Bug-

Drug-Host

Triad in the Era of Antibiotic Resistance: Focus on the Spectrum of ABSSSI

Supported by an educational grant from Melinta Therapeutics



THE BUG: KNOWN AND HIDDEN CHALLENGES OF INFECTIONS WITH STAPHYLOCOCCUS AUREUS

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This presentation has been updated on Nov 3, 2016 (see slides 45-50)



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Disclosures

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- Non-profit Institutions:
 - the Belgian *Fonds de la Recherche Scientifique* for basic research on pharmacology antibiotics and related topics
 - The European Union for applied research on optimization of β-lactams treatments through on-line monitoring of free serum levels
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- Industry:
 - AstraZeneca, GSK, Sanofi-Aventis, Bayer HealthCare, Cempra Pharmaceuticals, The Medicines Company, Northern Antibiotics, RibX, Cubist, Galapagos, ...

Other relationships in relation to this talk

- Belgian Antibiotic Policy Coordination Committee,
- European Committee for Antibiotic Susceptibility Testing (EUCAST)
- European Medicines Agency (EMA)





The problems

- the rise of resistance
 - from penicillin-S to MDR
- hidden habitats
 - intracellular forms
- unexpected forms
 - the small colony variants
- the communities
 - biofilms



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Towards solution(s)

- the antibiotics
 - success of the 10 x '20 initiative ?
- driving antibiotic into cells
 - is accumulation helping ?
- antibiotic combinations
 - overcoming poor susceptibility ?
- improving antibiotic penetration
 - disrupting the matrix ?



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The Staphylococcus *aureus* saga: 60 first years ...

1881: First observation of *staphylococci* in pus by Alexander Ogston



"Micrococci so deleterious when injected are seemingly harmless on the surface of wounds and ulcers". Br Med J 1881;1:369e375 1884: First distinction between *S. aureus* and *S. albus* by Friedrich Rosenbach



1914-1918:

Half of the casualties in the trenches of the First World War were due to septic wound infections with *S. aureus*.



1940-45:

the production process for penicillin (then still universally active against the bacterium*) was a military secret



* the original observation of Fleming (1928) was made on *S. aureus*

The Staphylococcus *aureus* saga: the next 17 years ...

1944: First description of a β-lactamase in *S. aureus* *





Lee, S. (2008). State of C2/C3 substituents of ?-lactam antibiotics in the β -lactam ring cleavage by β -lactamases. *PHILICA.COM Article number* 122.

The first description of a β-lactamase was made in
1940 in *E. coli* (Nature 146, 837 (28 December 1940)

1950-70: almost all strains of S. aureus produce a β-lactamase



Figure. Secular trends of approximate prevalence rates for penicilinase-producing, methicillin-susceptible strains of *Staphylococcus aureus* in hospitals (closed symbols) and the community (open symbols). 1960: introduction of methicillin ... and emergence of resistance to methicillin in 1961 694 SEPT. 3, 1960 BRITISH BRL 1241

MICROBIOLOGICAL STUDIES ON SODIUM 6-(2,6 DIMETHOXYBENZAMIDO) PENICILLANATE MONOHYDRATE (BRL 1241) IN VITRO AND IN PATIENTS

BY

G. T. STEWART, M.D., B.Sc.

With the Technical Assistance of PATRICIA M. HARRISON, B.Sc., and R. J. HOLT, F.I.M.L.T.

From Queen Mary's Hospital for Children and the Medical Research Council Laboratories, Carshalton, Surrey

A report in 1959 by Batchelor *et al.* on the isolation of 6-aminopenicillanic acid drew attention to the possibility of synthesizing new forms of penicillin by the introduction of side-chains. Derivatives prepared in this way may or may not possess antibacterial activity, but we were particularly impressed by the range and mode of action of one derivative, supplied to us in 1959 as BRL 1241 ("celbenin"). The compound—sodium 6-(2,6 dimethoxybenzamido)penicillanate monohydrate may be represented by the following structural formula:



Methicillin-resistant staphylococci

MARY BARBER

From the Department of Bacteriology, Postgraduate Medical School of London

SYNOPSIS Eighteen strains of *Staph. pyogenes* (nine penicillin-sensitive and nine penicillin-destroying) were passaged 40 to 50 times on Celbenin¹ ditch plates.

All strains developed an increase in resistance to Celbenin and eight strains (four penicillinsensitive and four penicillin-destroying) were able to grow in 100 μ g/ml. or more Celbenin. Resistance was of the drug-tolerant type and none of the cultures inactivated Celbenin. There was an associated increase in tolerance to benzyl penicillin.

The highly Celbenin-resistant cultures isolated from penicillin-destroying staphylococci were in sharp contrast to those from penicillin-sensitive strains, as well as to penicillin G-tolerant staphylococci isolated *in vitro*, because they retained the cultural characteristics, coagulase and haemolytic activity, and mouse virulence of the parent strains, and the degree of resistance remained stable after repeated passage in the absence of Celbenin.

Three naturally occurring Celbenin-resistant strains of *Staph. pyogenes* isolated from infective processes were also studied. All three strains grew luxuriantly in concentrations of Celbenin up to $12.5 \,\mu$ g/ml. but very poorly in higher concentrations.

The possible significance of these findings is discussed.

The *Staphylococcus aureus* saga: from 1961 onwards...

1970's: Spreading of methicillin resistance In hospitals 1980's: Large scale re-introduction of vancomycin *



FIG. 1. Usage of vancomycin (in kilograms) in the United States, France, Italy, Germany, United Kingdom, and The Netherlands.

Kirst et al. Antimicrob Agents Chemother. 1998: 42:1303-4.

1997: Strains with reduced susceptibility to vancomycin

Journal of Antimicrobial Chemotherapy (1997) 40, 135–146

Correspondence

Methicillin-resistant *Staphylococcus aureus* clinical strain with reduced vancomycin susceptibility

JAntimicrob Chemother 1997; 40: 135-136

K. Hiramatsu^{*a**}, H. Hanaki^{*a*}, T. Ino^{*b*}, K. Yabuta^{*b*}, T. Oguri^{*c*} and F. C. Tenover^{*d*}

^aDepartment of Bacteriology; ^bDepartment of Pedi atrics, Juntendo University, Tokyo; ^cClinical Labora tory, Juntendo Hospital, Tokyo, Japan; ^dNosocomial Pathogens Laboratory, Centers for Disease Control and Prevention, Atlanta, GA, USA





Staphylococcus aureus and linezolid

1996:

First description of linezolid

J. Med. Chem. 1996, 39, 673-679

Synthesis and Antibacterial Activity of U-100592 and U-100766, Two Oxazolidinone Antibacterial Agents for the Potential Treatment of Multidrug-Resistant Gram-Positive Bacterial Infections

Steven J. Brickner,* Douglas K. Hutchinson, Michael R. Barbachyn, Peter R. Manninen, Debra A. Ulanowicz, Stuart A. Garmon, Kevin C. Grega, Susan K. Hendges, Dana S. Toops, Charles W. Ford, and Gary E. Zurenko *Upjohn Laboratories, The Upjohn Company, Kalamazoo, Michigan 49001*

Received December 22, 1995[®]

2002:



Linezolid Resistance in *Staphylococcus aureus:* Characterization and Stability of Resistant Phenotype

Satish K. Pillai,^{1,3} George Sakoulas,^{1,3} Christine Wennersten,¹ George M. Eliopoulos,^{1,3} Robert C. Moellering, Jr.,^{1,3} Mary Jane Ferraro,^{2,3} and Howard S. Gold^{1,3} ¹Division of Infectious Diseases, Beth Israel Deaconess Medical Center, ²Departments of Pathology and Medicine, Massachusetts General Hospital, and ³Harvard Medical School, Boston, Massachusetts 2007: Resistance in S. aureus by mehylation (*cfr*)



Toh et al. Mol Microbiol. 2007;64:1506-14 PMID: <u>17555436</u>

Pillai et al. J Infect Dis. 2002;186:1603-7. PMID: 12447736.

2016: first report of cfr in a pandemic MRSA clone (ST22) from patients





First Report of *cfr*-Carrying Plasmids in the Pandemic Sequence Type 22 Methicillin-Resistant *Staphylococcus aureus* Staphylococcal Cassette Chromosome *mec* Type IV Clone

Anna C. Shore,^{a,b} Alexandros Lazaris,^a Peter M. Kinnevey,^a Orla M. Brennan,^a Gráinne I. Brennan,^{a,b} Brian O'Connell,^{b,c} Andrea T. Feßler,^d Stefan Schwarz,^d David C. Coleman^a

Microbiology Research Unit, Dublin Dental University Hospital, University of Dublin, Trinity College Dublin, Dublin, Ireland^a; National MRSA Reference Laboratory, St. James's Hospital, Dublin, Ireland^b; Department of Clinical Microbiology, School of Medicine, Trinity College Dublin, St. James's Hospital, Dublin, Ireland^c; Institute of Farm Animal Genetics, Friedrich Loeffler Institut, Neustadt-Mariensee, Germany^d

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Novel anti-MRSA antibiotics acting on resistant isolates *

* not an exhaustive list

already approved

- 2 β-lactams (ceftaroline / ceftobiprole ^a)
- 3 lipoglypopeptides (telavancin, dalbavancin, oritavancin)
- 1 oxazolidinone: tedizolid ^b

In development

- fusidic acid ^c
- radezolid ^d
- plazomycin
- new fluoroquinolones (delafloxacin, nadifloxacin, ...) e
- new topoisomerase type II inhibitors (gepotidacin, ...)
- fatty acid synthesis inhibitors (AFN-1252/Debio 1452, ...) f



a approved in Europe and other countries for pneumonia (CAP/HAP) - In discussion with FDA for ABSSSI and SAB

^b active against *cfr*+ linezolid resistant isolates

^c development for use in the US

 $^{^{\}rm d}$ currently in development for topical applications

^e very low MICs (overcoming current mutation and efflux-mediated resistance mechanisms)

^f very low MICs (typically 0.008 mg/L)

Novel anti-MRSA antibiotics acting on resistant isolates *

* not an exhaustive list

- already approved
 - 2 β-lactams (ceftaroline / ceftobiprole a)

- 3 - 1 agents, the antimicrobial pipeline for MRSA is potentiated with a number of agents under pre-clinical and clinical development. This is a hopeful sign that the IDSA's target might possibly be met by 2020.

Kumar & Chopra. J Antimicrob Chemother. 2013;68:1465-70. PMID: 23429643

- plazomyon
- new fluoroquinolones (delafloxacin, nadifloxacin, ...) e
- new topoisomerase type II inhibitors (gepotidacin, ...)
- fatty acid synthesis inhibitors (AFN-1252/Debio 1452, ...) f

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S. aureus in and released from neutrophils in a mouse osteomyelitis model

S. aureus as an intracellular parasite

S. aureus in THP-1 macrophages



Kalinka *et al.*, Int J Med Microbiol. 2014; 304:1038-49 - PMID: <u>25129555</u>

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A simple view in 1991



FACTORS AFFECTING THE ACTIVITY OF ANTIMICROBIALS AGAINST INTRACELLULAR BACTERIA



④ Metabolisation and inactivation

Figure 1: Pharmacokinetic and pharmacodynamic parameters involved in the activity of antimicrobial drugs against intracellular microorganisms.

Intracellular activity is not directly correlated to accumulation



AMP=ampicillin; AZM=azithromycin; CIP=ciprofloxacin; ETP=ertapenem; GEN=gentamicin; GRN=garenoxacin; LNZ=linezolid; LVX=levofloxacin; MEM=meropenem; MXF=moxifloxacin; NAF=nafcillin; ORI=oritavancin; OXA=oxacillin; PEN V=penicillin V; RIF=rifampicin; TEC=teicoplanin; TEL=telithromycin; VAN=vancomycin

24h pharmacodynamic dose-effect model

1. Cell exposure to a <u>a wide range of extracellular concentrations of the antibiotic</u>



Buyck et al. In vitro Models for the Study of the Intracellular Activity of Antibiotics; In "Bacterial Persistence", Molecular Biology Laboratory Protocols Series, J. Michiels and M. Fauvart, editors, 2016, p 147-157 - DOI: 10.1007/978-1-4939-2854-5



Interpretation of the results of the 24h dose-effect model



2. Analysis of the response

E_{min}: cfu increase (in log₁₀ units) at 24 h from the corresponding initial inoculum as extrapolated for an infinitely low antibiotic concentration

Static concentration (C_{stat}):

extracellular concentration resulting in no apparent bacterial growth (number of cfu identical to the initial inoculum)

E_{max}: cfu decrease (in log₁₀ units) at 24 h from the corresponding initial inoculum <u>as extrapolated from</u> <u>infinitely large antibiotic</u> <u>concentration</u>

<u>Reference:</u> Barcia-Macay M, Seral C, Mingeot-Leclercq MP, Tulkens PM, Van Bambeke F. Pharmacodynamic evaluation of the intracellular activity of antibiotics against *Staphylococcus aureus* in a model of THP-1 macrophages. Antimicrobial Agents and Chemotherapy (2006) 50:841-851 – PMID: <u>16495241</u>



Interpretation of the results of the 24h dose-effect model



2. Analysis of the response



<u>Reference:</u> Barcia-Macay M, Seral C, Mingeot-Leclercq MP, Tulkens PM, Van Bambeke F. Pharmacodynamic evaluation of the intracellular activity of antibiotics against *Staphylococcus aureus* in a model of THP-1 macrophages. Antimicrobial Agents and Chemotherapy (2006) 50:841-851 – PMID: <u>16495241</u>

Most antibiotics show a much lower E_{max} against intracellular bacteria than against bacteria in broth



Van Bambeke & Tulkens, ASM Microbe 2016 – poster SATURDAY 571 – Session 188 http://www.facm.ucl.ac.be/posters/2016/ASM-Microbe-2016/VanBambeke-Tulkens-Poster-SATURDAY-571-ASM-Microbe-2016.pdf

 Δ log₁₀ cfu from time 0 (24 h)

log₁₀ antibiotic concentration (multiples of MIC)

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Small Colony Variants



http://infekt.ch/2006/10/small-colony-variants-von-staphylococcusaureus-schwierig-zu-behandelnde-infektionen/ Klinik für Infektiologie/Spitalhygiene, St Gallen, Switzerland Last visited 09/10/2016

- tiny colonies difficult to see by naked eye and often mistakenly disregarded as contaminants
- usually recovered from protracted, difficult-totreat infections (osteomyelitis, infected prostheses, cystic fibrosis...)²
- slow grow (wait for 72h...)
- deficit in metabolic pathways
 (→ menadione, hemin, or thymine dependence) ³
- selected by antibiotics ... and, consequently, poor susceptibility to most antibiotics ⁴
- equally or more infective than their fastgrowing parents ⁵
- particularly prone to invade eukaryotic cells, persist therein and display resistance 6

- ¹ Proctor et al. Infect Agents Dis (1994) 3:302-312 PMID: <u>7889317</u>
- ² Besier et al. J Clin Microbiol (2007) 45:168-172 -PMID:<u>17108072</u>
- ³ Garcia et al, J Antimicrob Chemother (2013) 68:1455-1464 PMID: 23485724

- ⁵ Jonsson et al. Microb Pathog (2003) 34:73-79 PMID: <u>12623275</u> Bates et al. J Infect Dis (2003) 187:1654-1661 PMID: <u>12721946</u>
- ⁶ Proctor et al. Infect Agents Dis (1994) 3:302-312 PMID: <u>7889317</u> Tuchscherr et al. J Antimicrob Chemother (2016) 71:438-448 PMID: <u>26589581</u>

⁴ Baumert et al. Microb Drug Resist. 8:253-260 2002 PMID <u>12523621</u>

SCVs are a cause of resistance

J Antimicrob Chemother 2016; **71**: 438–448 doi:10.1093/jac/dkv371 Advance Access publication 20 November 2015

Journal of Antimicrobial Chemotherapy

Staphylococcus aureus develops increased resistance to antibiotics by forming dynamic small colony variants during chronic osteomyelitis

L. Tuchscherr^{1*}†, C. A. Kreis²†, V. Hoerr^{1,3}†, L. Flint⁴, M. Hachmeister⁴, J. Geraci¹, S. Bremer-Streck⁵, M. Kiehntopf⁵, E. Medina⁶, M. Kribus⁷, M. Raschke², M. Pletz⁸, G. Peters⁴ and B. Löffler^{1,9}

¹Institute of Medical Microbiology, Jena University Hospital, Jena, Germany; ²Department of Trauma, Hand and Reconstructive Surgery, University Hospital of Münster, Münster, Germany; ³Department for Clinical Radiology, University Hospital of Münster, Münster, Germany; ⁴Institute of Medical Microbiology, University Hospital of Münster, Münster, Germany; ⁵Department of Clinical Chemistry and Laboratory Medicine, Jena University Hospital, Jena, Germany; ⁶Helmholtz Center for Infection Research, Braunschweig, Germany; ⁷Department of Trauma, Hand and Reconstructive Surgery, Jena University Hospital, Jena, Germany; ⁹Center for Sepsis Control and Care (CSCC), Jena University Hospital, Jena, Germany; ⁹Center for Sepsis Control and Care (CSCC), Jena University Hospital, Jena, Germany; ⁹Center for Sepsis Control and Care (CSCC), Jena University Hospital, Jena, Germany; ⁹Center for Sepsis Control and Care (CSCC), Jena University Hospital, Jena, Germany; ⁹Center for Sepsis Control and Care (CSCC), Jena University Hospital, Jena, Germany; ⁹Center for Sepsis Control and Care (CSCC), Jena University Hospital, Jena, Germany; ⁹Center for Sepsis Control and Care (CSCC), Jena University Hospital, Jena, Germany; ⁹Center for Sepsis Control and Care (CSCC), Jena University Hospital, Jena, Germany; ⁹Center for Sepsis Control and Care (CSCC), Jena University Hospital, Jena, Germany; ⁹Center for Sepsis Control and Care (CSCC), Jena University Hospital, Jena, Germany; ⁹Center for Sepsis Control and Care (CSCC), Jena University Hospital, Jena, Germany; ⁹Center for Sepsis Control and Care (CSCC), Jena University Hospital, Jena, Germany; ⁹Center for Sepsis Control and Care (CSCC), Jena University Hospital, Jena, Germany; ⁹Center for Sepsis Control and Care (CSCC), Jena University Hospital, Jena, Germany; ⁹Center for Sepsis Control and Care (CSCC), Jena University Hospital, Jena, Germany; ⁹Center for Sepsis Control and Care (CSCC), Jena University Hospital, Jena, Germany; ⁹Center for Sepsis

Tuchscherr et al. J Antimicrob Chemother (2016) 71:438-448 - PMID: 26589581





SCVs are not easily eradicated from osteoblasts in vitro (7 days model)



Osteoblasts infected with S. aureus SCVS and exposed to antibiotics: surviving bacteria



SCVs are not easily eradicated from osteoblasts in vivo (chronic infection)



Magnetic resonance imaging showing osteomyelitis progression with and without therapeutic treatment

- Inflammatory lesions: orange and yellow (left) and red and brown (right)
- Non-inflamed area: magenta (left) and green (right)

Tuchscherr et al. J Antimicrob Chemother (2016) 71:438-448 - PMID: 26589581

A direct comparison of antibiotic efficacies between intracellular SCV and their isogenic fast growing parents (NP)



Efficacy lower against SCV, possibly related to slower metabolism ?

Nguyen et al. Antimicrob Agents Chemother (2009) 53:1434-1442 – PMID: <u>19188393</u>

Dose-effect studies comparing extracellular and intracellular SCVs

Vol. 53, 2009

ANTIBIOTICS AND INTRACELLULAR SCV 1439



FIG. 6. Dose-response curves of the three most active antibiotics against extracellular and intracellular SCVs. The graphs show the change in the number of CFU (Δ log CFU from the initial inoculum) per ml of broth (extracellular [extra]) or per mg of cell protein (intracellular [intra]) in THP-1 macrophages after a 24-h incubation at the extracellular concentrations (total drug) shown in the abscissa. Curves were constructed by sigmoidal regression using the Hill equation (Table 2 gives the regression parameters); for oritavancin, for which a bimodal curve is clearly seen intracellularly, two successive sigmoidal regressions with variable slopes were used to fit the data. The zone highlighted in gray corresponds to the clinically relevant concentration range (total drug). Data are means \pm standard deviations (n = 3); most error bars are smaller than the symbols.





Dose-effect studies comparing extracellular and intracellular SCVs



FIG. 6. Dose-response curves of the three most active antibiotics against extracellular and intracellular SCVs. The graphs show the change in the number of CFU (Δ log CFU from the initial inoculum) per ml of broth (extracellular [extra]) or per mg of cell protein (intracellular [intra]) in THP-1 macrophages after a 24-h incubation at the extracellular concentrations (total drug) shown in the abscissa. Curves were constructed by sigmoidal regression using the Hill equation (Table 2 gives the regression parameters); for oritavancin, for which a bimodal curve is clearly seen intracellularly, two successive sigmoidal regressions with variable slopes were used to fit the data. The zone highlighted in gray corresponds to the clinically relevant concentration range (total drug). Data are means \pm standard deviations (n = 3); most error bars are smaller than the symbols.

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Antibiotic combinations may be a solution



Let us vary the ratio of A to B



Antibiotic combinations may be a solution



rifampin / oritavancin (H) Fractional maximal effect (FME) RIF / ORI 0.1/0.9 0.3/0.7 0.5/0.5 0.7/0.3 0.9/0.1 0.002 0.042 0.161 -D-RIF 0.007 0.018 mg/L 171.72 44.52 19.08 8.17 -0- 0RI 2.12

Here is what you expect !

Li et al. Antimicrob. Agents Chemother (1993) 37:523–531 - PMID: <u>8460921</u> Desbiolles et al. Agents Chemother (2001) 45:3328–3333 - PMID:<u>11709304</u> and this is what we saw !

Nguyen et al. Antimicrob Agents Chemother (2009) 53:1434-

1442 - PMID: 19188393



Antibiotic combinations may be a solution



https://de.wikipedia.org/wiki/Heinrich_von_Opel Last visited: 22 Oct 2016

More combinations must be studied ...

and this is what we saw !

0.018

19.08

0.007

44.52

0.042

8.17



0.161 - C- RIF

-0- 0RI

2.12

Nguyen et al. Antimicrob Agents Chemother (2009) 53:1434-1442 – PMID: <u>19188393</u>

0.002

171.72

mg/L

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Biofilms (microbial communities)







- ancient multicellular life forms ¹ with key pathogenic roles in up to 80 % of infections processes (first recognized with *P. aeruginosa*²)
- structured aggregation of surface-attached microcolonies encased in an extracellular matrix and separated by fluid-filled channels ³
- substratum conditioned by host matrix proteins (fibrinogen, fibronectin, and collagen)⁴
- resistance mediated through a dormant phenotype (anoxic environment and nutrient deprivation) with cell division occurs very slowly, producing persister cells ⁵

1 Henrici AT J Bacteriol (1933) 25:277–87 - PMID: <u>16559616</u> 2 Lam et al. Infect Immun (1980) 28:546–56 -PMID: <u>6772562</u> 3 O'Toole et al. Annu Rev Microbiol (2000) 54:49-79. PMID: <u>11018124</u> 4 François et al. J Lab Clin Med (2000) 135:32–42 -PMID: <u>10638692</u> 5 Lewis K Annu Rev Microbiol (2010) 64:357-372 - PMID: <u>20528688</u>

Biofilms: the problems



pharmacokinetics

- diffusibility through the matrix
- bioavailability within the biofilm
- access to bacteria
- efflux out of bacteria

pharmacodynamics

- bacterial responsiveness (metabolic activity of bacteria)
- antibiotic expression of activity (local environment [O₂, pH, ..])



From F. Van Bambeke: Anti-staphylococcal activity of antibiotics in biofilm and host cell. ESCMID Postgraduate educational course, Lyon, France, 2016 <u>http://www.facm.ucl.ac.be/conferences/2016/06-Lyon-ESCMID/Vanbambeke-ESCMID-intra-biofilm.pdf</u> Last visited: 10 Oct 2016



Biofilms respond poorly to many antibiotics... but not all...



Activities of antibiotics against biofilms. Concentration-response activities of antibiotics against 24-h biofilms of strain 2011S027 incubated with increasing concentrations of antibiotics for 48 h (**DFX, delafloxacin**; **DAP, daptomycin**; **VAN, vancomycin**).

Siala et al. Antimicrob Agents Chemother (2014) 58:6385-6398 – PMID: 25114142

Delafloxacin activity in biofilms is boosted by acid pH

Biofilm pH



Influence of pH on delafloxacin MIC



Micro-pH in biofilms and influence on delafloxacin activity

- Top: micro-pH within biofilms using C-SNARF-4 as a pH-sensitive probe (fluorescence emitted shifting from red to green upon acidification).
- Middle: micro-pH in the depth of the biofilm.
- Lower: influence of pH on the MIC of delafloxacin in MHB. The gray squares highlight the range of pH measured in the corresponding biofilm.



Moving to a dynamic model...







Combinations may be the way to go for biofilms...



Siala et al. Antimicrobial resistance in microbial biofilms and options for treatment ESCMID Course, 5-7 Oct 2016, Ghent, Belgium



Combinations may be the way to go for biofilms...



Stewart et al. PLoS One. 2012;7(11):e50560 - PMID: 23185637

Siala et al. Antimicrobial resistance in microbial biofilms and options for treatment ESCMID Course, 5-7 Oct 2016, Ghent, Belgium

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Fig. 2 Strategies in the management of biofilms (adapted from wikipedia.org/wiki/File:Biofilm.jpg).









ARTICLE

Received 23 Feb 2016 | Accepted 20 Sep 2016 | Published 3 Nov 2016

DOI: 10.1038/ncomms13286

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The antifungal caspofungin increases fluoroquinolone activity against *Staphylococcus aureus* biofilms by inhibiting *N*-acetylglucosamine transferase

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Activity (dose response) of delfloxacin alone or combined with caspofungin on S. aureus biofilms in vivo: (mouse subcutaneous biofilm model). Animals were treated for 7 days with caspofungin (CAS; 4 mg/kg of body weight) once daily, delafloxacin twice daily, or with delafloxacin at each of these doses combined with caspofungin Statistical analysis (one-way ANOVA; Tukey post-hoc test): groups with different letters are significantly different from one another (P<0.05).







Activity of delafoxacine alone or combined with caspofungin against the bioluminescent strain Xen36 in vivo. Bioluminescent signal emitted from catheters infected by Xen36, implanted at day 0 in the back of mice treated 24 h after implantation and for the next 7 days with caspofungin (CAS) (4mg/kg/of body weight once daily), or delafloxacin (DFX) (40 mg/kg of body weight twice daily) or with a delafloxacin and caspofungin (each injected separately and according to its own schedule; Intensity of the transcutaneous photon emission represented as a pseudocolor image.



Disrupting the Matrix ? Here is why ...





Ica A (N-acetylglucosamine transferase) activity and its inhibition by caspofungin. (a) Reaction catalyzed by IcaA. (b) Activity of IcaA in membrane protein extracts from strain ATCC33591 and its *∆icaA* mutant. Activity was evaluated by the amount of UDP liberated in the reaction medium.

Messages

- S. aureus (and other staphylococci) have been of medical concern for many years...
- Resistance was long considered as <u>the</u> first danger ... but may have been put under (provisional) control by the wave of new drugs (success of the 10 x 20's initiative ?)
- Intracellular forms, SCV's, and biofilms still require much attention as current therapies are often inadequate...
 - Novel drugs need to be tested for activity against these recalcitrant forms of infection
 - Fluoroquinolones (delafloxacin in particular) and caspofungin may cooperate for biofilm elimination (through bacterial killing and matrix disruption)