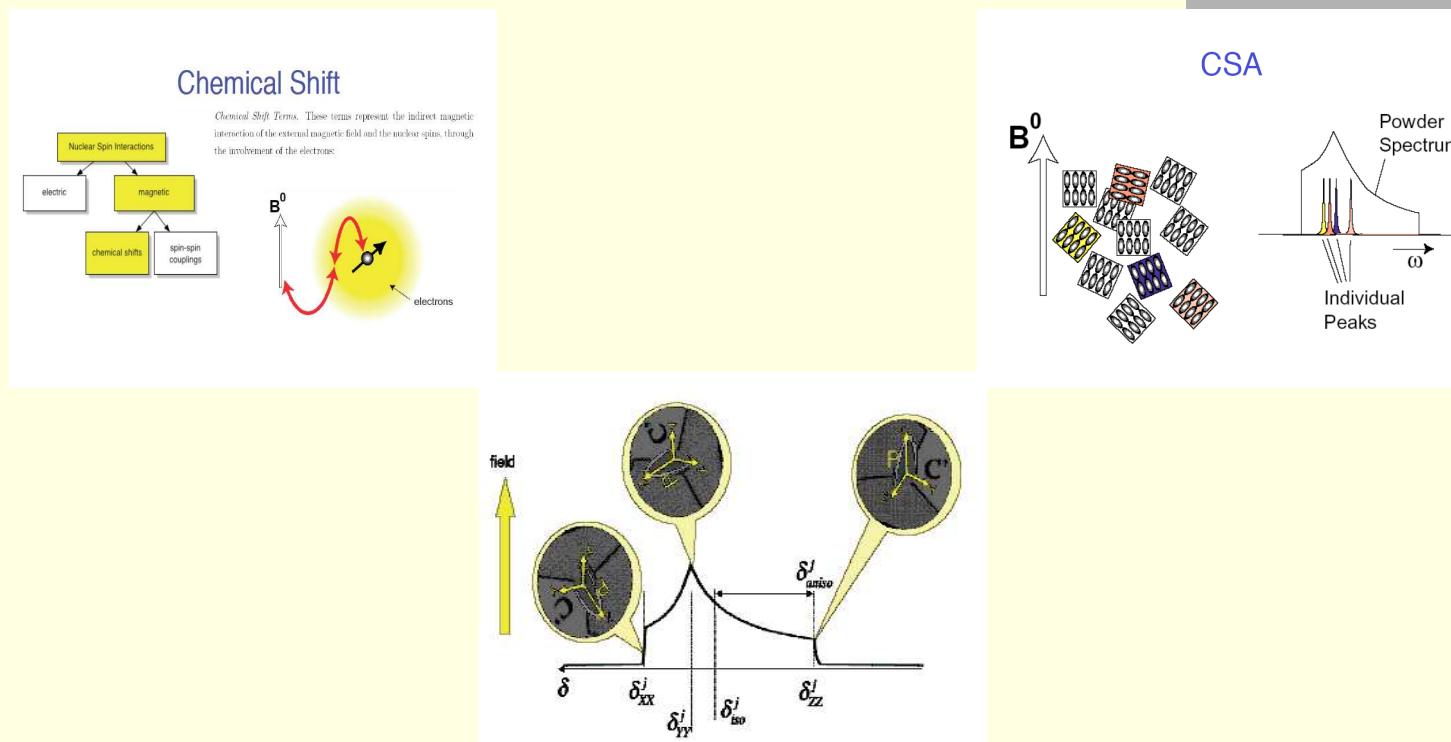
LUV liposomes Composition	$K_{app}(10^5 M^{-1})$
DPPG	8.6±0.5
DPPC	2.5±0.1
DOPC:DPPG (1:1, M:M)	3.2±0.9
DOPC:DPPC (1:1, M:M)	1.1±0.2

⇒ CIP has more affinity for negative charge liposomes
(DPPG Vs DPPC)

b) Mobility of phosphate heads of phospholipids

Nuclear Magnetic Resonance (^{31}P NMR)

Chemical shift anisotropy $\Delta\sigma$

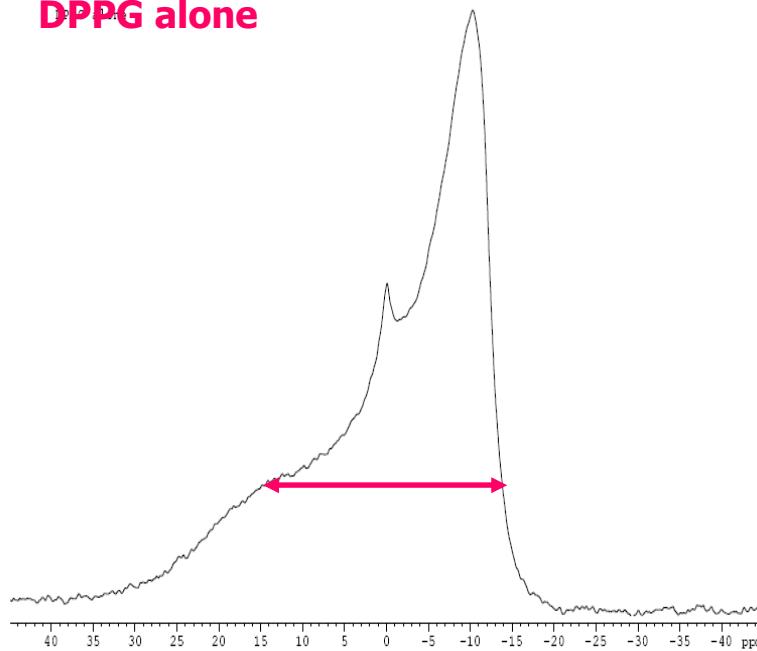


$$\Delta\sigma = \sigma_{\parallel} - \sigma_{\perp}$$

σ_{\parallel} : The low field shoulder
 σ_{\perp} : The high field peak

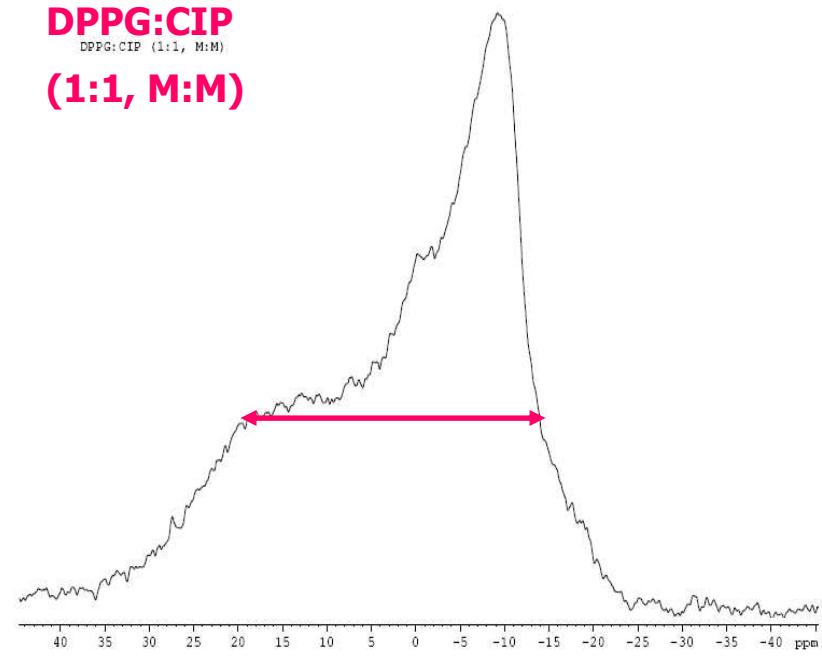
b) CIP effect on the membrane mobility
i) DPPG

DPPG alone



$$\Delta\sigma = 25.5 \pm 0.5 \text{ ppm}$$

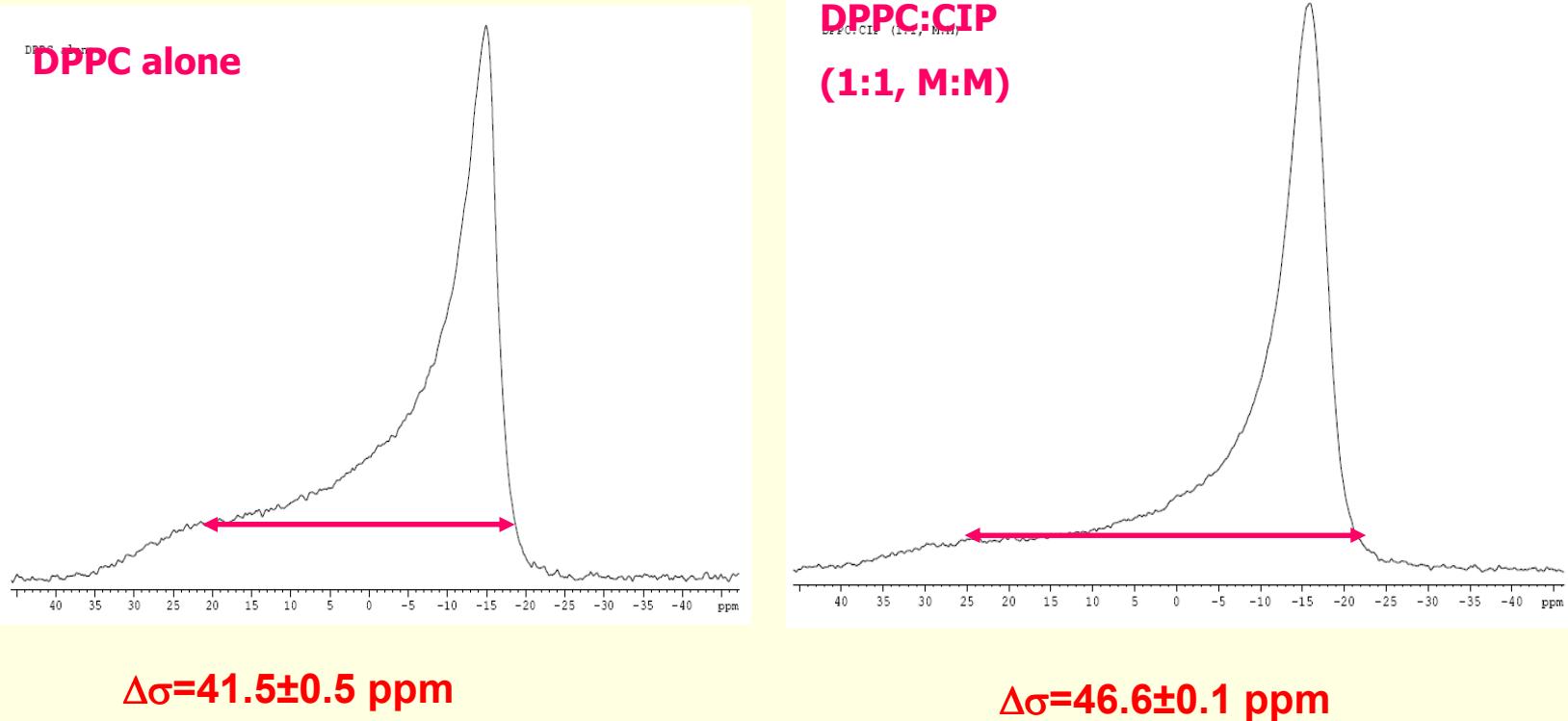
DPPG:CIP
(1:1, M:M)



$$\Delta\sigma = 34.0 \pm 1.0 \text{ ppm}$$

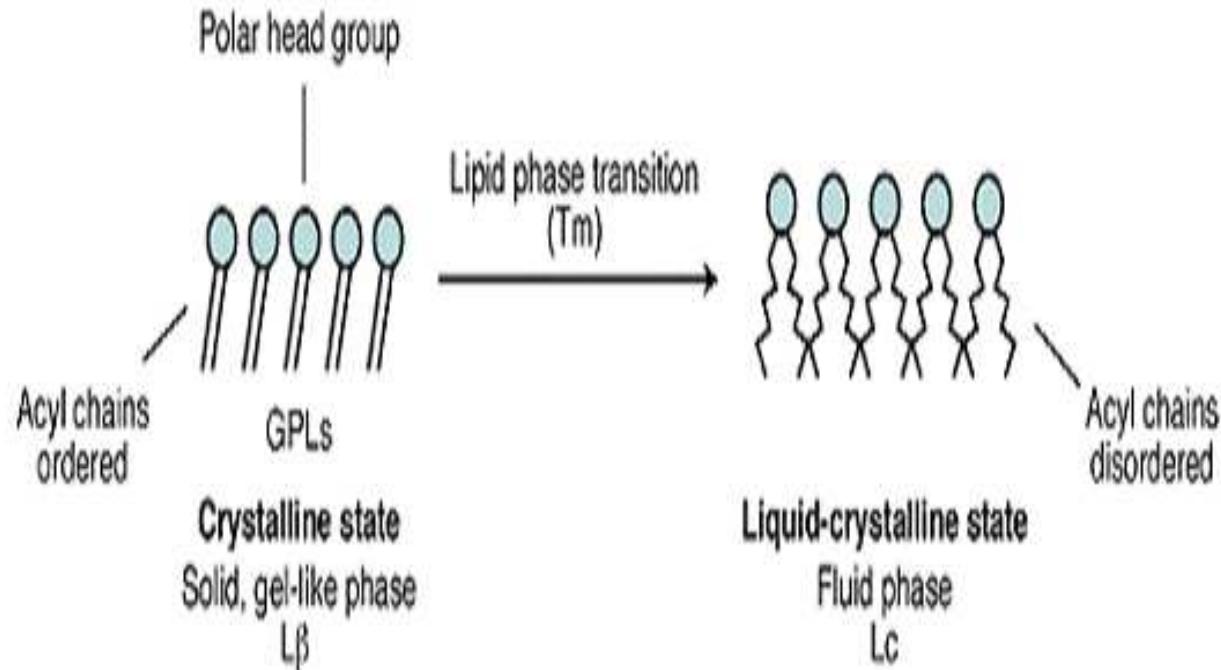
⇒ The difference of $\Delta\sigma$ of DPPG:CIP and DPPG is 9 ppm

b) CIP effect on the membrane mobility
ii) DPPC



⇒ The difference of $\Delta\sigma$ of DPPC:CIP and DPPC is 5 ppm

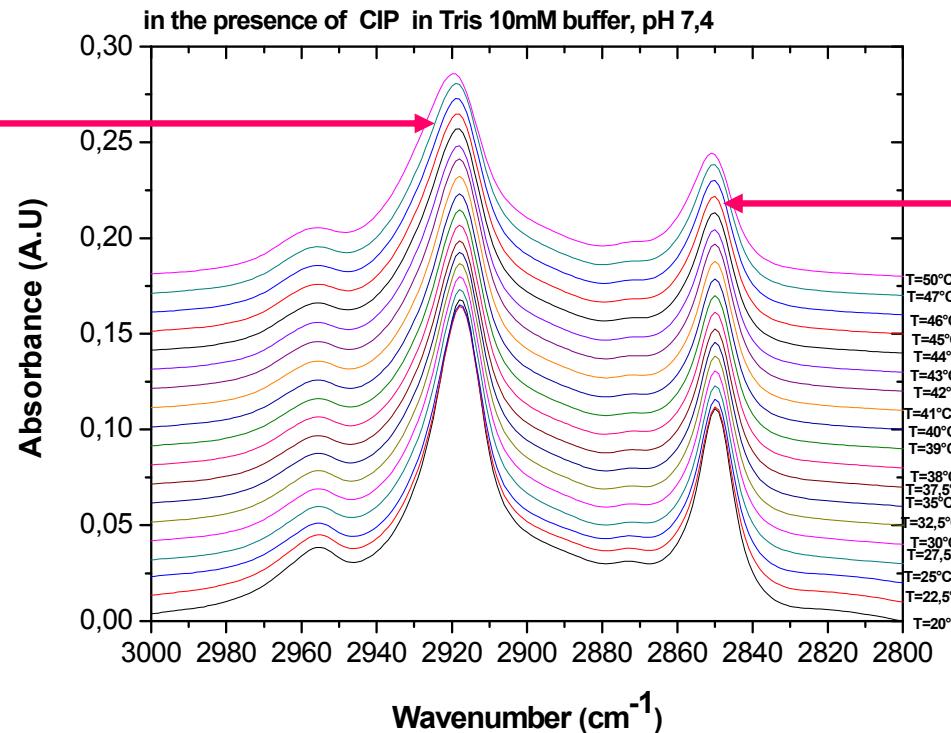
c) CIP effect on the melting temperature of lipid



c) Melting temperature of lipid by ATR-FTIR



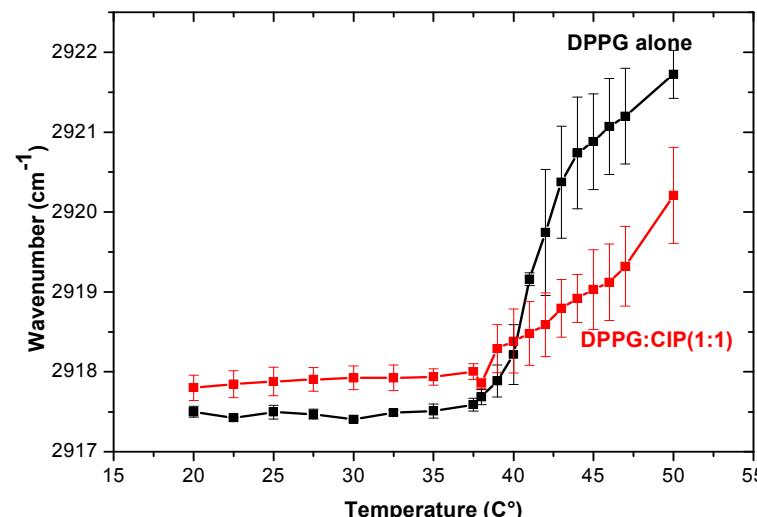
Evolution of stretching vibration bands of CH_2 of DPPG as function of temperature



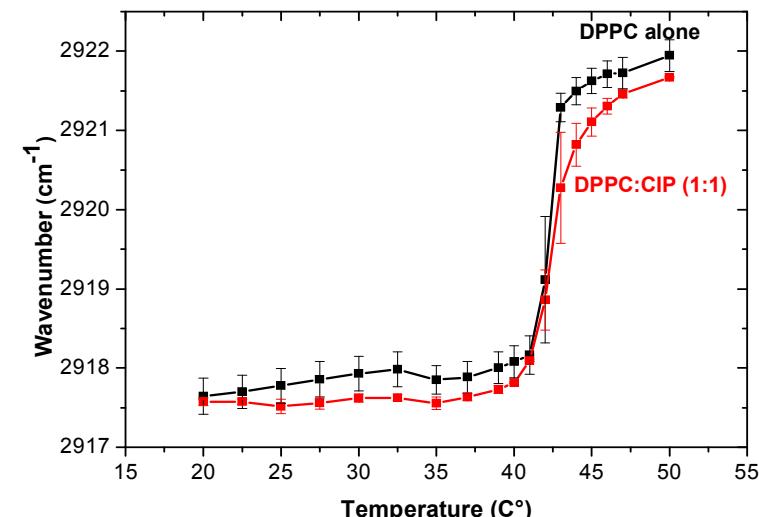
c) CIP effect on the melting temperature of DPPG and DPPC

CH₂ asymmetric stretching band

Melting temperature of DPPG and
DPPG:CIP (1:1)



Melting temperature of DPPC and
DPPC:CIP (1:1)

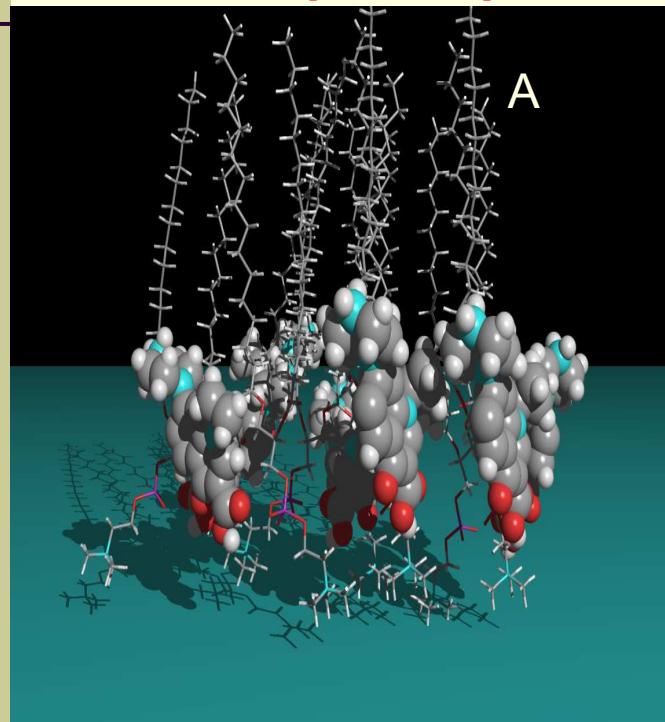


$$\text{Tm(DPPG)} = 40 \text{ } ^{\circ}\text{C}, \text{Tm(DPPC)} = 42 \text{ } ^{\circ}\text{C}$$

- CIP did not affect dramatically the DPPC melting curve
- CIP : ↓ order of acyl chain of DPPG < Tm
↑ order of acyl chain of DPPG > Tm

d) Assembly of CIP with phospholipids by molecular modeling using the Hypermatrix procedure

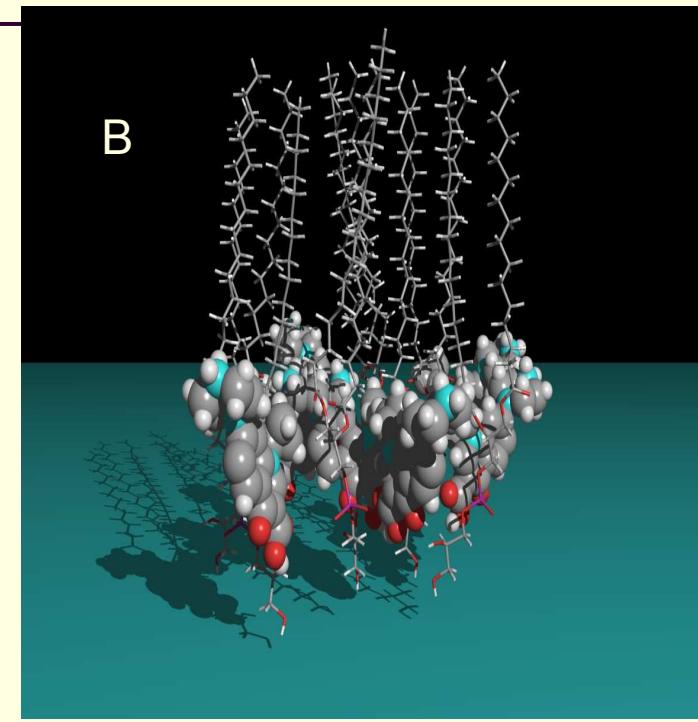
DPPC:CIP (1:1, M:M)



$E = -44.4 \text{ Kcal/mol}$

Area = 66 \AA^2

DPPG:CIP (1:1, M:M)



$E = -53.4 \text{ Kcal/mol}$

Area = 76 \AA^2

=> The interaction of DPPG:CIP is more stable than DPPC:CIP

Conclusion

- The binding constants K_{app} were in the order of $10^5 M^{-1}$ and the affinity appeared dependent on the negative charge of liposomes: DPPG > DOPC: DPPG (1:1) > DPPC > DOPC: DPPC (1:1)
- The major effect of CIP on DPPG as compared to DPPC suggested a role of the electrostatic interactions between CIP and lipids and the importance of the nature of the polar heads of phospholipids

Bensikaddour H. et al., (2008): *Biochim Biophys Acta* **1778**:2535-2543



Acknowledgement



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Thanks for your attention !