

# Susceptibility trends in pneumonia pathogens and current prescribing.



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Brussels, Belgium



24th **ECCMID** Barcelona, Spain  
10 – 13 May 2014

 **ESCMID** EUROPEAN SOCIETY OF CLINICAL  
MICROBIOLOGY AND INFECTIOUS DISEASES



**Session: Optimising diagnosis and appropriate antibiotic prescribing in pneumonia**



*With approval of the Belgian Common Ethical Healthplatform – visa no. V1/14/04/30/060865*

# Disclosures

Financial support from

- the Belgian *Fonds de la Recherche Scientifique* for basic research on pharmacology antibiotics and related topics
- *Université catholique de Louvain* for past personal support
- Commercial Relationships:
  - AstraZeneca, GSK, Sanofi-Aventis, Bayer HealthCare, Cempra Pharmaceuticals, The Medicines Company, Northern Antibiotics, RibX, Cubist, Galapagos, ...
- Other relationships in relation to this talk
  - Belgian Antibiotic Policy Coordination Committee,
  - European Medicines Agency (as expert for the agency and for Industry)

**Slides: <http://www.facm.ucl.ac.be> → Lectures**

# Do we have a problem ?

Obituary

**J.-M. Ghuysen**



This man discovered the mode of action of penicillin

*Ann. Rev. Biochem. 1979. 48:73-101  
Copyright © 1979 by Annual Reviews Inc. All rights reserved*

## USE OF MODEL ENZYMES IN THE DETERMINATION OF THE MODE OF ACTION OF PENICILLINS AND $\Delta^3$ -CEPHALOSPORINS<sup>1</sup>

*Jean-Marie Ghuysen, Jean-Marie Frère, Mélina Leyh-Bouille,  
Jacques Coyette, Jean Dusart, and Martine Nguyen-Distèche*

Service de Microbiologie, Faculté de Médecine, Institut de Botanique,  
Université de Liège, 4000 Sart Tilman, Liège, Belgium

**and died from invasive pneumococcal infection ...**

<http://www.cip.ulg.ac.be/newsite/pdf/jmghuysen.pdf>

# Which problem ?

- **Community-acquired pneumonia (CAP):**
  - remains a major acute cause of death (6<sup>th</sup> in patients > 65 y);
  - *Streptococcus pneumoniae* is the most commonly identified pathogen, but other bacteria may be critical in specific environments (the causative organisms remains, however, unidentified in 30% to 50% of cases)
  - Resistance to "older" antibiotics is growing ...
- **Hospital/Health Care-acquired/Ventilator associated pneumonia (HCAP/HAP/VAP)**
  - 2nd most frequent acquired infection in the hospital
  - carries a still higher mortality burden (13-55 %)
  - can be caused by a larger variety of organisms highly influenced by prior exposure to antibiotics, type of patient and comorbidities
  - Enteric Gram (-), *S. aureus*, and *P. aeruginosa* are predominant with resistance increasing if late onset (hospital strains)

Infectious Diseases (Cohen, Opal & Powderly, eds), 3d edition, Elsevier 2010,  
• Niederman M.: Community-acquired pneumonia (chapter 27)  
• Papazian L & Donati SY: Hospital-acquired pneumonia (chapter 28)  
available on line at <http://www.expertconsultbook.com> (last access: 18-03-2014)

# BUT...

- **Community-acquired pneumonia (CAP):**
  - remains a major acute cause of death (6<sup>th</sup> in patients > 65 y);
  - *Streptococcus pneumoniae* is the most commonly identified pathogen, but other bacteria, viruses, fungi, and parasites (e.g. *Pneumocystis carinii*) are also causative
  - Resistance to "older" antibiotics is growing ...
- **Hospital/Health Care-acquired Ventilator associated pneumonia (HCAP/HAP/VAP):**
  - 2<sup>nd</sup> most frequent acquired infection in the hospital
  - carries a still higher mortality burden (13-55 %)
  - can be caused by a larger variety of organisms highly influenced by prior exposure to antibiotics, type of patient and comorbidities
  - Enteric Gram (-), *S. aureus*, and *P. aeruginosa* are predominant with resistance increasing if late onset (hospital strains)

Those categorical differences  
are now blurring...

# A quick survey of the main (common) bacterial causative organisms

CAP and HCAP	
Outpatient, no significant comorbidity	<b><i>Streptococcus pneumoniae</i></b> <i>Mycoplasma pneumoniae</i> , <i>Chlamydomphila pneumoniae</i> , <i>Haemophilus influenzae</i> , <i>Legionella</i> spp., <i>Mycobacterium tuberculosis</i> , endemic fungi)
Outpatient, comorbidities or HCAP with no resistance risk factors	<b>Drug resistant <i>Streptococcus pneumoniae</i> (DRSP)</b> Enteric Gram-negative; anaerobes (with aspiration)
Inpatient, with comorbidities or HCAP with no resistance risk factors	<b><i>Streptococcus pneumoniae</i> (including DRSP),</b> <i>Haemophilus influenzae</i> , <i>Mycoplasma pneumoniae</i> , <i>C. pneumoniae</i> , <i>Legionella</i> spp. Enteric Gram-negatives, anaerobes, others...
Severe CAP, with no risks for <i>Pseudomonas aeruginosa</i>	<b><i>Streptococcus pneumoniae</i> (including DRSP),</b> <i>Haemophilus influenzae</i> , <i>Mycoplasma pneumoniae</i> , <i>Legionella</i> spp., <b><i>Staphylococcus aureus</i></b> Gram-negative bacilli, others
Severe CAP, with risks for <i>P. aeruginosa</i> , or HCAP with resistance risk factors	All of the above pathogens, plus <b><i>P. aeruginosa</i></b>

Infectious Diseases (Cohen, Opal & Powderly, eds), 3d edition, Elsevier 2010,  
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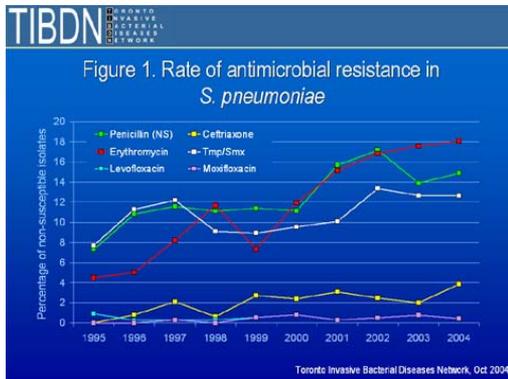
# A quick survey of the main (common) bacterial causative organisms

HAP	
Early pneumonia	<b><i>Streptococcus pneumoniae</i></b> <b><i>Haemophilus influenzae</i></b> , <b>Methicillin-sensitive <i>Staphylococcus aureus</i> (MSSA)</b> Escherichia coli and non-resistant EGNB,
Late pneumonia	<b><i>Pseudomonas aeruginosa</i></b> <i>Acinetobacter</i> spp., <b>Antibiotic-resistant <i>Enterobacteriaceae</i></b> , <b>Methicillin-resistant <i>Staphylococcus aureus</i> (MRSA)</b>
Other situations	Coagulase-negative <i>staphylococci</i>  <i>Neisseria</i> spp., <i>Moraxella</i> spp. <i>Enterobacter</i> spp., <i>Proteus</i> spp. <i>Burkholderia cepacia</i> <i>Acinetobacter</i> spp. <i>Stenotrophomonas maltophilia</i>  Anaerobes ( <i>Peptostreptococcus</i> , <i>Veillonelia</i> , <i>Bacteroides</i> spp. <i>Fusobacterium</i> spp., <i>Prevotella</i> spp., <i>Actinomyces</i> spp.  Intracellular ( <i>Legionella</i> spp. <i>Chlamydia pneumoniae</i> , <i>Mycoplasma pneumoniae</i> )

Infectious Diseases (Cohen, Opal & Powderly, eds), 3d edition, Elsevier 2010,  
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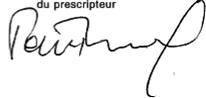
# What is my goal ?

- Discuss with you the trends of resistance of some of these organisms
- how it may impact on you prescription habits...
- leaving to the next speakers the discussion of guidelines ...



<http://microbiology.mtsinai.on.ca/tibdn/data/arspneumo.asp>  
Last accessed: 19 March 2014



 1:26519.66.860		Cachet du pharmacien au verso Exécuté le : <b>TULKENS PAUL</b>
A REMPLIR PAR LE PRESCRIPTEUR: nom et prénom du patient: ..... Titulaire - Conjoint - Enfant - Ascendant (Souligner la mention adéquate)		
Réservé à la vignette du conditionnement	R/ <i>Three weeks                  best in!                  (Refill twice!)</i>	
Cachet du prescripteur <b>Dr TULKENS Paul</b> Biologie Clinique Ctr. de Soignes, 5 B-1404 NIVELEES OM 0519 B1 - tel: 0577/21 80 95	Date et signature du prescripteur 	



**GUIDELINES**



# *Streptococcus pneumoniae*

REVIEW ARTICLE

Drugs 2007; 67 (16): 2355-2382  
0012-6667/07/0016-2355/\$49.95/0

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## Multidrug-Resistant *Streptococcus pneumoniae* Infections Current and Future Therapeutic Options

Françoise Van Bambeke,<sup>1</sup> René R. Reinert,<sup>2</sup> Peter C. Appelbaum,<sup>3</sup> Paul M. Tulkens<sup>1</sup>  
and Willy E. Peetermans<sup>4</sup>

- 1 Unité de Pharmacologie Cellulaire et Moléculaire, Université Catholique de Louvain, Brussels, Belgium
- 2 Institute for Medical Microbiology, National Reference Center for Streptococci, University Hospital (RWTH), Aachen, Germany
- 3 Department of Pathology, Hershey Medical Center, Hershey, Pennsylvania, USA
- 4 Department of Internal Medicine-Infectious Diseases, Katholieke Universiteit Leuven, University Hospital Gasthuisberg, Leuven, Belgium



Colonies of  
*S. pneumoniae*  
CDC Public Health  
Image Library  
<http://phil.cdc.gov/phil>

# Streptococcus pneumoniae: main mechanisms of resistance

Antibiotic class	Mechanism	Genetic support	Drugs affected	Consequence
<b>β-lactams</b>	↘ Affinity of PNP1a, PBP2x and PBP2b	mosaic genes	all (variable extent)	↘ susceptibility
<b>Macrolides</b>	Methylation of 23S rRNA	<i>erm(B)</i>	all except ketolides unless multiple mutations	full resistance
	active efflux	<i>mef(A)</i>	14- and 15-membered ring	moderate (?) resistance
<b>Fluoroquinolones</b>	↘ affinity to DNA-gyrase/topoisomerase complex	point mutations	all (variable extent)	full resistance if several mutations
	active efflux	<i>(pmrA)</i> <i>patA-patB</i>	gatifloxacin, gemifloxacin <sup>1</sup>	↘ susceptibility
<b>Tetracyclines</b>	ribosomal protection	<i>tet(A), tet(O)</i>	all except glycylicyclines	Full resistance
<b>Sulfonamides</b>	↘ of inhibition of dihydropteroate synthase	repetition of codons for aminoacids	all	Full resistance

<sup>1</sup> also norfloxacin and ciprofloxacin (not recommended)

Adapted from Van Bambeke, *et al.* Drugs. 2007;67:2355-82

See also Lismond, *et al.* JAC. 2011;66:948-51, Lismond, *et al.* Intern J Antimicrob Ag. 2012;39:208– 10

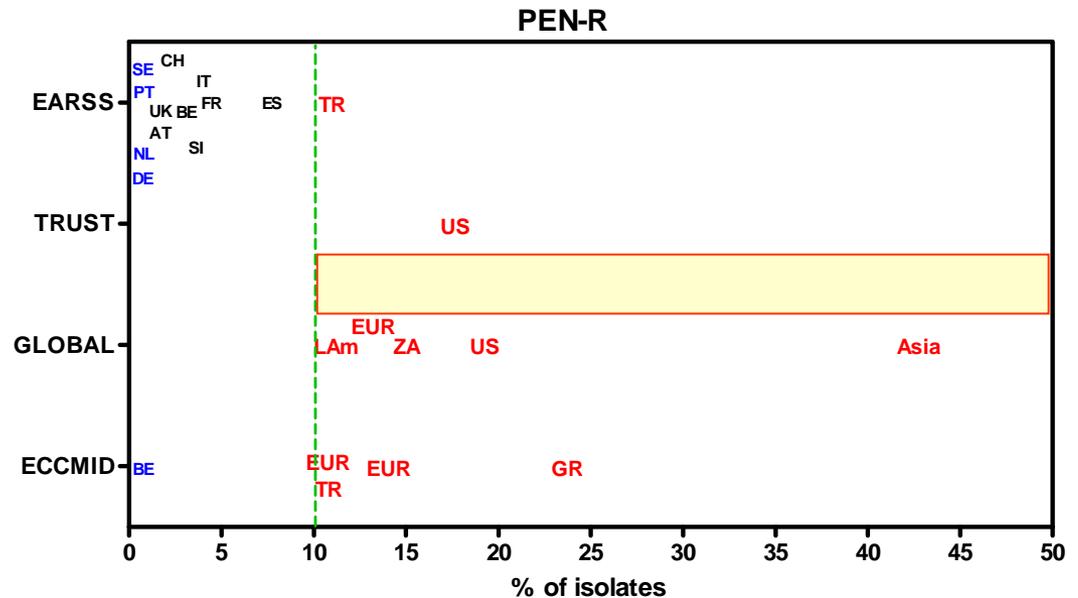
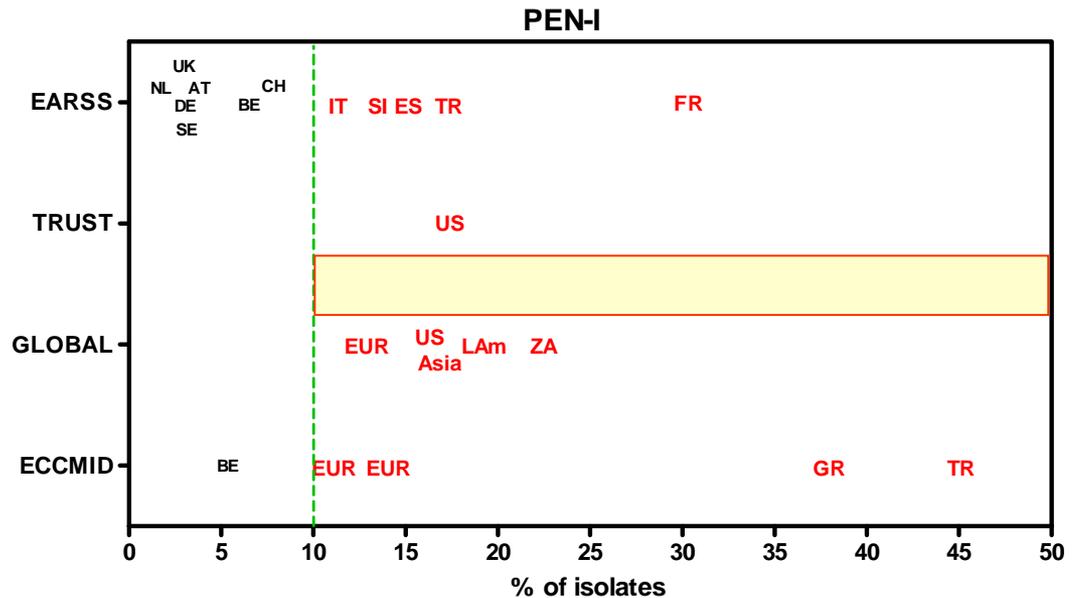
# Resistance of *S. pneumoniae* to penicillins \*

\*Analysis of resistance to penicillins (with CAP as main indication) in surveillance systems or publications (*S. pneumoniae*)

- **EARSS**: European Antimicrobial Surveillance system
- **TRUST**: Tracking Resistance in the United States Today
- **GLOBAL**: Global Landscape On the Bactericidal Activity of Levofloxacin
- **ECCMID**: abstracts of the 18-20th European Congress of Clinical Microbiology and Infectious Diseases

**Most studies used CLSI (non-meningitis) breakpoints**

CAP: community acquired pneumonia  
 CLSI: Clinical and Laboratory Standards Institute (<http://clsi.org>)



Lismond *et al.*, in preparation

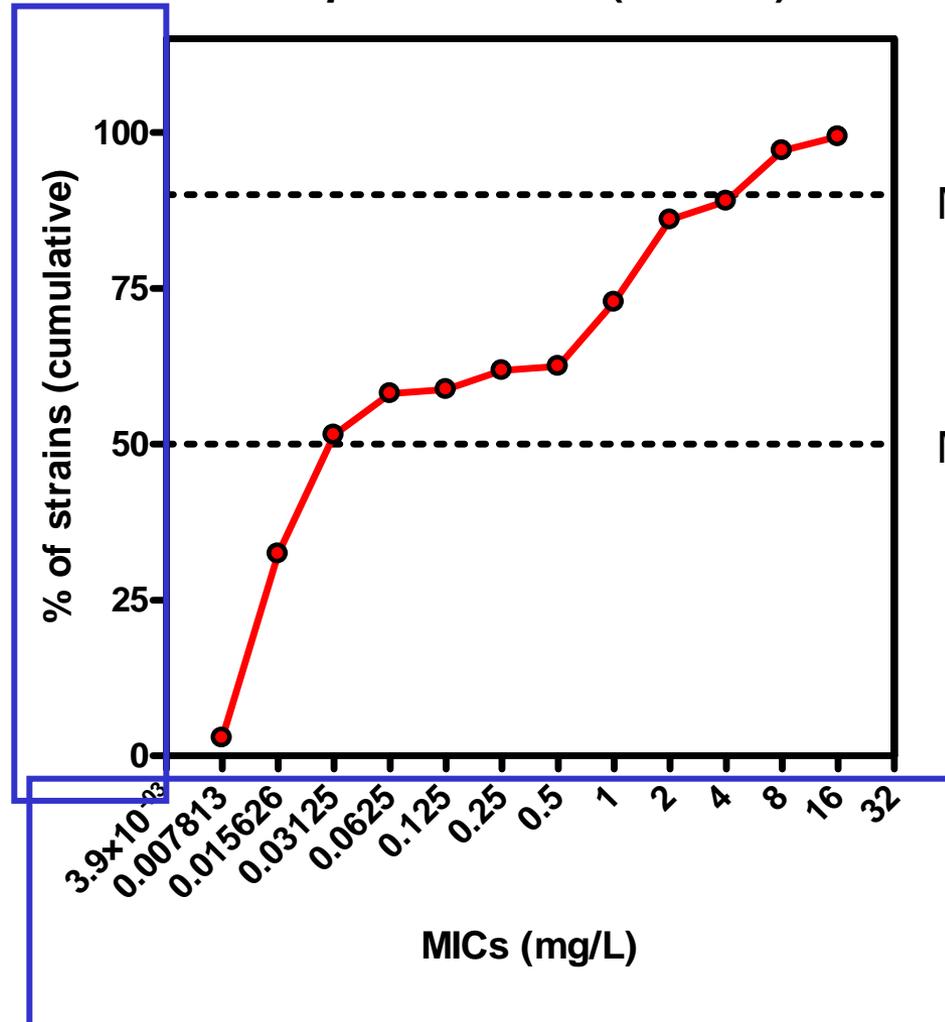
# But which breakpoints do we need to use ?

To be honest, I always wondered ...



# MIC distribution is a continuous variable...

amoxicillin vs.  
*S. pneumoniae* (n = 136)



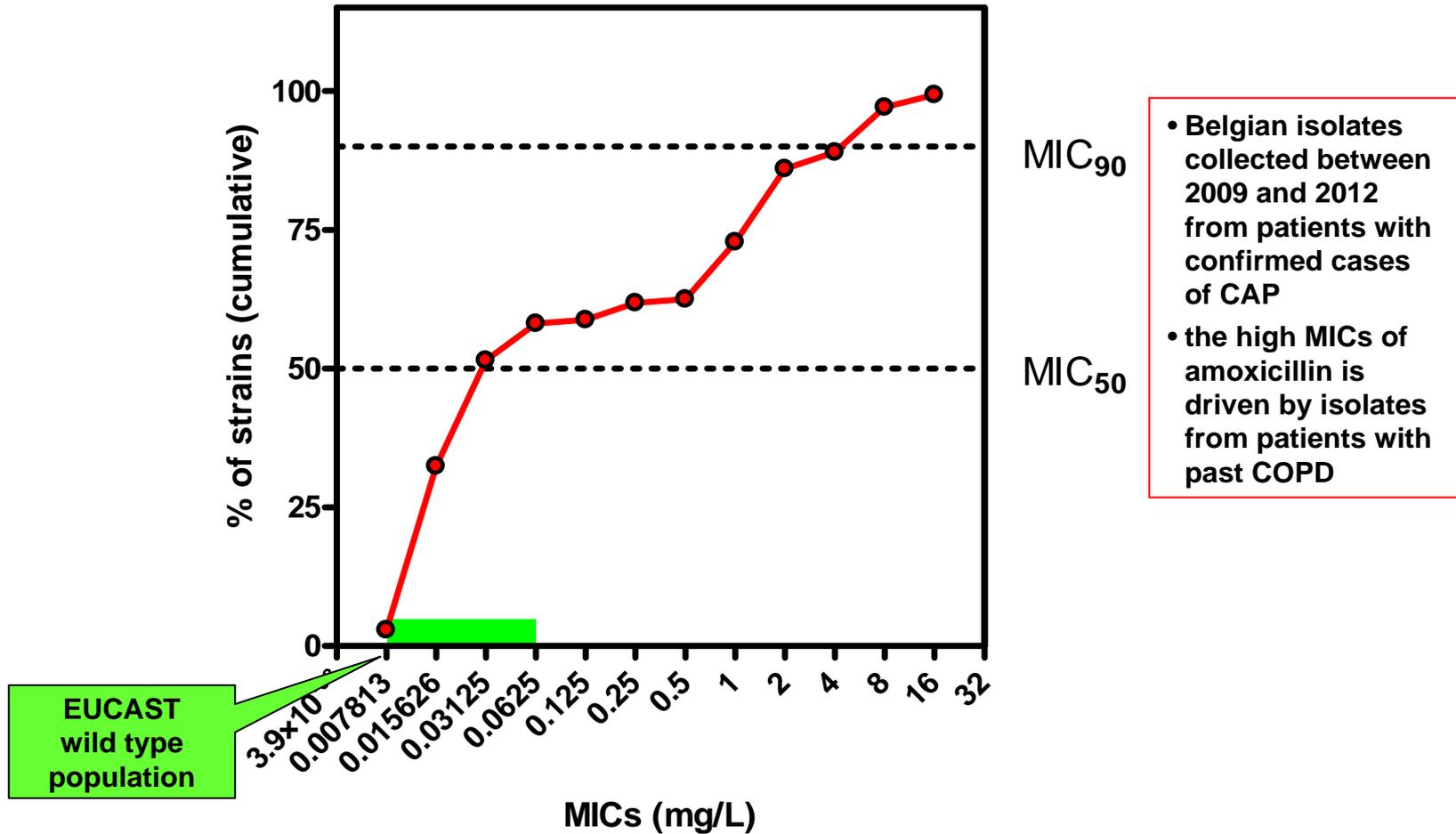
- Belgian isolates collected between 2009 and 2012 from patients with confirmed cases of CAP
- the high MICs of amoxicillin is driven by isolates from patients with past COPD

MIC minimum inhibitory concentration  
CAP community-acquired pneumonia  
COPD chronic obstructive pulmonary disease

Tulkens, unpublished

# MIC distribution is a continuous variable...

amoxicillin vs.  
*S. pneumoniae* (n = 136)



EUCAST: European Committee on Antimicrobial Susceptibility Testing (<http://www.eucast.org>)

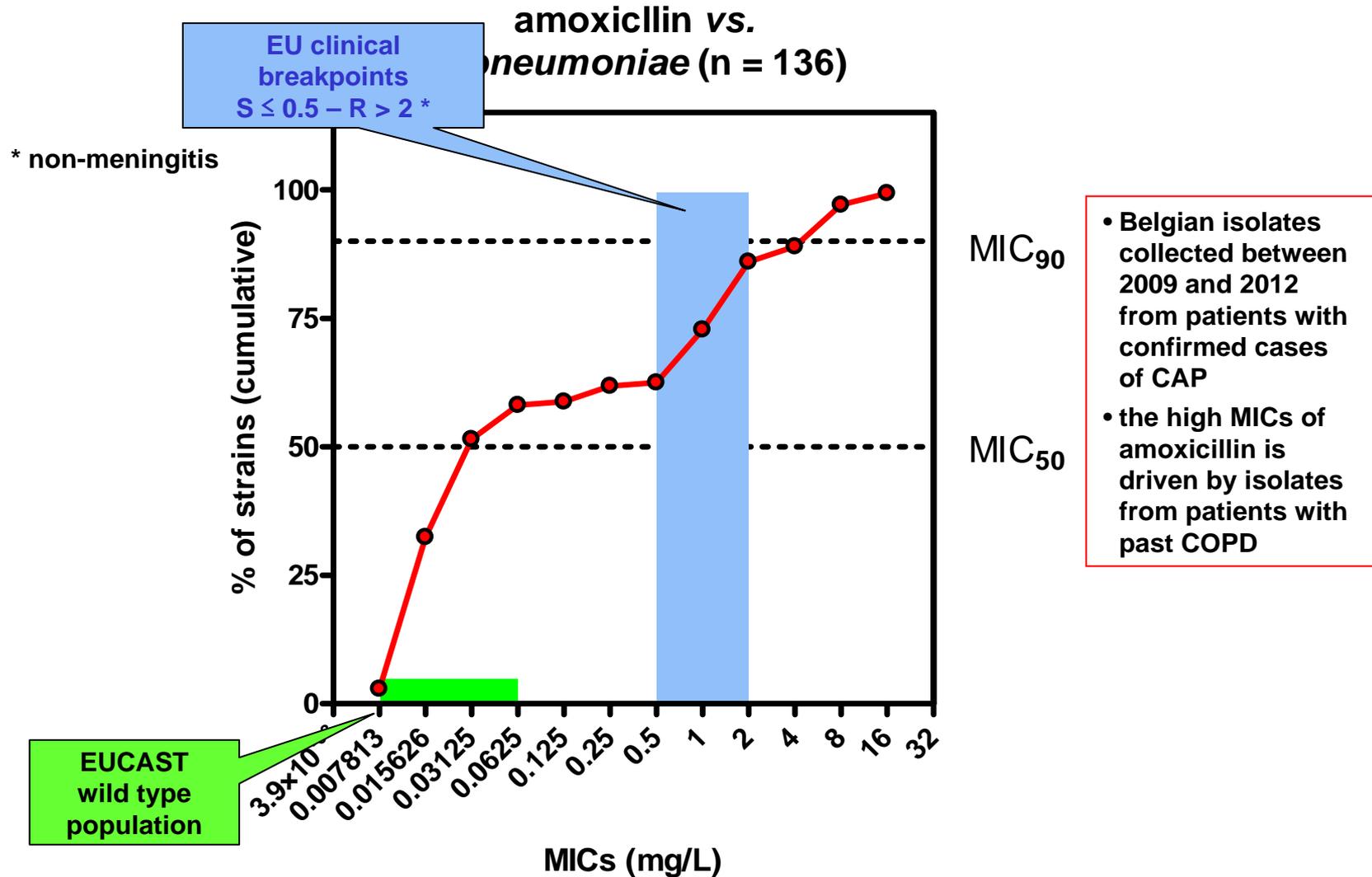
MIC: minimum inhibitory concentration

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COPD: chronic obstructive pulmonary disease

Tulkens, unpublished

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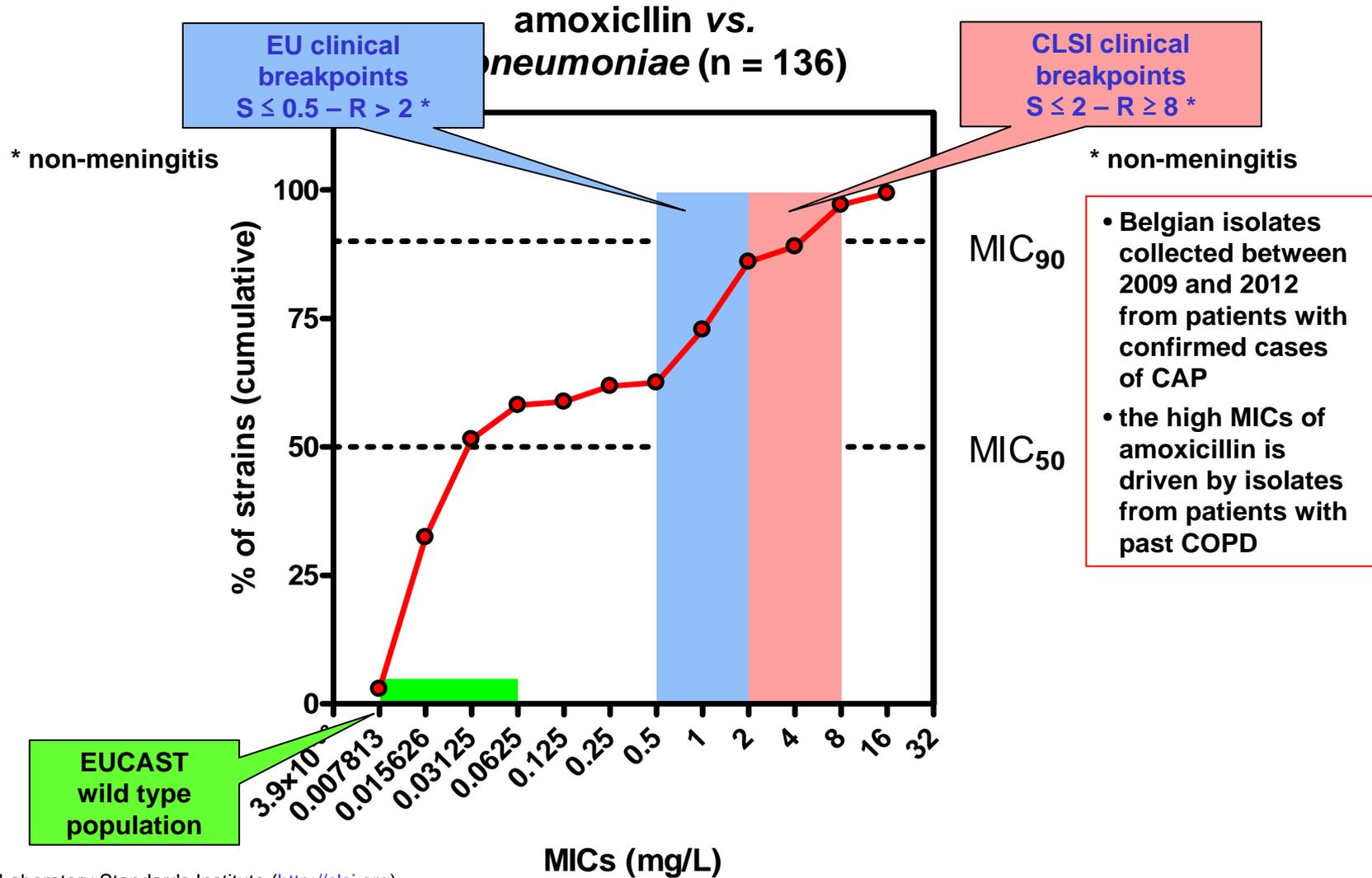
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Tulkens, unpublished

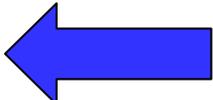
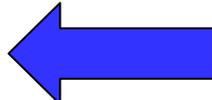
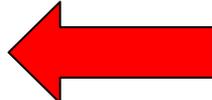
# MIC distribution is a continuous variable...



CLSI: Clinical and Laboratory Standards Institute (<http://clsi.org>)  
 EUCAST: European Committee on Antimicrobial Susceptibility Testing (<http://www.eucast.org>)  
 MIC: minimum inhibitory concentration  
 CAP: community-acquired pneumonia  
 COPD: chronic obstructive pulmonary disease

Tulkens, unpublished

# Warning about breakpoints (EUCAST vs. CLSI) for *S. pneumoniae* (non meningitis)

- With the [new] CLSI breakpoint (MIC  $\geq$  8 mg/L ), very few isolates will be defined as resistant.... 
- In fact, most experts believe that CAP caused by organisms with a penicillin MIC of 4 mg/L or higher (still an uncommon finding), can lead to an increased risk of death.<sup>1</sup> 
- For that reason, Europe has set its "R" breakpoint at > 2 mg/L.<sup>2</sup> 
- **Dosage adaptation over the original 250 mg BID is necessary for isolates with MIC between 0.25 and 2 mg/L ( → 0.5 g TID, 1 g TID, or extended-release forms ...)** 

CLSI: Clinical and Laboratory Standards Institute  
EUCAST: European Committee on Antimicrobial Susceptibility Testing  
MIC: minimum inhibitory concentration  
CAP: community acquired pneumonia  
R: resistance  
BID: twice daily; TID: 3 times daily

1. Feikin DR, *et al.* *Am J Public Health* 2000;90(2):223-9.  
2. EUCAST clinical breakpoints (<http://www.eucast.org>)  
(accessed 20/04/2014)

# Resistance of *S. pneumoniae* to macrolides and tetracyclines \*

\*analysis of resistance to erythromycin and doxycycline (with CAP as main indication) in surveillance systems or publications (*S. pneumoniae*)

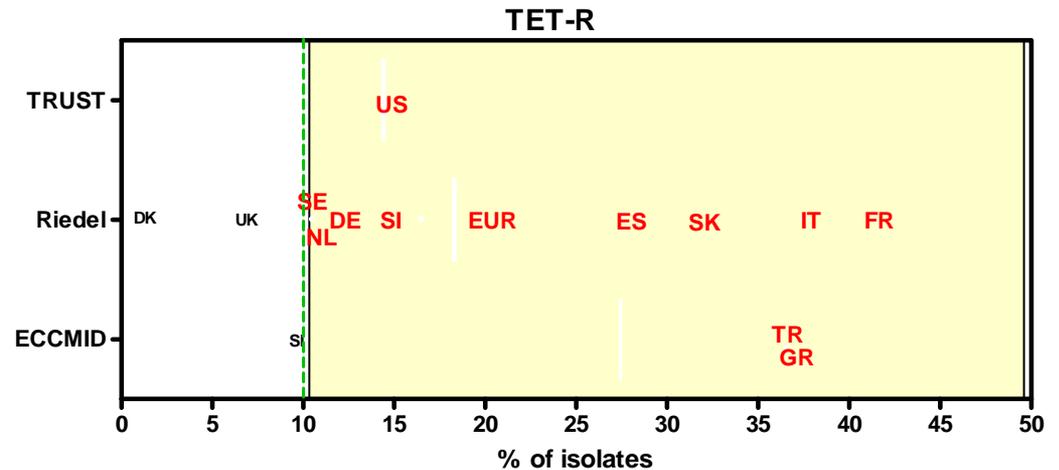
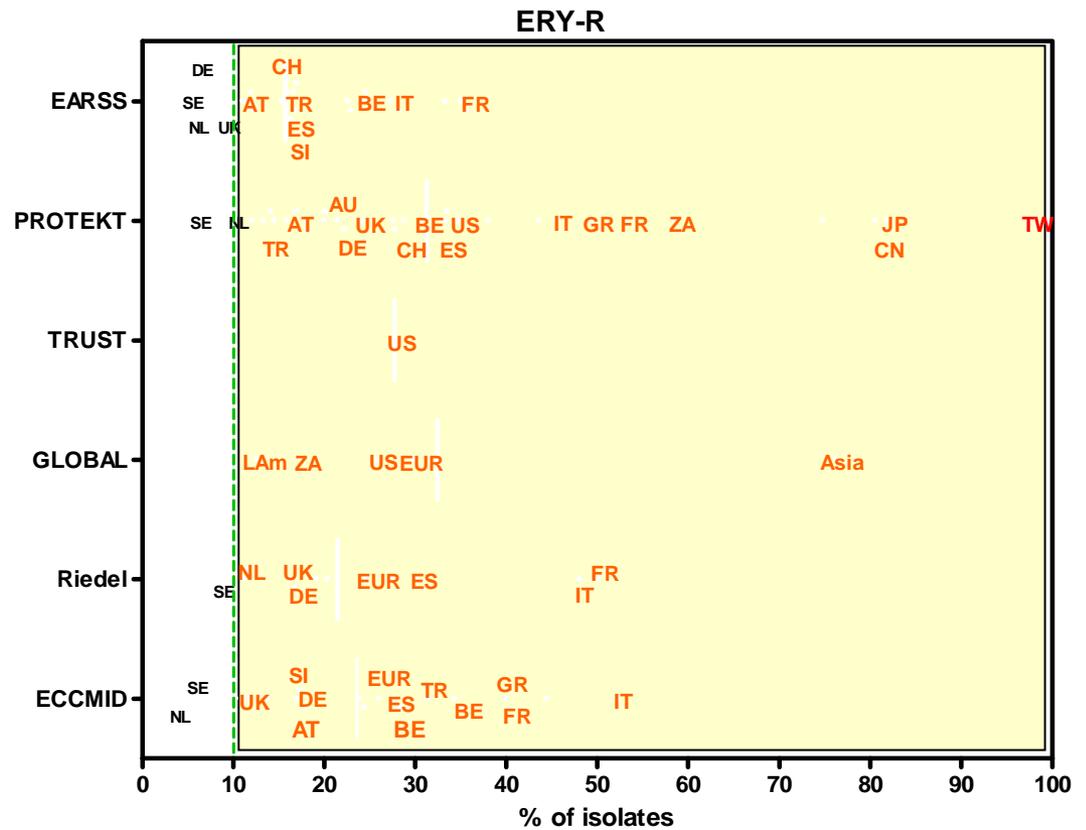
- **EARSS:** European Antimicrobial Surveillance system
- **PROTEKT:** Prospective Resistant Organism Tracking and Epidemiology for the Ketolide Telithromycin
- **TRUST:** Tracking Resistance in the United States Today
- **GLOBAL:** Global Landscape On the Bactericidal Activity of Levofloxacin
- **Riedel:** Eur J Clin Microbiol Infect Dis. 2007 Jul;26(7):485-90.
- **ECCMID:** abstracts of the 18th European Congress of Clinical Microbiology and Infectious Diseases

## Most studies used CLSI breakpoints

- erythromycin: S ≤0.25 – R ≥1
- Doxycycline: S ≤0.25 – R ≥1

Lismond *et al.*, in preparation

CAP: community-acquired pneumonia



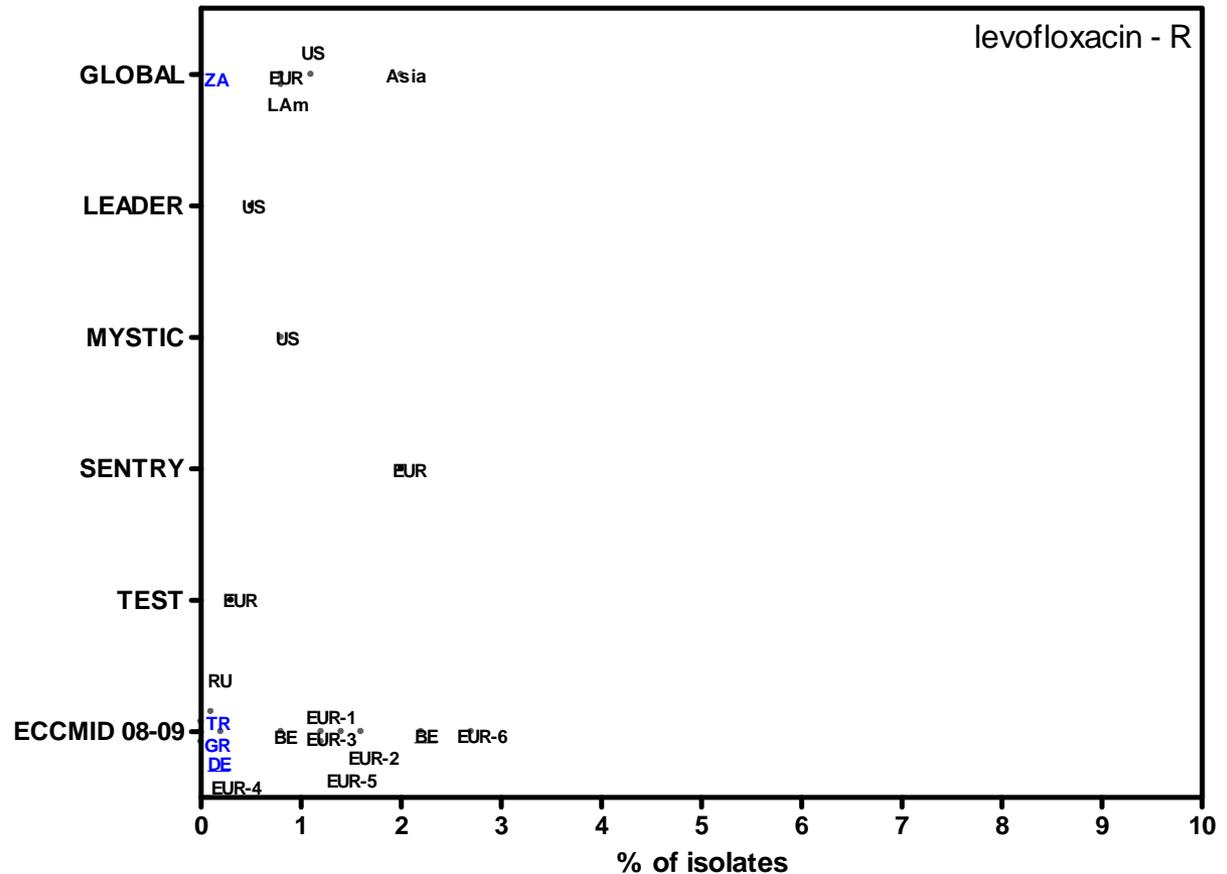
# Resistance of *S. pneumoniae* to fluoroquinolones

\*analysis of resistance of erythromycin and doxycycline (with CAP as main indication) in surveillance systems or publications (*S. pneumoniae*)

- **GLOBAL:** Global Landscape On the Bactericidal Activity of Levofloxacin
- **LEADER:** Linezolid Surveillance Program
- **MYSTIC:** Meropenem Yearly Susceptibility Test Information Collection
- **SENTRY:** Antimicrobial Surveillance Program (2005–2006)
- **TEST:** Tigecycline Evaluation Surveillance Trial
- **ECCMID 08-09 :** abstracts of the 18th and 19th European Congresses of Clinical Microbiology and Infectious Diseases

## Most studies used CLSI breakpoints

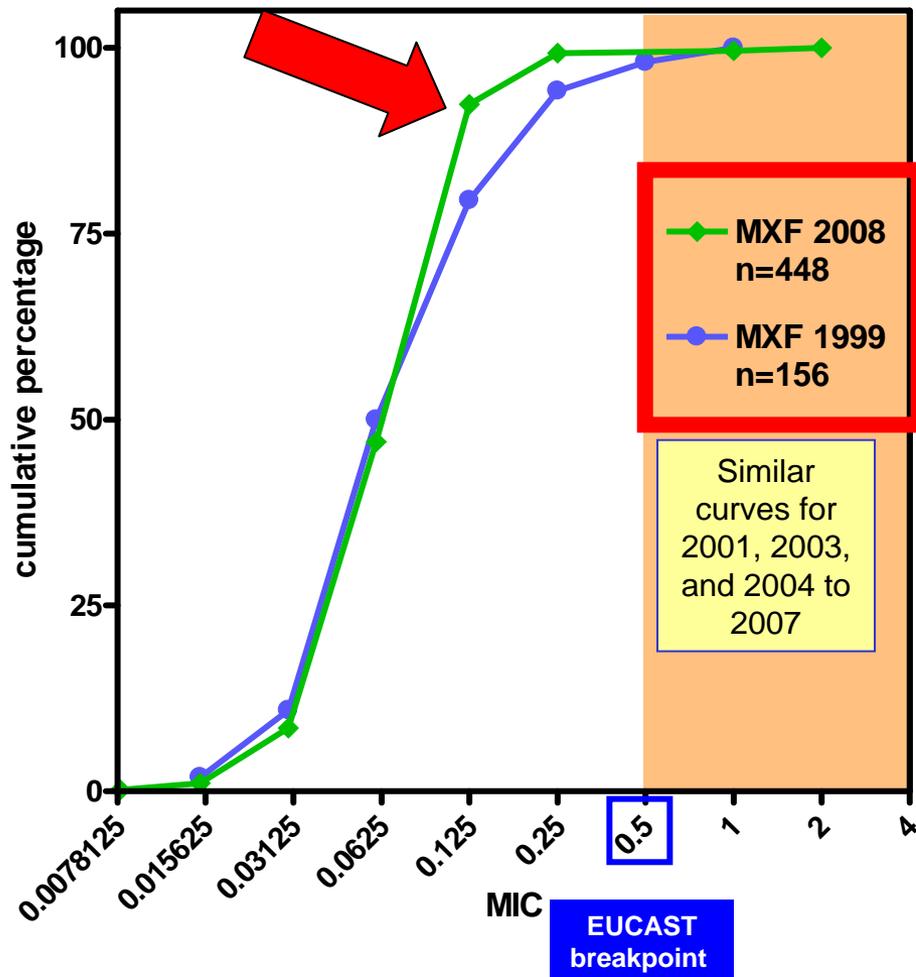
- levofloxacin: S  $\leq$  2 – R  $\geq$  8
- doxycycline: S  $\leq$  1 – R  $\geq$  4



Lismond *et al.*, in preparation

# Moxifloxacin MIC's against *S. pneumoniae* in Belgium from 1999 to 2008

## *S. pneumoniae* susceptibility to moxifloxacin in Belgium



- Extract from the data of a national collection based on annual surveys made by the Belgian Scientific Institute for Public Health for *S. pneumoniae* from community isolates [<https://www.wiv-isp.be/Programs/communicable-infectious-diseases/Pages/EN-BacterialDiseases.aspx?pfq=1033>] and presented at the 19th ECCMID. May, 16-19 2009, Helsinki (Vanhoof *et al* abstract no. O467 [<http://www.blackwellpublishing.com/eccmid19/abstract.asp?id=74082>; last visited: 2 may 2014])
- See also
  - Vanhoof *et al* Acta Clin Belg. 2006;61:49-57
  - Vanhoof *et al* Pathol Biol (Paris) 2010;58:147-151
- Confirmed in an independent study for the period 2004-2009 (Simoens *et al* Antimicrob Agents Chemother 2011;55:3051-3)
- Similar distribution for blood-stream isolates from patients with clinically confirmed diagnostic of CAP in 2007-2010 (Lismond *et al* Int J Antimicrob Agents. 2012;39(3):208-216)

MXF: moxifloxacin  
 CAP: community-acquired pneumoni  
 MIC: minimum inhibitory concentration

# Resistance of *S. pneumoniae* to fluoroquinolones

- The situation may be different in other countries (Asia)
  - 4% resistance to levofloxacin for PNRSP in China <sup>1</sup>
  - 8.6 % (6/70) in adults in China <sup>2</sup>
  - 4.7 % in Asian Countries (all cases from Korea, Hong-Kong, Taiwan) in association with previous treatment with fluoroquinolones, cerebrovascular disease, and healthcare-associated infection <sup>3</sup>

1. Jones RN, et al. *Diagn Microbiol Infect Dis.* 2013;77:258-66  
2. Guo Q, et al. *Eur J Clin Microbiol Infect Dis* 2014;33:465–70  
3. Kang CI, et al. *Eur J Clin Microbiol Infect Dis.* 2014;33:55-9

# Resistance of *S. pneumoniae* to fluoroquinolones

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  - 4.7 % in Asian Countries (all cases from Korea, Hong-Kong, Taiwan) in a study of 100 cases of pneumonia associated with levofloxacin-resistant *S. pneumoniae* <sup>3</sup>

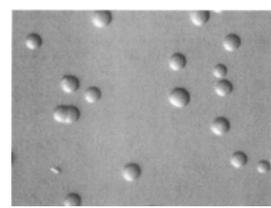
**Table 2** Independent risk factors associated with pneumonia caused by levofloxacin-nonsusceptible *S. pneumoniae*

Variables	Adjusted OR (95 % CI)	P
Previous treatment with fluoroquinolone	3.22 (1.05–9.85)	0.041
Cerebrovascular disease	2.88 (1.36–6.06)	0.005
Healthcare-associated infection	2.28 (1.14–4.55)	0.019

Kang et al. Eur J Clin Microbiol Infect Dis. 2014;33:55-9

1. Jones RN, et al. *Diagn Microbiol Infect Dis.* 2013;77:258-66
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# *Mycoplasma pneumoniae*



Waites & Talkington,  
Clin. Microbiol. Rev.  
2004;17:697-728

- must be recognized as a real potential pathogen if performing active surveillance

CLINICAL MICROBIOLOGY REVIEWS, Oct. 2004, p. 697-728  
0893-8512/04/\$08.00+0 DOI: 10.1128/CMR.17.4.697-728.2004  
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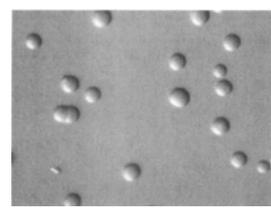
Vol. 17, No. 4

## *Mycoplasma pneumoniae* and Its Role as a Human Pathogen

Ken B. Waites<sup>1\*</sup> and Deborah F. Talkington<sup>2</sup>

*Department of Pathology, University of Alabama at Birmingham, Birmingham, Alabama 35249,<sup>1</sup> and  
Division of Bacterial and Mycotic Diseases, National Center for Infectious Diseases,  
Centers for Disease Control and Prevention, Atlanta, Georgia 30333<sup>2</sup>*

# Mycoplasma pneumoniae



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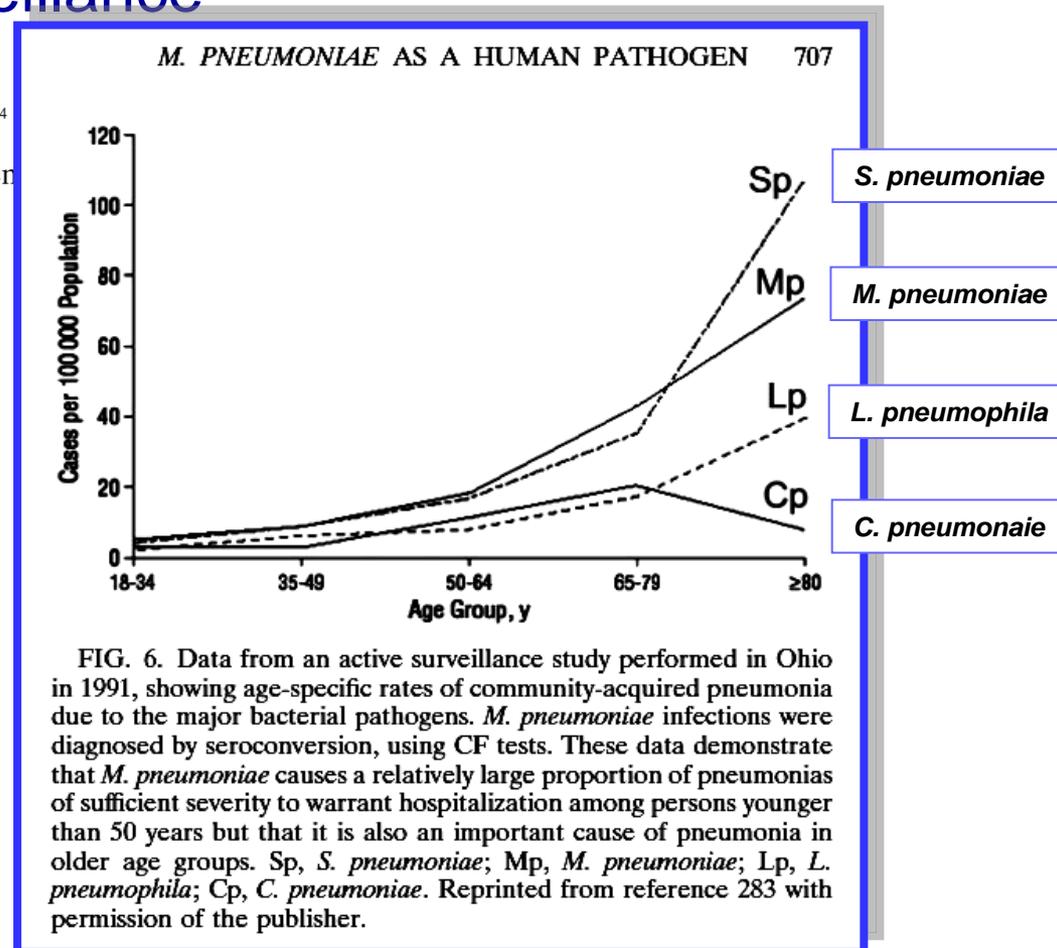
CLINICAL MICROBIOLOGY REVIEWS, Oct. 2004, p. 697-728  
0893-8512/04/\$08.00+0 DOI: 10.1128/CMR.17.4.697-728.2004  
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Vol. 17, No. 4

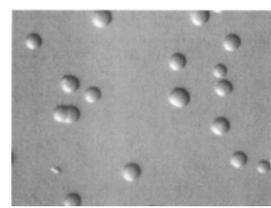
## *Mycoplasma pneumoniae* and Its Role as a Human Pathogen

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# *Mycoplasma pneumoniae*



Waites & Talkington,  
Clin. Microbiol. Rev.  
2004;17:697-728

- was long considered as universally susceptible to macrolides...
- but this was no longer true in Asia since several years ...



**Antimicrob Agents Chemother. 2013;57:4046-9.**

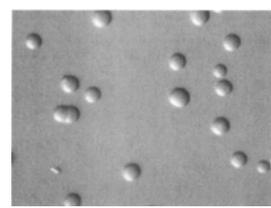
## Nationwide Surveillance of Macrolide-Resistant *Mycoplasma pneumoniae* Infection in Pediatric Patients

Yasuhiro Kawai,<sup>a</sup> Naoyuki Miyashita,<sup>b</sup> Mika Kubo,<sup>a</sup> Hiroto Akaike,<sup>a</sup> Atsushi Kato,<sup>a</sup> Yoko Nishizawa,<sup>a</sup> Aki Saito,<sup>a</sup> Eisuke Kondo,<sup>a</sup> Hideto Teranishi,<sup>a</sup> Tokio Wakabayashi,<sup>a</sup> Satoko Ogita,<sup>a</sup> Takaaki Tanaka,<sup>a</sup> Kozo Kawasaki,<sup>a</sup> Takashi Nakano,<sup>a</sup> Kihei Terada,<sup>a</sup> Kazunobu Ouchi<sup>a</sup>

Department of Pediatrics<sup>a</sup> and Department of Internal Medicine 1,<sup>b</sup> Kawasaki Medical School, Okayama, Japan

We conducted nationwide surveillance to investigate regional differences in macrolide-resistant (MR) *Mycoplasma pneumoniae* strains in Japan. The prevalence of MR *M. pneumoniae* in pediatric patients gradually increased between 2008 and 2012. Although regional differences were observed, high levels of MR genes were detected in all seven surveillance areas throughout Japan and ranged in prevalence from 50% to 93%. These regional differences were closely related to the previous administration of macrolides.

# Mycoplasma pneumoniae



Waites & Talkington,  
Clin. Microbiol. Rev.  
2004;17:697-728

- was long considered as universally susceptible to macrolides...
- but this was no longer true



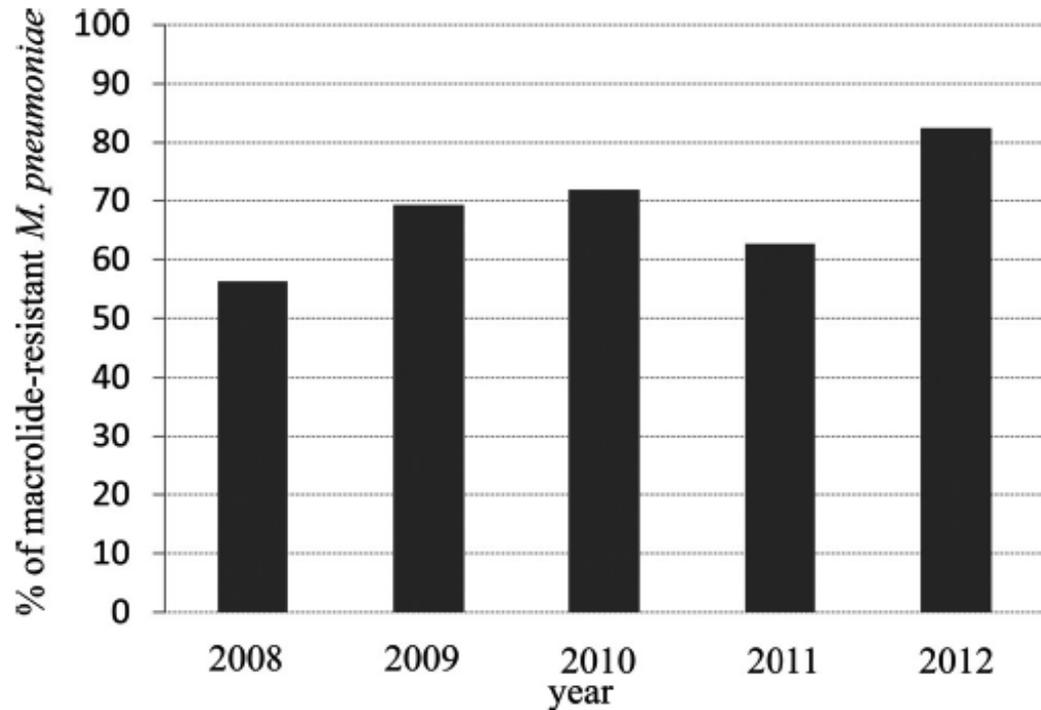
Antimicrob Agents Chemother

## Nationwide Surveillance of Macrolide-Resistant *M. pneumoniae* Infection in Pediatric Patients

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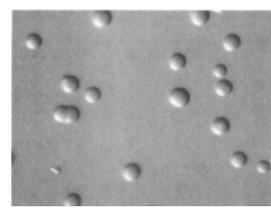
Department of Pediatrics<sup>a</sup> and Department of Internal Medicine 1,<sup>b</sup> Kawasaki Medical School, Okayama, Japan

We conducted nationwide surveillance to investigate regional differences in *M. pneumoniae* strains in Japan. The prevalence of MR *M. pneumoniae* in pediatric patients increased from 50% to 93% from 2008 to 2012. Regional differences were observed, high levels of MR genes were detected in all regions. These regional differences were closely related to the prevalence from 50% to 93%. These regional differences were closely related to the prevalence from 50% to 93%.



1 Year-by-year increases in the frequency of macrolide-resistant *Mycoplasma pneumoniae* cases from 2008 to 2012.

# *Mycoplasma pneumoniae*



Waites & Talkington,  
Clin. Microbiol. Rev.  
2004;17:697-728

- and Resistance is arriving in Europe



Antimicrobial Agents and Chemotherapy 2014 58 1265–1266

LETTER TO THE EDITOR

## First Report of Macrolide Resistance in a *Mycoplasma pneumoniae* Isolate Causing Community-Acquired Pneumonia in Spain

Juan de Dios Caballero,<sup>a,b,c</sup> Rosa del Campo,<sup>a,b,c</sup> María del Carmen Mafé,<sup>d</sup> María Gálvez,<sup>a</sup> Mario Rodríguez-Domínguez,<sup>a,b,c</sup> Rafael Cantón,<sup>a,b,c</sup> María Antonia Meseguer,<sup>a</sup> José Manuel Hermida<sup>d</sup>

Servicio de Microbiología, Hospital Universitario Ramón y Cajal, Madrid, Spain<sup>a</sup>; Instituto Ramón y Cajal de Investigación Sanitaria (IRYCIS), Madrid, Spain<sup>b</sup>; Spanish Network for Research in Infectious Diseases (REIPI), Instituto de Salud Carlos III, Madrid, Spain<sup>c</sup>; Servicio de Enfermedades Infecciosas, Hospital Universitario Ramón y Cajal, Madrid, Spain<sup>d</sup>

**A previously healthy 23-year-old Chinese who had been studying in Spain for 1 year but returned from a 1-month trip to China and Korea 13 days before the onset of symptoms.**

# Haemophilus: is it important ?



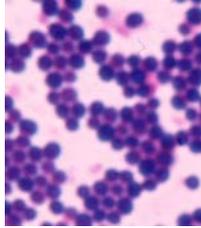
<http://www.pathologyoutlines.com/topic/lymphnodeshinfluenzae.html>

- *Haemophilus* is often considered as a colonizer of the upper respiratory tract with risks only for patients with COPD
- However, in coinfection with a preceding viral infection, *Haemophilus* may easily colonize the lung, leading to lethal secondary bacterial pneumonia.
  - We may now understand the corresponding genetic background (e.g. overexpression of an anti-oxidant protein) <sup>1</sup>
- $\beta$ -lactamase-negative ampicillin-resistant (BLNAR) *Haemophilus* may be on the rise in some regions in Europe (but not all) <sup>2</sup>
  - antibiotic discs may fail to fully separate between BLNAS and BLNAR populations <sup>3</sup>
  - the majority of invasive *H. influenzae* (including BLNAR) remain susceptible to third-generation cephalosporins and fluoroquinolones in Europe <sup>4</sup>
- Resistance of *Haemophilus* to fluoroquinolones may be on the rise in Asia <sup>5</sup>

1. Wong, *et al. Proc Natl Acad Sci U S A.* 2013;110:15413-8.
2. Dabernat, *et al. Eur J Clin Microbiol Infect Dis.* 2012;31:2745-53  
Geelen, *et al. Scand J Infect Dis.* 2013;45:606-11
3. Garcia-Cobos, *et al. JAC.* 2013;68: 159–63
4. Garcia-Cobos, *et al JAC.* 2014;69:111-6  
Puig, *et al.. PLoS One.* 2013;13-8:e82515
5. Shoji, *et al. J Infect Chemother.* 2014;20:250-5

COPD chronic obstructive pulmonary disease  
BLNAR  $\beta$ -lactamase-negative ampicillin-resistant  
BLNAS  $\beta$ -lactamase-negative ampicillin-sensitive

# Staphylococcus aureus



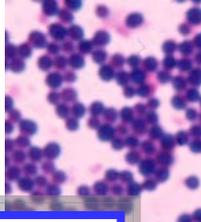
[http://www.microbeworld.org/index.php?option=com\\_jlibrary&view=article&id=7611](http://www.microbeworld.org/index.php?option=com_jlibrary&view=article&id=7611)

- Nosocomial pneumonia involving hospital-acquired (HA) *S. aureus* is becoming increasingly frequent <sup>1,2</sup>
- In parallel, pneumonia caused by community-acquired (CA) MRSA while remaining rare in Europe<sup>2</sup> are becoming common in several other parts of the world including Asia <sup>3</sup>
- As many strains (even MSSA) produce toxins, they cause major tissue damage, and, hence a high mortality <sup>3,4,5</sup>

1. Jones, *Clin Infect Dis.* 2010;51(suppl 1):S81-7
2. Valour, *et al Rev Pneumol Clin.* 2013;69:368-82
3. Karampela, *et al. Minerva Anesthesiol.* 2012 Aug;78(8):930-40  
Kang & Song. *Infect Chemother.* 2013;45:22-31
4. Papazian & Donati. Nosocomial pneumonia. *In Infectious Diseases*, 3rd Edition, Cohen, Powderly & Opal, eds. Elsevier  
(available on line at <http://www.expertconsultbook.com> ; last visited: 4 April 2014)
5. Catena, *et al Infez Med.* 2012;20:205-10 /.

MRSA methicillin-resistant *Staphylococcus aureus*  
MSSA methicillin-sensitive *Staphylococcus aureus*

# S. aureus



- Nosocomial pneumonia
- S. aureus* is becoming

- In parallel pneumonia

"*S. aureus* accounts for 2 to 5% of the etiologies of community-acquired pneumonia"

- major tissue damage

"*S. aureus* represents 20 to 30% of cases of hospital-acquired pneumonia, including ventilator-associated pneumonia"

Revue de Pneumologie clinique (2013) xxx, xxx–xxx



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SÉRIE : MICROBIOLOGIE ET APPAREIL RESPIRATOIRE  
**Infections broncho-pulmonaires à *Staphylococcus aureus***  
*Staphylococcus aureus* broncho-pulmonary infections

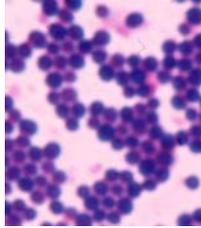
F. Valour<sup>a,b,c,d</sup>, N. Chebib<sup>a</sup>, Y. Gillet<sup>b,c,d,e</sup>, P. Reix<sup>b,f</sup>,  
F. Laurent<sup>b,c,d,g</sup>, C. Chidiac<sup>a,b,c,d</sup>, T. Ferry<sup>a,\*,b,c,d</sup>

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<sup>b</sup> Université Claude-Bernard Lyon 1, 69008 Lyon, France  
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<sup>f</sup> Service de pneumologie, allergologie, mucoviscidose, hospices civils de Lyon, hôpital Femme-Mère-Enfant, 69500 Bron, France  
<sup>g</sup> Laboratoire de bactériologie, hospices civils de Lyon, groupement hospitalier Nord, 69004 Lyon, France

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- Jones CJ, et al. (2013) S81-7
-

# S. aureus



[http://www.microbeworld.org/index.php?option=com\\_jlibrary&view=article&id=7611](http://www.microbeworld.org/index.php?option=com_jlibrary&view=article&id=7611)

- Nosocomial pneumonia involving hospital-acquired (HA) *S. aureus*
- In parallel (CA) MRSA in several
- As many as major tissue

**Table 3. Regional Incidence of Pathogens Isolated from Patients Hospitalized with Pneumonia in the Last 5 Years of the SENTRY Antimicrobial Surveillance Program (31,436 Cases).**

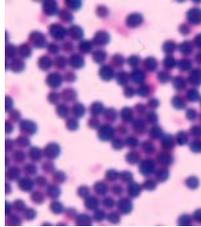
Pathogen	Incidence, %			
	All regions	United States	Europe	Latin America
<i>Staphylococcus aureus</i>	28.0	36.3	23.0	20.1
<i>Pseudomonas aeruginosa</i>	21.8	19.7	20.8	28.2
<i>Klebsiella</i> species	9.8	8.5	10.1	12.1
<i>Escherichia coli</i>	6.9	4.6	10.1	5.5
<i>Acinetobacter</i> species	6.8	4.8	5.6	13.3
<i>Enterobacter</i> species	6.3	6.5	6.2	6.2
<i>Serratia</i> species	3.5	4.1	3.2	2.4
<i>Stenotrophomonas maltophilia</i>	3.1	3.3	3.2	2.3
<i>Streptococcus pneumoniae</i>	2.9	2.5	3.6	2.4
<i>Haemophilus influenzae</i>	2.7	2.5	3.7	1.3

- Jones Clin Infect Dis 2010;51(suppl 1):S81-7
- Karampela et al Minerva Medica 2010;111(1):1-6
- Papazian & Donati Nosocomial Infection 2006;11(1):1-6  
<http://www.expertconsult.com>

Jones Clin Infect Dis 2010;51(suppl 1):S81-7

32.

# S. aureus



[http://www.microbeworld.org/index.php?option=com\\_jlibrary&view=article&id=7611](http://www.microbeworld.org/index.php?option=com_jlibrary&view=article&id=7611)

- Nosocomial pneumonia  
S. aureus is becoming
- In parallel, pneumonia  
(CA) MRSA which  
in several other
- As many strains  
major tissue damage

**Table 4. Frequency of Bacterial Pathogens Associated with Hospital-Acquired Bacterial Pneumonia (HABP) and Ventilator-Associated Bacterial Pneumonia (VABP)**

Organism	Percentage of isolates (no)	
	HABP (n = 835)	VABP (n = 499)
MRSA	47.1 (48.6)	42.5 ( <b>34.4</b> )
<i>Pseudomonas</i> species	18.4	21.2
<i>Klebsiella</i> species	7.1	8.4
<i>Haemophilus</i> species	5.6	<b>12.2</b>
<i>Enterobacter</i> species	4.3	5.6
<i>Streptococcus pneumoniae</i>	3.1	5.8
<i>Acinetobacter</i> species	2.0	3.0

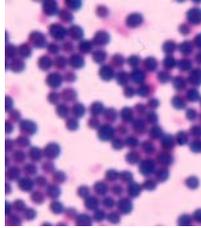
**NOTE.** Data are from [2, 7]. Boldface indicates a significant change or difference in incidence compared with HABP. MRSA, methicillin-resistant *Staphylococcus aureus*.

- Jones Clin Infect Dis 2010;51(suppl 1):S81-7
- Karampela et al Minerva Anestesiol. 2013;69:368-82.
- Papazian & Donati Nosocomial pneumonia  
<http://www.expertconsultbook.com> (L

HABP: hospital-acquired bacterial pneumonia  
VABP: ventilator-associated bacterial pneumonia  
MRSA: methicillin resistant *Staphylococcus aureus*

Jones, et al. Clin Infect Dis. 2010;51(suppl 1):S81-7

# S. aureus



[http://www.microbeworld.org/index.php?option=com\\_jlibrary&view=article&id=7611](http://www.microbeworld.org/index.php?option=com_jlibrary&view=article&id=7611)

- Nosocomial *S. aureus*
- In parallel (CA) MRSA in several
- As many major tiss

*Le Infezioni in Medicina*, n. 3, 205-210, 2012

*Casi clinici*

*Case reports*

## **Necrotizing pneumonia caused by Panton-Valentine leukocidin-producing methicillin-susceptible *Staphylococcus aureus* (MSSA)**

***Polmonite necrotizzante causata da Staphylococcus aureus meticillino sensibile produttore di leucocidina di Panton-Valentine (MSSA)***

Vincenzo Catena<sup>1,2</sup>, Marco Baiocchi<sup>2</sup>, Paolo Lentini<sup>3</sup>, Luigi Badolati<sup>2</sup>,  
Monica Baccarin<sup>2</sup>, Daniele D. Del Monte<sup>1</sup>, Alessandro Rubini<sup>4</sup>

<sup>1</sup>Dipartimento di Emergenza e Terapia Intensiva, U.L.S.S. 2, Feltre, Belluno, Italy;

<sup>2</sup>Dipartimento di Emergenza e Terapia Intensiva, Ospedale "San Bassiano", Bassano del Grappa, Vicenza, Italy;

<sup>3</sup>Dipartimento Nefrologia e Dialisi, Ospedale "S. Bassiano", Bassano del Grappa, Vicenza, Italy;

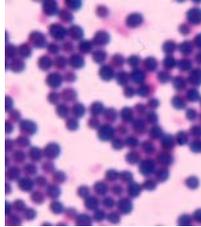
<sup>4</sup>Dipartimento Scienze Biomediche, Università di Padova, Padova, Italy

1. Jones Clin Infect Dis
2. Karampela et al Mine
3. Papazian & Donati N  
<http://www.expertcons>

8-82.

Catena, et al *Infez Med.* 2012;20:205-10

# S. aureus



[http://www.microbeworld.org/index.php?option=com\\_jlibrary&view=article&id=7611](http://www.microbeworld.org/index.php?option=com_jlibrary&view=article&id=7611)

- Nosocomial pneumonia caused by *S. aureus* is becoming increasingly common.
- In parallel, pneumonia caused by community-acquired MRSA (CA-MRSA) which is resistant to several other antibiotics.
- As many strains of MRSA are causing major tissue damage.



Figure 1 - Chest X-Ray at the admission.



Figure 2 - Chest CT scan at the admission: diffuse bilateral alveolar infiltrates.

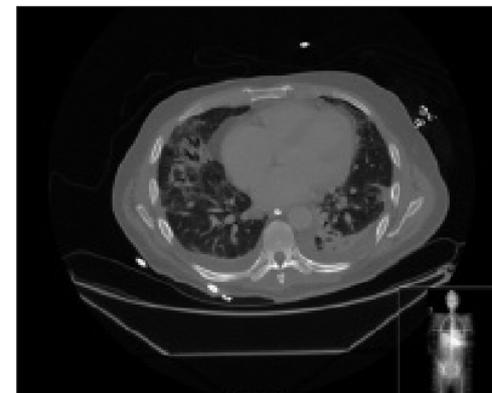
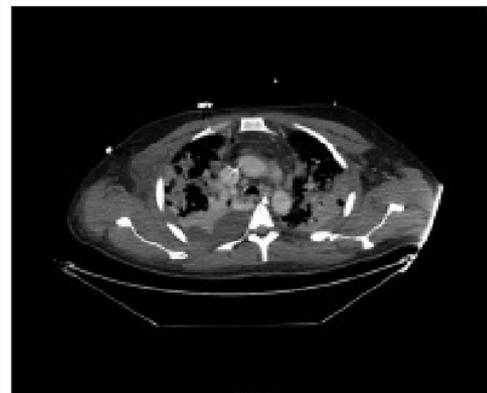


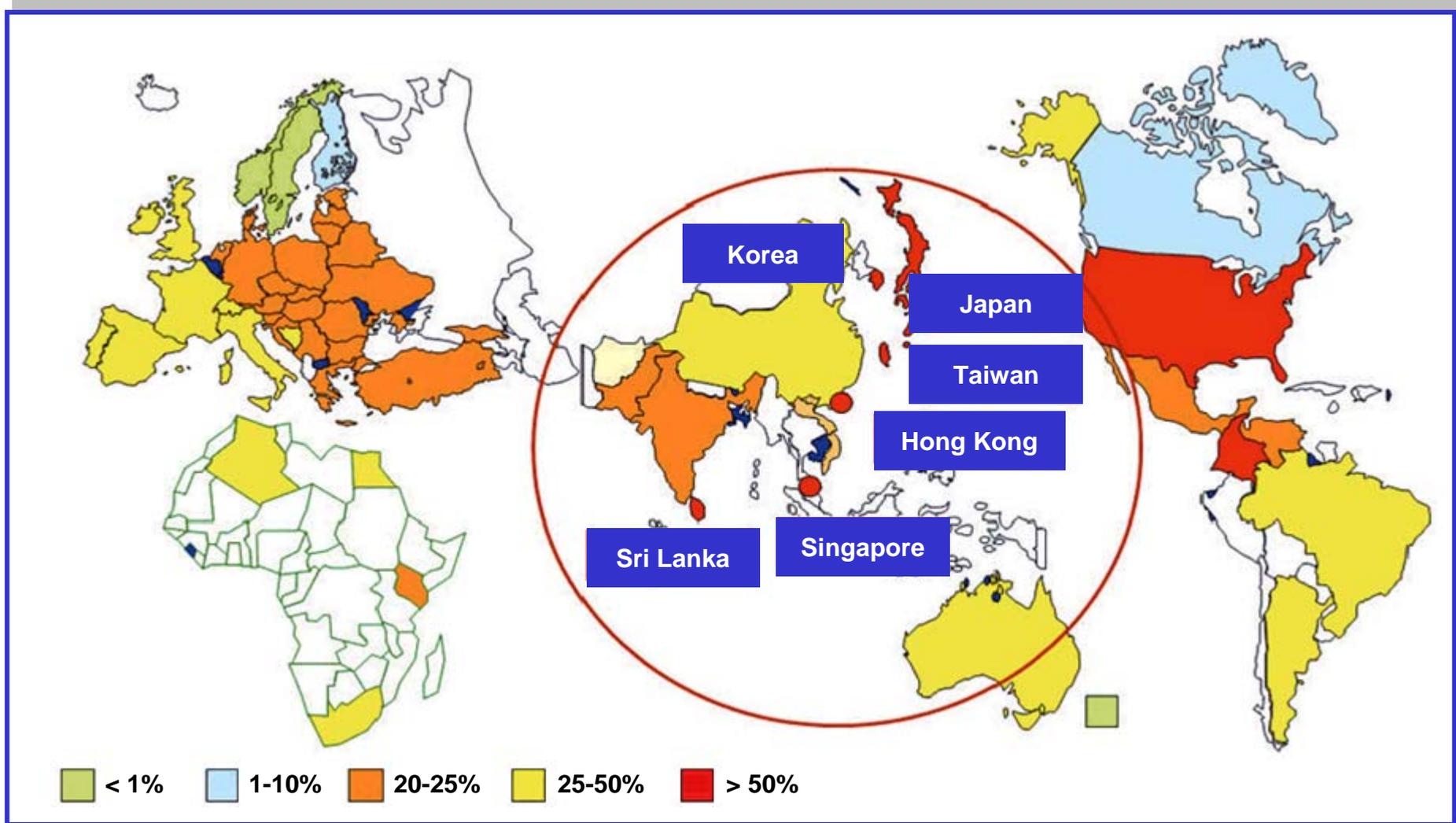
Figure 3 - Chest CT scans during ICU stay: extensive bilateral pleural effusions, diffuse bilateral alveolar infiltrates and nodular opacities with cavity forming consistent with necrotizing pneumonia.

1. Jones Clin Infect Dis 2010;51(suppl 1)
2. Karampela et al Minerva Anestesiol. 2010
3. Papazian & Donati Nosocomial pneumonia <http://www.expertconsultbook.com> (Lancet 2013;381:1237-45)

Catena, et al. Infez Med. 2012;20:205-10

# MRSA in Asia

Prevalence of methicillin resistance among *S. aureus* isolates.  
Some Asian countries have shown the highest prevalence rates of MRSA

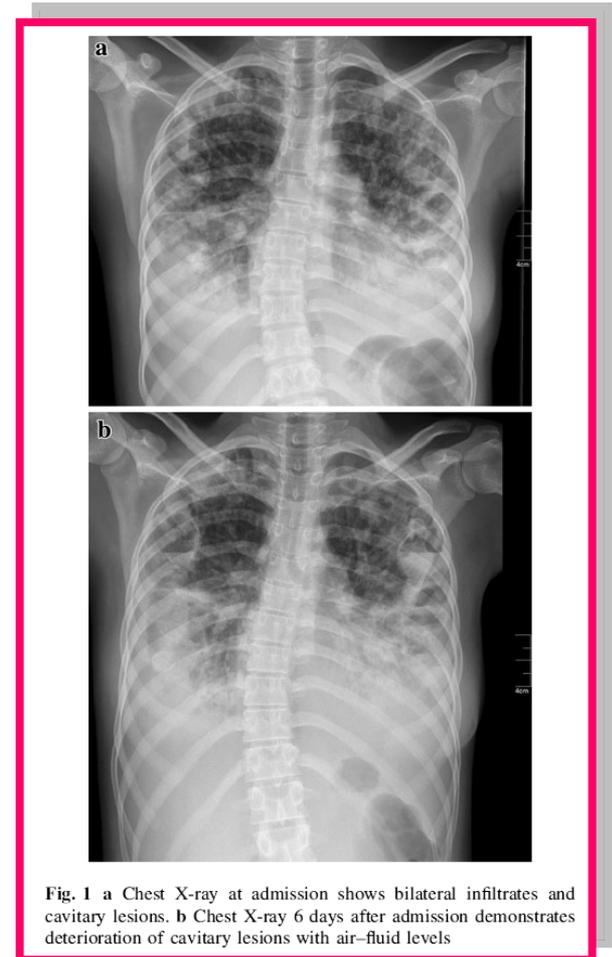


# *S. aureus* in Africa

- Very little is known about Africa !
- But data that are coming are challenging...

**Huson *et al.*** *Infection*. 2014 Jan 25. [Epub ahead of print]

- high prevalence (20-28 %) of MRSA in urban hospitals (as opposed to rural) in Cameroon
- typical case of *S. aureus* pneumonia (strain resistant to penicillin, cloxacillin, ciprofloxacin, and erythromycin)



# Anaerobes and lung diseases



*B. fragilis*  
CDC Public Health  
Image Library  
<http://phil.cdc.gov/phil>

- Anaerobic bacteria are frequent in aspiration pneumonia and associated complications (aspiration pneumonitis, lung abscess, necrotizing pneumonia and empyema) <sup>1</sup>
- While microbiological documentation is difficult, failure to direct adequate therapy against anaerobes (if present) may lead to clinical failures <sup>2</sup>
- Treatment of anaerobic infection is complicated by the slow growth of these organisms, by the polymicrobial nature of the infections, and by the growing resistance of anaerobic bacteria to antimicrobials (see next slide but only very rare cases for metronidazole <sup>3</sup>)

1. Bartlett. *Anaerobe*. 2012;18:235-9
2. Brook. *Adv Exp Med Biol*. 2011;697:117-52 - In N. Curtis et al. (eds.), *Hot Topics in Infection and Immunity in Children VII*, Springer.
3. Centers for Disease Control and Prevention (CDC) *MMWR Morb Mortal Wkly Rep*. 2013;62:694-6.

# Anaerobes: resistance to other antibiotics than metronidazole

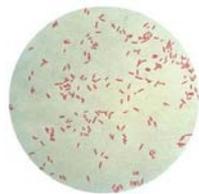
Percent resistance of *Bacteroides fragilis* group isolates to antimicrobial agents<sup>a,b</sup>

	Antimicrobial	CLSI breakpoints → MIC breakpoint (μg/ml)		% resistance to antimicrobial	
		Susceptible	Resistant	<i>B. fragilis</i>	<i>B. fragilis</i> group
≤4 >8	Ampicillin-sulbactam	≤8/4	≥32/16	2.8–11	
≤4 >8	Amoxicillin-clavulanate	≤4/2	≥16/8	4–37	10–20
≤8 >16	Piperacillin-tazobactam	≤32/4	≥128/4	0–5	0–8
NA	Cefoxitin	≤16	≥64	4–25	17–33
≤1 >1	Ertapenem	≤4	≥16	1.4–10	
≤2 >8	Imipenem	≤4	≥16	0.3–7	<1–1
≤2 >8	Meropenem	<4	≥16	1.2–22	
≤1 >1	Doripenem	≤4	≥16	1.3–12	
≤4 >4	Clindamycin	≤2	≥8	10–42	32–52
IE	Moxifloxacin	≤2	≥8	10–41	14–57
no correl.	Tigecycline	≤4	≥16	2–11	2–13

<sup>a</sup> Including intermediate-resistant strains. Metronidazole is not included since >99% of Gram-negative strains are susceptible.

# Gram-negatives

(beyond *Haemophilus*, *Legionella*, ...)



*P.aeruginosa*  
CDC Public Health  
Image Library  
<http://phil.cdc.gov/phil>

- May coexist with Gram-positive organisms...
- But remain the primary causative pathogens of nosocomial pneumonia <sup>1</sup>
- Main organisms include *Escherichia coli*, *Klebsiella pneumoniae*, *Enterobacter species*, *Pseudomonas aeruginosa*, and *Acinetobacter baumannii* <sup>2</sup>
- Inadequate initial therapy is unambiguously linked with increase mortality rate <sup>3</sup>
- Resistance rates vary very much from hospital to hospital and from ward to ward, but risk factors may be identified <sup>1</sup>:
  - hospitalization > 5 days, high resistance rates in the area or specific hospital,
  - immunosuppressive diseases and/or drugs, use of antibiotics in the last 90 days,
  - for health care-associated pneumonia: hospitalization for 2 days within 90 days, residence in a nursing home or extended care facility, home infusion therapy, chronic dialysis within 30 days, home wound care, or a family member with an MDR pathogen

1. Arnold *et al* Intensive Care Med. 2010;25:259-70
2. Boucher, *et al. Clin Infect Dis.* 2009;48:1-12.
3. Leibovici, *et al. J Intern Med* 1998;244:379-86  
Ibrahim, *et al. Chest.* 2000; 118:146-55  
Regui *et al* Chest. 2002;122:262-8  
Micek *et al* AAC. 2005;1306-11.

MDR multi drug resistant

# Gram-negatives

(beyond *Haemophilus*, *Legionella*, ...)

## Resistance

- must be assessed locally \*
- often anticipated to be high \*
- could be influenced by previous treatments

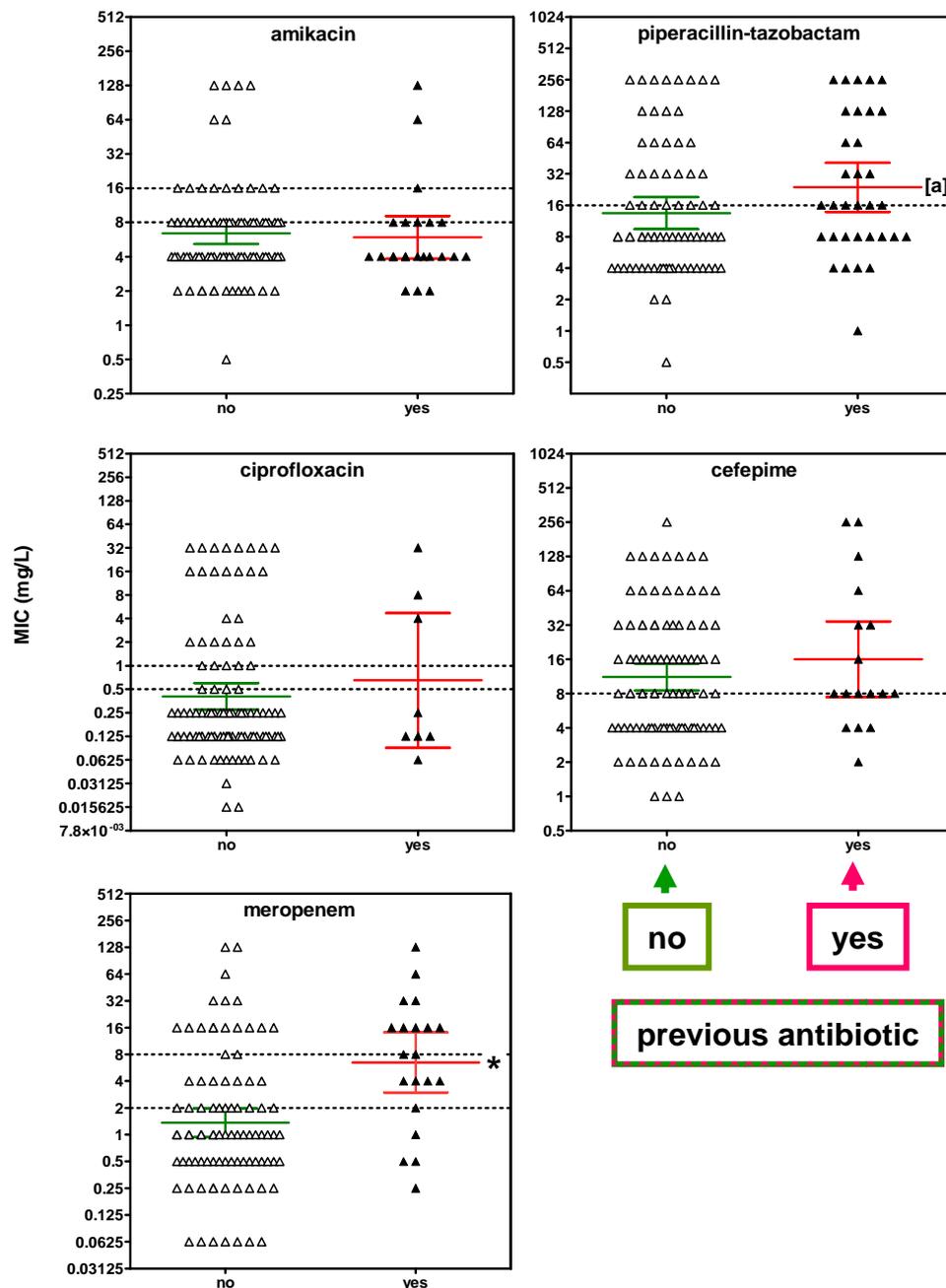
\* may not be the case in all hospitals  
(Sivert, *et al. Infect Control Hosp Epidemiol* 2013;34:1-14)

**MIC of 5 antibiotics used in empiric antipseudomonal therapy towards initial *P. aeruginosa* isolates of ICU patients with suspected nosocomial infection in 5 hospitals in Belgium**

- stratification between patients having either not received (no) or received (yes) the corresponding drug within 1 month prior to the collection of the isolate.
- the horizontal dotted lines are the corresponding S and R EUCAST breakpoints

ICU intensive care unit

Riou, *et al. Int J Antimicrob Agents.* 2010;36:513-22



# Gram-negatives in Europe and Mediterranean Area

International Journal of Antimicrobial Agents 43 (2014) 328–334



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journal homepage: <http://www.elsevier.com/locate/ijantimicag>



Antimicrobial susceptibility of Gram-negative organisms isolated from patients hospitalised with pneumonia in US and European hospitals: Results from the SENTRY Antimicrobial Surveillance Program, 2009–2012

Helio S. Sader\*, David J. Farrell, Robert K. Flamm, Ronald N. Jones

*JMI Laboratories, 345 Beaver Creek Center, Suite A, North Liberty, IA 52317, USA*



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JMI Laboratories, 345 Beaver Creek Center, Suite A, No

Antimicrobial susceptibility of Gram-negative bacterial organisms isolated from patients hospitalised with pneumonia in US and Europe and the Mediterranean region (EMR) medical centres (2009–2012).

Organism/antimicrobial agent	%S/%R (no. tested)		
	EUCAST <sup>a</sup>		
	USA	EMR	
<i>Pseudomonas aeruginosa</i>	(1439)	(1250)	
TZP	72.9/27.1	63.9/36.1	←
Ceftazidime	79.6/20.4	68.7/31.3	←
Cefepime	80.4/19.6	72.1/27.9	←
Meropenem	76.3/9.0	65.8/14.4	
Amikacin	92.2/3.8	82.8/11.2	
Gentamicin	87.0/13.0	75.2/24.8	←
Tobramycin	91.7/8.3	76.9/23.1	←
Levofloxacin	59.1/29.5	53.8/36.6	←
Colistin	98.9/1.1	99.0/1.0	

TZP: piperacillin/tazobactam

Sader, et al. Int J Antimicrob Agents. 2014;43:328–34

# Gram-negatives in Europe and Mediterranean Area

International Journal of Antimicrobial Agents 43 (2014) 328–334



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Organism/antimicrobial agent	%S/%R (no. tested)	
	EUCAST <sup>a</sup>	
	USA	EMR
<b>Klebsiella spp.</b>	(666) <sup>b</sup>	(695) <sup>c</sup>
TZP	77.6/15.6	64.7/29.4 ←
Ceftriaxone	81.4/18.2	67.6/31.7 ←
Ceftazidime	82.0/16.4	68.9/27.9 ←
Cefepime	83.8/13.2	71.7/24.6 ←
Meropenem	93.1/5.9	93.1/5.0
Amikacin	90.8/8.0	89.0/5.2
Gentamicin	89.3/9.0	81.2/17.7 ←
Levofloxacin	83.6/15.3	74.0/24.3 ←
Tigecycline <sup>d</sup>	93.5/2.1	94.4/0.7
Colistin	97.3/2.7	96.4/3.6

TZP: piperacillin/tazobactam

Sader, et al. Int J Antimicrob Agents. 2014;43:328–34

# Gram-negatives in Europe and Mediterranean Area

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International  
journal homepage

Antimicrobial susceptibility of Gram-negative bacterial organisms isolated from patients hospitalised with pneumonia in US and Europe and the Mediterranean region (EMR) medical centres (2009–2012). Results from the SENTRY Program, 2009–2012

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JMI Laboratories, 345 Beaver Creek Center, Suite A, North Liberty, IA, USA

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Organism/antimicrobial agent	%S/%R (no. tested)	
	EUCAST <sup>a</sup>	
	USA	EMR
<b>Escherichia coli</b>	(375)	(705)
TZP	87.2/9.6	81.1/12.9
 Ceftriaxone	83.7/16.0	81.8/17.7
 Ceftazidime	83.2/12.5	82.3/11.9
 Cefepime	85.1/11.2	83.4/13.1
 Meropenem	99.5/0.0	100.0/0.0
 Amikacin	97.9/0.5	95.7/1.4
 Gentamicin	81.6/15.8	84.0/15.0
 Levofloxacin	61.1/38.9	67.1/32.8
 Tigecycline <sup>d</sup>	100.0/0.0	99.9/0.0
 Colistin	99.7/0.3	99.9/0.1



TZP: piperacillin/tazobactam

# Gram-negatives in Europe and Mediterranean Area



Intern  
journal

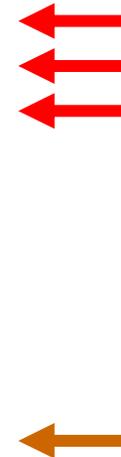
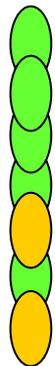
Antimicrobial susceptibil  
from patients hospitalis  
hospitals: Results from  
Program, 2009–2012

Helio S. Sader\*, David J. Farrell

JMI Laboratories, 345 Beaver Creek Center, Suite A,

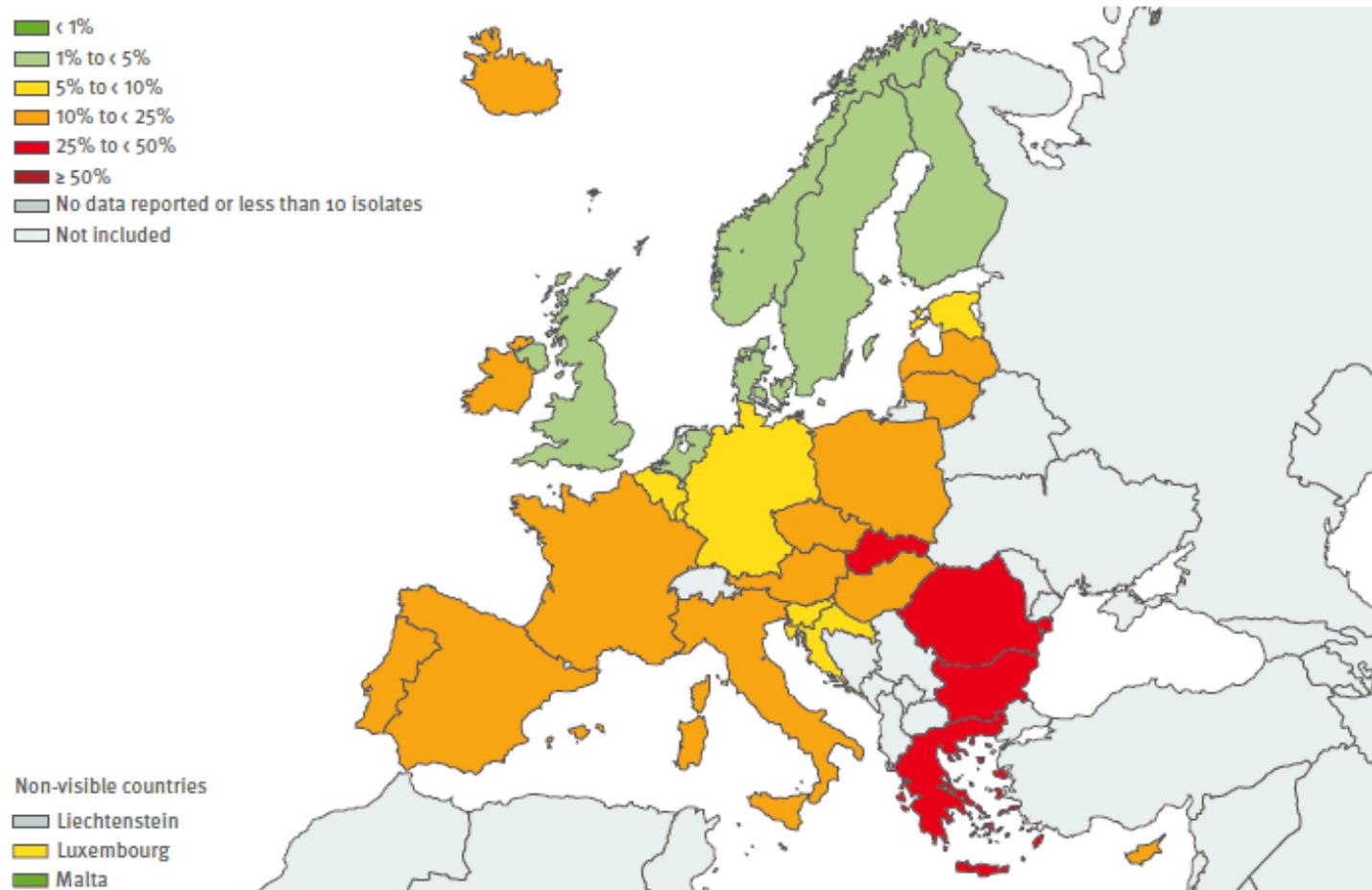
Antimicrobial susceptibility of Gram-negative bacterial organisms isolated from patients hospitalised with pneumonia in US and Europe and the Mediterranean region (EMR) medical centres (2009–2012).

Organism/antimicrobial agent	%S/%R (no. tested)	
	EUCAST <sup>a</sup>	
	USA	EMR
<b>Enterobacter spp.</b>	(407) <sup>f</sup>	(330) <sup>g</sup>
TZP	74.0/20.6	67.0/28.2
Ceftriaxone	69.0/29.1	60.6/35.8
Ceftazidime	69.6/26.9	61.2/32.7
Cefepime	86.7/4.7	85.5/4.8
Meropenem	99.0/0.5	99.1/0.3
Amikacin	99.5/0.0	97.6/1.5
Gentamicin	94.1/5.2	92.7/6.4
Levofloxacin	93.4/4.9	89.7/8.2
Tigecycline <sup>d</sup>	95.6/0.7	96.4/1.2
Colistin	86.4/13.6	85.0/15.0



# The problem of multiresistance: *P.aeruginosa* as an example

**Figure 3.25.** *Pseudomonas aeruginosa*. Percentage (%) of invasive isolates with combined resistance (resistance to three or more antimicrobial classes among piperacillin (± tazobactam), ceftazidime, fluoroquinolones, aminoglycosides and carbapenems), by country, EU/EEA countries, 2012



European Centre for Disease Prevention and Control. Antimicrobial resistance surveillance in Europe 2012. Annual Report of the European Antimicrobial Resistance Surveillance Network (EARS-Net). Stockholm: ECDC; 2013. p.36

# Emergence of resistance during treatment

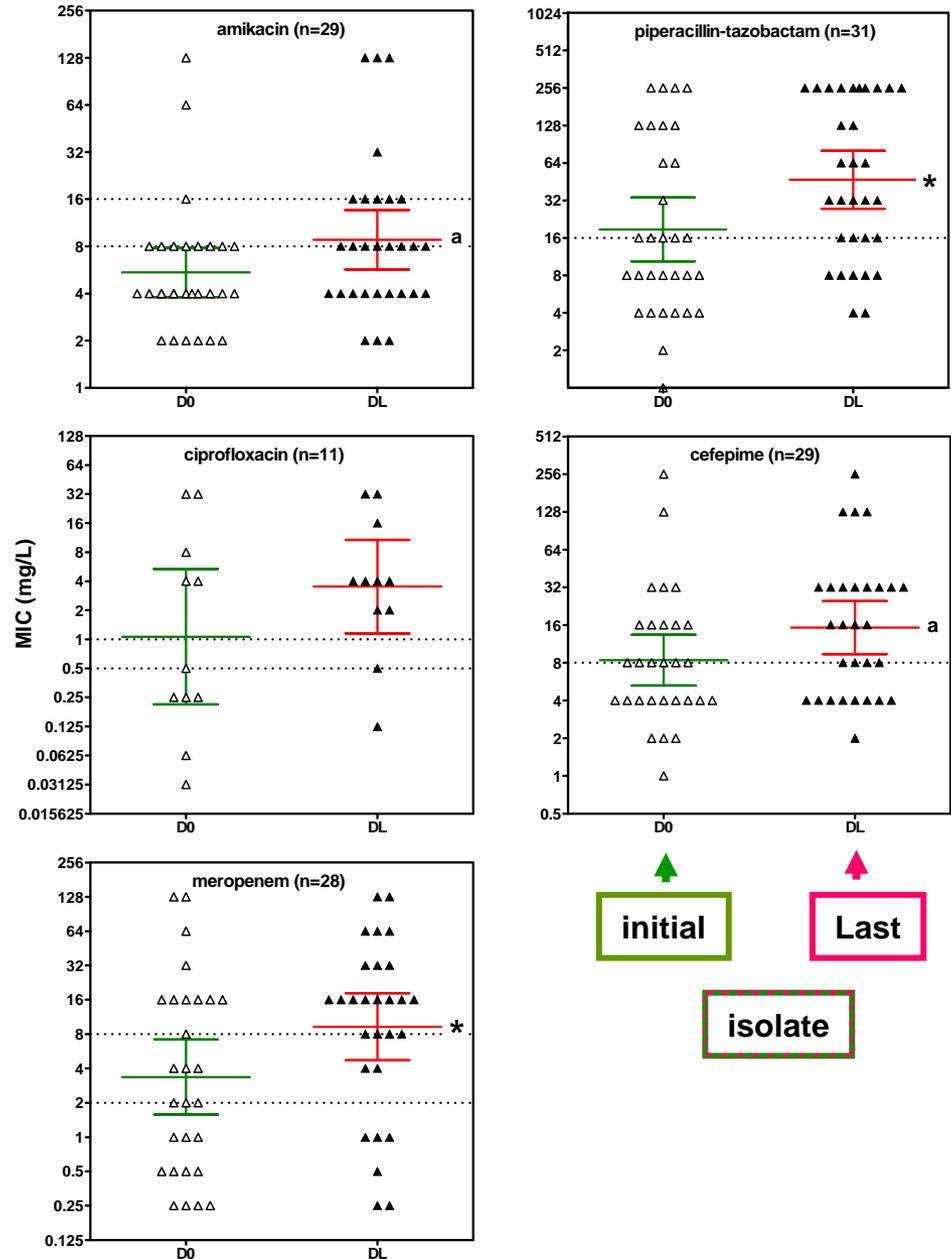
*P. aeruginosa* successive clonal isolates from the same patient (all patients treated with large doses of 1 to 3 antibiotics)

- D0: initial isolate
- DL: last isolate obtained
- individual values with geometric mean (95 % CI)
- S (lowest line) and R (highest line) EUCAST breakpoints

\*  $p < 0.05$  by paired t-test (two-tailed) and Wilcoxon non-parametric test

<sup>a</sup>  $p < 0.05$  by Wilcoxon non-parametric test only

Note: stratification by time between D0 and DL gave no clue (too low numbers)



# Emergence of resistance during treatment if persistent

## Original Article

<http://dx.doi.org/10.3947/ic.2013.45.3.283>  
Infect Chemother 2013;45(3):283-291  
pISSN 2093-2340 · eISSN 2092-6448



## Correlations between Microbiological Outcomes and Clinical Responses in Patients with Severe Pneumonia

Sungmin Kiem<sup>1</sup>, and Jerome J. Schentag<sup>2</sup>

<sup>1</sup>Department of Internal Medicine, Inje University College of Medicine, Busan, Korea; <sup>2</sup>School of Pharmacy and Pharmaceutical Sciences The University at Buffalo, Buffalo, NY, USA

- **3 clinical trials (US – 1984-1993) with PK/PD optimized dosages**
- **146 bacterial strains from 76 patients**
- **non-eradicated strains (71%) already had or developed resistance**

# Emergence of resistance during treatment if persistent ore relapse

## Original Article

<http://dx.doi.org/10.3947/ic.2013.45.3.283>  
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## Correlations between Microbiological Outcomes and Clinical Responses in Patients with Severe Pneumonia

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- 3 clinical trials (US – 1984-1993) with PK/PD optimized dosages
- 146 bacterial strains from 76 patients
- non-eradicated strains (71%) already had or developed resistance

Microbiological outcomes	Susceptible	Resistant	Development of resistance	Total
<i>Enterobacter spp.</i> <sup>a</sup>				12
Eradication	4	0	1	5
Persistence	0	0	3	3
Relapse	0	0	2	2
Colonization	0	2	0	2
<i>Pseudomonas spp.</i> <sup>d</sup>				31
Eradication	7	1	0	8
Persistence	4	4	9	17
Relapse	1	0	4	5
Colonization	1	0	0	1

# What can we do ?

- **Carbapenems** ... but may be a risk factor <sup>1</sup> for carbapenemase and neither is better <sup>2</sup>
- **Ceftozolane** may help for *P. aeruginosa* (with tazobactam) <sup>3</sup>
- **Avibactam** may restore susceptibility to ceftazidime to a high proportion of Gram-negatives including *P. aeruginosa* <sup>4</sup>
- **Combining** antibiotics (based on checker board <sup>5</sup>) or associating of glycopeptides with colistin for  $\geq 5$  days <sup>6</sup> could help
- **Extended infusion** (of cefepime) may improve mortality, and decrease mean length of stay and hospital costs <sup>7</sup>
- **Continuous infusion** may be a promising approach <sup>8</sup> ... but may not solve the problem of emergence of resistance... (see next slide)

1. Kim, *et al. Diag Microbiol Infect Dis.* 2014;78:457–61
2. Luyt, *et al. AAC.* 2014;58:1372-80.
3. Zhanel, *et al. Drugs.* 2014;74:31–51
4. Flamm, *et al. JAC.* 2014; Advance Access  
Chalhoub *et al* ECCMID 2014; e-poster 440
5. Nakamura, *et al. J Infect Chemother.* 2014;20:266e269
6. Petrosillo, *et al. AAC.* 2014;58:851-8
7. Bauer. *et al. AAC.* 2013;57:2907-12
8. Van Herendael, *et al. Ann Intensive Care.* 2012;2:22  
Dulhunty *et al. Clin Infect Dis.* 2013;56:236-44

# Bolus / Continuous infusion and resistance



Felton et al Antimicrob Agents Chemother 2013;57:5811-5819

## Impact of Bolus Dosing versus Continuous Infusion of Piperacillin and Tazobactam on the Development of Antimicrobial Resistance in *Pseudomonas aeruginosa*

T. W. Felton,<sup>a</sup> J. Goodwin,<sup>a,b</sup> L. O'Connor,<sup>a</sup> A. Sharp,<sup>a,b</sup> L. Gregson,<sup>a,b</sup> J. Livermore,<sup>a,b</sup> S. J. Howard,<sup>a,b</sup> M. N. Neely,<sup>c</sup> W. W. Hope<sup>a,b</sup>

The University of Manchester, Manchester Academic Health Science Centre, NIHR Translational Research Facility in Respiratory Medicine, University Hospital of South Manchester NHS Foundation Trust, Manchester, United Kingdom<sup>a</sup>; Antimicrobial Pharmacodynamics and Therapeutics, Department of Molecular and Clinical Pharmacology, University of Liverpool, Liverpool, United Kingdom<sup>b</sup>; Laboratory of Applied Pharmacokinetics, University of Southern California, School of Medicine, Los Angeles, California, USA<sup>c</sup>

# What do we need for efficacy ?



## Impact of Bolus and Tazobactam on *Pseudomonas*

T. W. Felton,<sup>a</sup> J. Goodwin,<sup>b</sup>  
The University of Manchester, M  
Manchester NHS Foundation Tru  
Pharmacology, University of Live  
Angeles, California, USA<sup>c</sup>

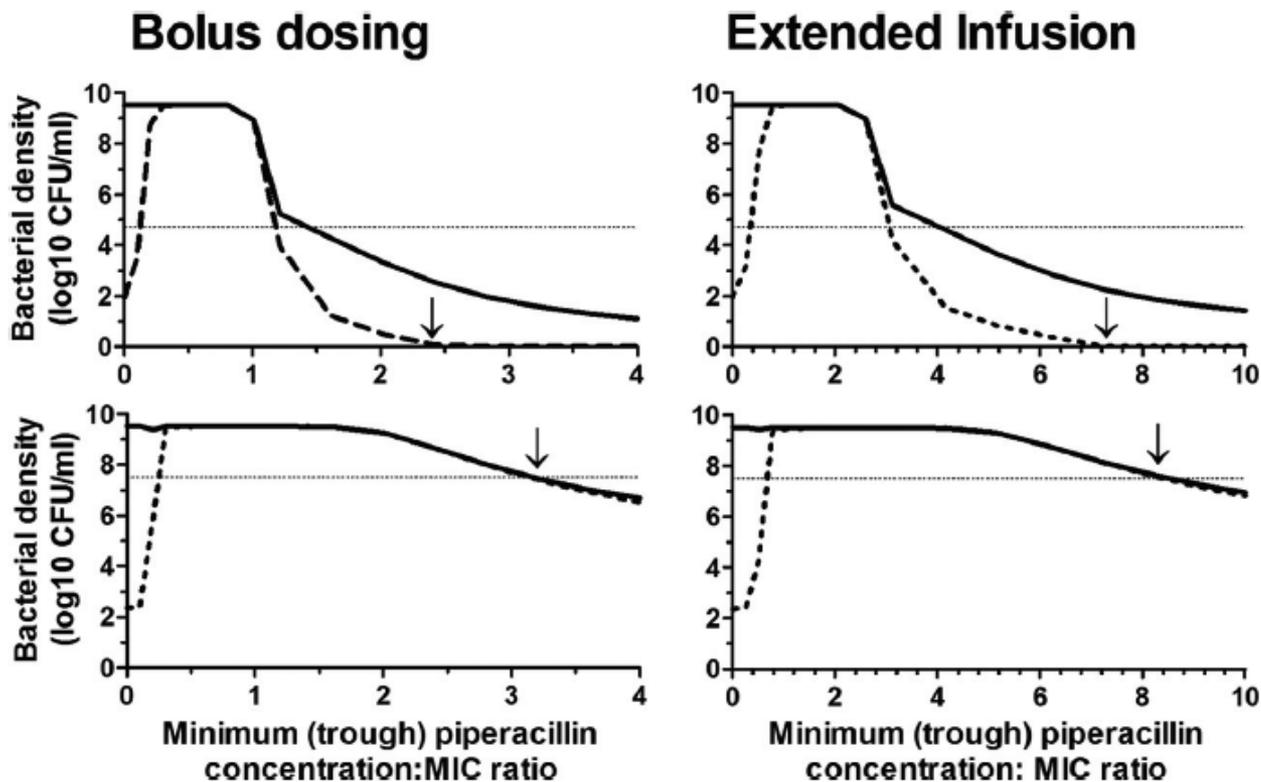


FIG 7 Change in bacterial density with the trough free piperacillin/MIC ratio following 5 days of treatment and target attainment of clinical regimens. Solid line, total population; dashed line, resistant subpopulation; dotted line, stasis line. The arrow indicates the relevant  $C_{\min}/MIC$  ratio.

# What do we need for suppression of resistance?



## Impact of Bolus Dosing and Tazobactam on the *Pseudomonas aeruginosa*

T. W. Felton,<sup>a</sup> J. Goodwin,<sup>a,b</sup> L. O'Connor,<sup>a</sup> A. ...  
 The University of Manchester, Manchester Academic Health Sciences Centre, Manchester NHS Foundation Trust, Manchester, United Kingdom  
 Pharmacology, University of Liverpool, Liverpool, United Kingdom  
 Los Angeles, California, USA<sup>c</sup>

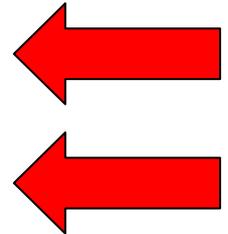
TABLE 3  $C_{\min}$ /MIC ratios required to achieve stasis, 1-, 2-, and 3-log bacterial killing and suppression of emergence of resistance

Bacterial density and status	$C_{\min}$ /MIC (mg/liter)			
	Bolus		Extended infusion	
	Hollow fiber	Predicted plasma <sup>a</sup>	Hollow fiber	Predicted plasma <sup>a</sup>
<b>Low</b>				
Bacterial stasis (total bacteria)	1.4	2.0	4.1	5.9
1-log reduction in total CFU/ml	1.8	2.6	5.2	7.4
2-log reduction in total CFU/ml	2.4	3.4	6.7	9.6
3-log reduction in total CFU/ml	3.2	4.6	8.8	12.6
Suppression of resistance	2.4	3.4	7.3	10.4
<b>High</b>				
Bacterial stasis (total bacteria)	3.2	4.6	8.3	11.9

<sup>a</sup> Protein binding is assumed to be 30% (31).

# Key questions to ask when **using** guidelines in infectious diseases (with application to pneumonia)

- How sure are you of the diagnosis ?
- **Which are the main pathogens ?**
- **What are their current resistance patterns and how can you avoid emergence of further resistance?**
- How should the therapy be initiated (empiric vs. directed) ?
- Which level of adverse effects is acceptable ?
- Which patients do you mainly treat?
- Does cost matter?
- What are your real choices?

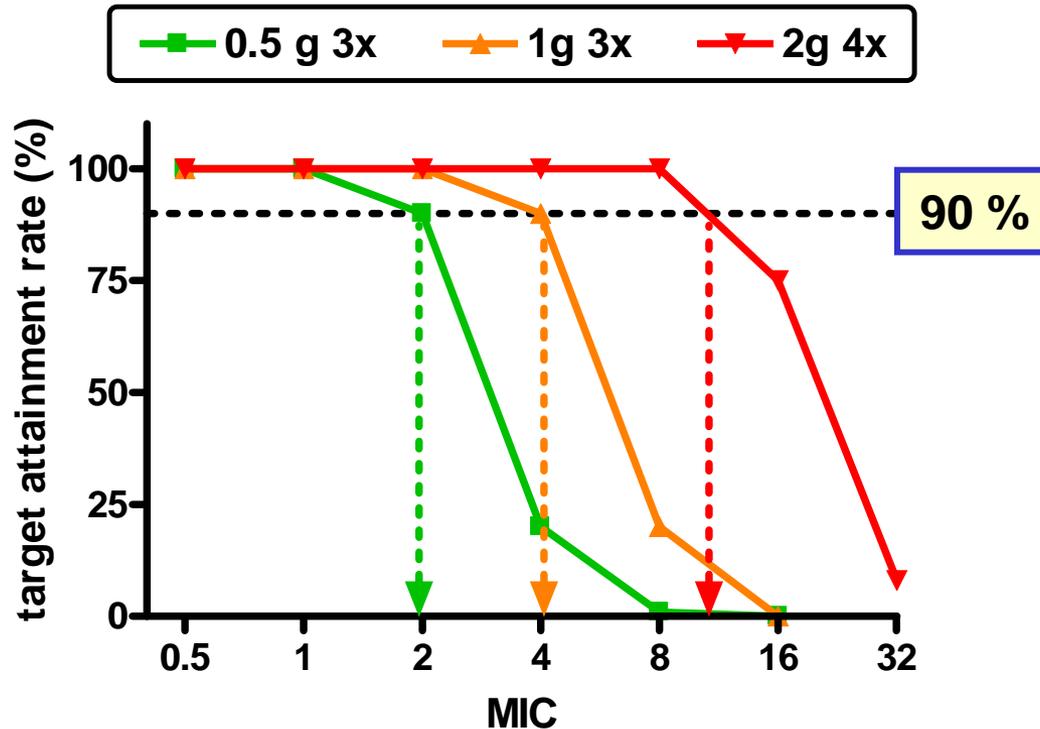


# What did I not speak about ... but should have done ... since it may impact your practice

- SCVs
- persisters and biofilms
- rapid diagnostic (including resistance phenotypes and mechanisms) and moving to personalized medicine
- TDM of  $\beta$ -lactams and fluoroquinolones
- new drugs
- pharmacoeconomy and approaches in case of limited resources...
- ....

# Back-up

# EUCAST calculations of target attainment rate for amoxicillin against *S. pneumoniae*



\* for  $fT > MIC = 40\%$

By increasing the dose and multiplying the number of daily administration, you may cover bacteria with MIC up to 8 mg/L... but the total daily dose will be very high and

Graph prepared from data in [http://www.eucast.org/fileadmin/src/media/PDFs/EUCAST\\_files/Rationale\\_documents/Amoxicillin\\_rationale\\_Nov2010\\_v\\_1.0.pdf](http://www.eucast.org/fileadmin/src/media/PDFs/EUCAST_files/Rationale_documents/Amoxicillin_rationale_Nov2010_v_1.0.pdf)

# Are macrolides still useful ?

- not as only agents if resistance rates > 20 % <sup>1</sup>
- but if used in combination with  $\beta$ -lactams to
  - act against organisms with low susceptibility to  $\beta$ -lactams (*Mycoplasma*, *Chlamydia*, *Legionella*) <sup>2</sup> when these are expected to be present and important (to be discussed)
  - to provide a so-called "antiinflammatory activity" (highly discussed <sup>3</sup>, but possible development with non-antibiotic derivatives [see next slide]).

1 a value often considered as being a critical threshold in a context of empirical therapy (Limond, *et al. Int J Antimicrob Agents*. 2012;39:208-16)

2 Baum: *Mycoplasma* and *Ureaplasma* / Stamm & Bateiger: *Chlamydia* and *Chlamydochila* /

Edelstein & Cianciotto: *Legionella* / In Mandell, Douglas, and Bennett's Principles and Practice of Infectious Diseases, 7<sup>th</sup> edition available on line at <https://expertconsult.inkling.com> (accessed: 4 April 2014)

3 Spagnolo, *et al. Eur Respir J*. 2013;42:239-51

# Anti-inflammatory action of "macrolides" ?

## RESEARCH PAPER

### Azithromycin analogue CSY0073 attenuates lung inflammation induced by LPS challenge

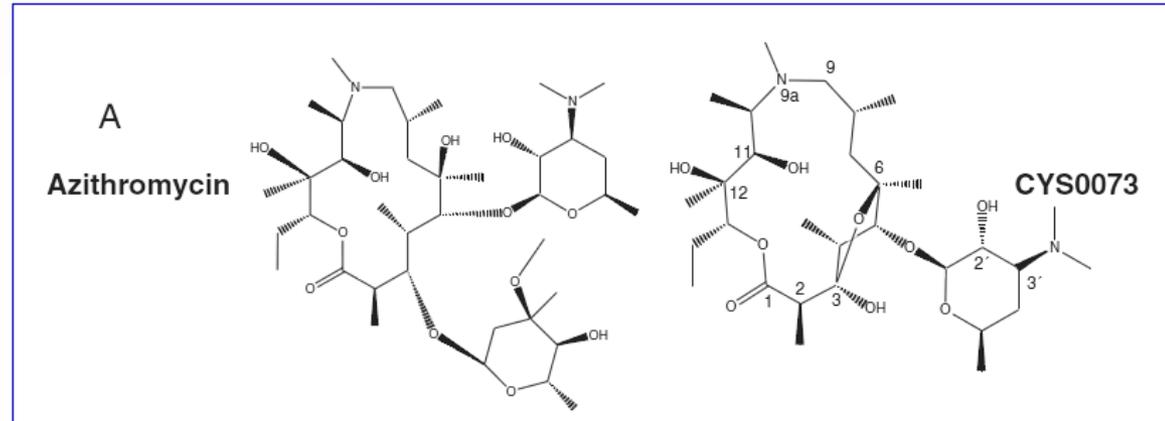
V Balloy<sup>1,2,3,4</sup>, A Deveaux<sup>1,2</sup>, D Lebeaux<sup>5</sup>, O Tabary<sup>1,2</sup>, P le Rouzic<sup>1,2</sup>,  
J M Ghigo<sup>5</sup>, P F Busson<sup>6,7</sup>, P Y Boëlle<sup>6,7</sup>, J Guez Guez<sup>8</sup>, U Hahn<sup>8</sup>,  
A Clement<sup>1,2,9</sup>, M Chignard<sup>3,4</sup>, H Corvol<sup>1,2,9</sup>, M Burnet<sup>8</sup> and L Guillot<sup>1,2</sup>

<sup>1</sup>INSERM, UMR\_S 938, CDR Saint-Antoine, Paris, France, <sup>2</sup>Sorbonne Universités, UPMC Univ Paris 06, UMR\_S 938, CDR Saint-Antoine, Paris, France, <sup>3</sup>Inserm U874, Paris, France, <sup>4</sup>Unité de défense Innée et Inflammation, Institut Pasteur, Paris, France, <sup>5</sup>Unité de Génétique des Biofilms, Institut Pasteur, Paris, France, <sup>6</sup>INSERM, UMR\_S 707, Paris, France, <sup>7</sup>Sorbonne Universités, UPMC Univ Paris 06, UMR\_S 707, Paris, France, <sup>8</sup>Synovo, Tübingen, Germany, and <sup>9</sup>Pneumologie pédiatrique, APHP, Hôpital Trousseau, Paris, France

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**Keywords**  
macrolides; lung inflammation;  
cystic fibrosis; COPD

**Received**  
5 November 2013  
**Revised**  
16 December 2013  
**Accepted**  
7 January 2014



# Global Resistance of *S. pneumoniae*: additional information

- Resistance to  $\beta$ -lactams and macrolides may be higher in children<sup>1</sup>
- Global resistance rates in Asia may be worse than currently reported
  - Erythromycin: > 70% of clinical isolates resistant<sup>2</sup>
  - High prevalence of penicillin resistance if using “old” CLSI or EUCAST breakpoints<sup>3</sup>

1. Diekema, *et al. Int. JAC.* 2002;20:412-8 / Brown & Farrell. *JAC.* 2004;54 Suppl 1:i23-9.2 / Hoban, *et al. Int. J. Infect. Dis.* 2005; 262-273 / Sanchez *et al. Rev. Esp. Quimioter.* 2007;20:421-8 / Lee *et al Int J Antimicrob Agents.* 2013;42:395-402.
2. Jean & Hsueh. *Int J Antimicrob Agents.* 2011;37:291-5 / Nickerson *et al Lancet Infect Dis* 2009;9:130-5.
3. Song, *et al. Clin Infect Dis* 1999;28:1206-11 / Song *et al Antimicrob Agents Chemother* 2004;48:2101-7 / Mendes *et al Antimicrob Agents Chemother.* 2013;57:5721-6.

# Global Resistance of *S. pneumoniae*: additional information

- Resistance to  $\beta$ -lactams and macrolides may be higher in certain regions
- Global trends in current prescribing

**TABLE 2** Comparative antimicrobial activities of selected agents tested against key Gram-positive pathogens for the APAC region RRS program (2011)

Organism (no. of strains tested) and antimicrobial agent	MIC ( $\mu\text{g/ml}$ )			% S/% R <sup>a</sup>	
	50%	90%	Range	CLSI	EUCAST
<i>S. pneumoniae</i> (42) <sup>e</sup>					
Penicillin <sup>f</sup>	$\leq 0.06$	4	$\leq 0.06$ –8	76.2/4.8	66.7/23.8
Amoxicillin-clavulanate	$\leq 1$	8	$\leq 1$ –>8	76.2/21.4	—/—
Ceftriaxone	$\leq 0.06$	8	$\leq 0.06$ –8	78.6/14.3	66.7/14.3
Clindamycin	$\leq 0.25$	>2	$\leq 0.25$ –>2	50.0/50.0	50.0/50.0
Erythromycin	1	>16	$\leq 0.12$ –>16	47.6/52.4	47.6/52.4
Levofloxacin	1	1	0.5–2	100.0/0.0	100.0/0.0

APAC: Asisa/Pacific [Australia, Hong Kong, India, Indonesia, Japan, South Korea, Malaysia, New Zealand, Philippines, Singapore, Taiwan, Thailand

RRS: Regional Resistance Surveillance programme

1. Diekema, *et al. Int. JAC.* 2002;20:412-8 / Brown & Farrell. *JAC.* 2004;54 Suppl 1:i23-9.2 / Hoban, *et al. Int. J. Infect. Dis.* 2005; 262-273 / Sanchez *et al. Rev. Esp. Quimioter.* 2007;20:421-8 / Lee *et al Int J Antimicrob Agents.* 2013;42:395-402.
2. Jean & Hsueh. *Int J Antimicrob Agents.* 2011;37:291-5 / Nickerson *et al Lancet Infect Dis* 2009;9:130-5.
3. Song, *et al. Clin Infect Dis* 1999;28:1206-11 / Song *et al Antimicrob Agents Chemother* 2004;48:2101-7 / Mendes *et al Antimicrob Agents Chemother.* 2013;57:5721-6.

# Resistance of *S. pneumoniae* to fluoroquinolones

- Several countries noted no or little resistance over time if used appropriately even with relatively large use

## Example #1: Canada

**Table 1.** *In vitro* activities of fluoroquinolones against selected pathogens in the CANWARD study 2007–11 as well as prevalence of MDR isolates involving fluoroquinolone over time

Organism/resistance phenotype	Year				
	2007	2008	2009	2010	2011
<i>S. pneumoniae</i>					
% R <sup>a</sup> levofloxacin (no. of isolates tested)	0.3 (591)	1.5 (514)	0 (129)	0.6 (168)	0 (138)
% MDR <sup>b</sup>	3.2	6.8	3.9	7.1	7.2
% MDR isolates resistant to levofloxacin	5.3	11.4	0	8.3	0

<sup>a</sup>R includes both intermediate and resistant isolates.

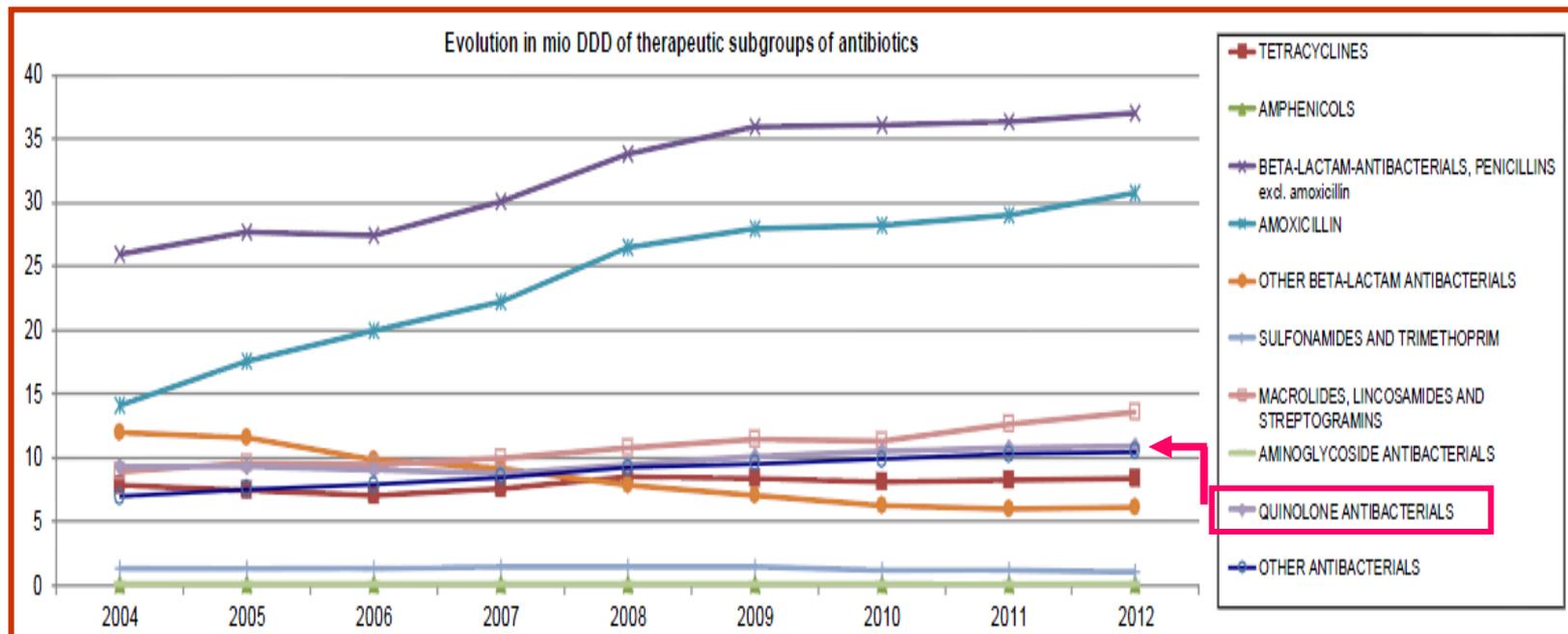
<sup>b</sup>MDR includes both intermediate and resistant isolates for the following agents for the following organisms: *E. coli*, *K. pneumoniae* and *E. cloacae* (ciprofloxacin, ceftriaxone, meropenem, piperacillin/tazobactam, trimethoprim/sulfamethoxazole and gentamicin); *P. aeruginosa* (ciprofloxacin, ceftazidime, meropenem, piperacillin/tazobactam and gentamicin); *S. aureus* (ciprofloxacin, clarithromycin, oxacillin and trimethoprim/sulfamethoxazole); and *S. pneumoniae* (levofloxacin, penicillin, clarithromycin and trimethoprim/sulfamethoxazole).

# Resistance of *S. pneumoniae* to fluoroquinolones

- Several countries noted no or little resistance over time if used appropriately even in relatively large use

## Example #2: Belgium

Antibiotics used in the ambulatory care in Belgium (reimbursement data [ >95% of total use])



Source: Belgian National Institute for Sickness and Invalidation Insurance: "Tableaux de bord pharmaceutiques: Délivrances pharmaceutiques dans le secteur ambulatoire – année 2012"

<http://www.inami.be/drug/fr/statistics-scientific-information/pharmanet/pharmaceutical-tables/pdf/2012/tables2012.pdf>

Last accessed: 20/01/2014

# A comparison of three CAP guidelines separated by (some) water



Table 4. Recommended community-acquired pneumonia therapy and management from published international guidelines \*

	BTS guidelines [24]	ATS/IDSA guidelines [25]	ERS/ESCMID guidelines [26]
Low severity patients*	<p>Use CURB65 score with clinical judgement</p> <p>Treat with oral amoxicillin or (doxycycline or clarithromycin if hypersensitive).</p>	<p>Use CURB65 or PSI score to guide Outpatient treatment</p> <p>Stratify by risk for drug resistant <i>S. pneumoniae</i></p> <p>Low risk: Treat with macrolide or doxycycline</p> <p>High risk: Treat with respiratory fluoroquinolone or b-lactam+macrolide</p>	<p>Use CRB65 to guide Outpatient treatment</p> <p>Treat with one of: aminopenicillin ± macrolide</p> <p>Aminopenicillin/b-lactamase inhibitor ± macrolide</p> <p>Non-antipseudomonal cephalosporin</p> <p>Cefotaxime or ceftriaxone ± macrolide</p> <p>Levofloxacin</p> <p>Moxifloxacin</p> <p>Penicillin g ± macrolide</p>

\*These are not necessarily the terms used in the guidelines but give a broad translation of what the guidelines state.

Khan & Woodhead, F1000 Prime Rep. 2013;;5:43

Free access: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC3790563/pdf/medrep-05-43.pdf>

Note: *S. pneumoniae* is (probably) the most frequent isolated organism in CAP (~ 20 %), but others may need to be considered (*Mycoplasma pneumoniae* ~ 11 %; *Chlamydia pneumoniae* ~ 8.0 %; *Haemophilus influenzae* ~ 3.3 %), and ~ 50% of cases remain without successful isolation

(Woodhead. Eur Respir J 2002; 36:20s-27s)

# *S. aureus* in Asia: VISA and hVISA

- VRSA and true VISA are rare<sup>1</sup>
- hVISA phenotype is much more frequent
  - 1/3 of MRSA isolates in Korea and was independently associated with a vancomycin MIC  $\geq 2$  mg/L and rifampicin resistance<sup>2</sup>
- VISA and hVISA are associated with a longer period of prior glycopeptide use, bone/joint and prosthetic infections, and treatment failure<sup>3</sup>

hVISA heterogeneous vancomycin-intermediate-resistant *S. aureus* :

VISA: vancomycin-intermediate *S. aureus*

VRSA vancomycin-resistant *S. aureus*

MRSA methicillin-resistant *S. aureus*

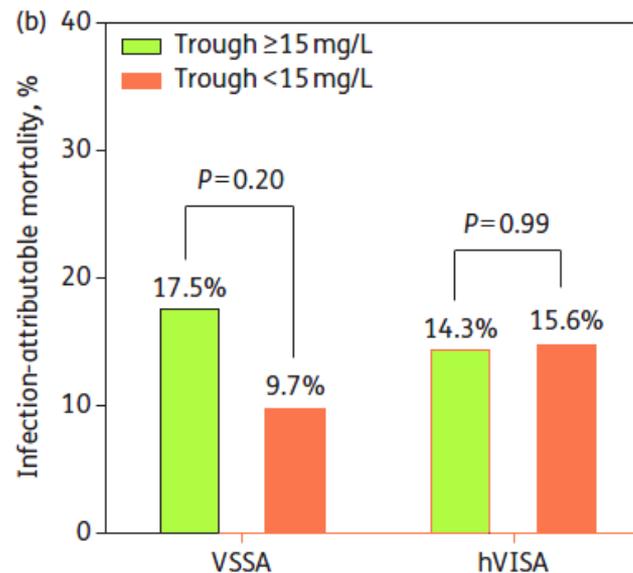
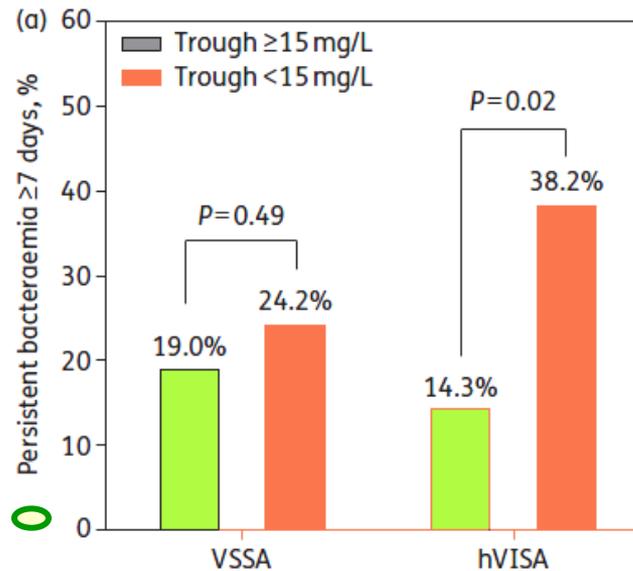
1. Kang & Song. *Infect Chemother.* 2013;45:22-31
2. Park, et al. *J Antimicrob Chemother.* 2012;67:1843-9
3. Fong, et al. *Eur J Clin Microbiol Infect Dis.* 2009;28:983-7

# S. Aureus in Asia:

- VRSA and true VISA are rare
- hVISA is much more common in Korea
- VISA and hVISA are associated with prior glycopeptide use, hospital-acquired infections, and treatment failures

higher trough levels may be necessary

1. Kang & Song *Infect Chemother* 2013;45:22-31
2. Park *et al* *J Antimicrob Chemother* 2012;67:1843-9.
3. Fong *et al* *Eur J Clin Microbiol Infect Dis* 2009;28:983-7.
4. Park *et al* *J Antimicrob Chemother.* 2012;67:1843-9.



**Figure 1.** Clinical outcomes of patients with hVISA and VSSA bacteraemia based on initial vancomycin trough levels.

# *S. aureus* in Africa

- Very little is known about Africa !
- But data that are coming are challenging...

Infection

DOI 10.1007/s15010-014-0589-1

REVIEW

## **Methicillin-resistant *Staphylococcus aureus* as a cause of invasive infections in Central Africa: a case report and review of the literature**

M. A. M. Huson · R. Kalkman · J. Remppis ·  
J. O. Beyeme · C. Kraef · F. Schaumburg ·  
A. S. Alabi · M. P. Grobusch

**Infection. 2014 Jan 25. [Epub ahead of print]**

# S. aureus in Africa

- Very little
- But data

**Table 1** Epidemiology of methicillin-resistant *Staphylococcus aureus* (MRSA) in Central Africa

Country <sup>a</sup>	References	Study period	Type of specimens	Setting	Total number of isolates screened	Total number of <i>S. aureus</i> isolates	MRSA isolates (%) <sup>b</sup>
Cameroon	Kesah et al. [30]	1996–1997	Clinical isolates: including surgical samples (swabs and pus), ear, nose, and throat samples, urine, sputum, cerebrospinal fluid, aspirates, and blood cultures	Urban, university hospital	Not indicated	127	27 (21.3)
	Breurec et al. [31]	January 2007–March 2008	Clinical isolates from patients with suspected staphylococcal infection	Urban, university hospital	Not indicated	32	9 (28.1)
Gabon	Schaumburg et al. [33]	2008–2010	Healthy volunteers: isolates from anterior nares, the axilla, and groin	Semi-urban	Not indicated	163	6 (3.7)
			Clinical isolates: mainly wound infection, bacteremia, and abscesses	Semi-urban, regional referral hospital	Not indicated	54	6 (11.1)
	Schaumburg et al. [36]	2009	Healthy volunteers: nasal carriage	Remote rural	100	34	0
	Ateba Ngoa et al. [32]	February–July 2009	Asymptomatic volunteers <sup>c</sup> : isolates from anterior nares, the axilla, and groin	Rural and semi-urban	500	146	5 (3.4)
	Alabi et al. [34]	January 2009–September 2012	Clinical isolates: mainly bloodstream, ear-eye-nose-throat, and skin and soft tissue infection	Semi-urban, regional referral hospital	Not indicated	328	19 (5.8)
	Schaumburg et al. [35]	March 2010–January 2013	Healthy mothers (nares, mamillae) and infants (nares, throat)	Rural	Not indicated	474	9 (1.9)

<sup>a</sup> No data were available for countries other than Cameroon and Gabon

<sup>b</sup> Percentage of the total *S. aureus* isolates

<sup>c</sup> 198 inpatients asymptomatic for *S. aureus*-related disease and 302 healthy volunteers

Infection  
DOI 10.1007/s15010-014-0589-1

REVIEW

## Methicillin-resistance infections in Central Africa: a review of the literature

M. A. M. Huson · R. Kall  
J. O. Beyeme · C. Kraef ·  
A. S. Alabi · M. P. Grob

Infection. 2014 Jan 2

# *S. aureus* in Africa

- Very little is known about African
- But data that are coming are

Infection

DOI 10.1007/s15010-014-0589-1

## REVIEW

### Methicillin-resistant *Staphylococcus aureus* as a cause of community-acquired pneumonia infections in Central Africa: a case report and review of the literature

M. A. M. Huson · R. Kalkman · J. Remppis ·  
J. O. Beyeme · C. Kraef · F. Schaumburg ·  
A. S. Alabi · M. P. Grobusch

Infection. 2014 Jan 25. [Epub ahead of print]

#### *S. aureus*

- resistant to penicillin, cloxacillin, ciprofloxacin, and erythromycin,
- sensitive to gentamicin, clindamycin, trimethoprim-sulfamethoxazole and rifampicin.

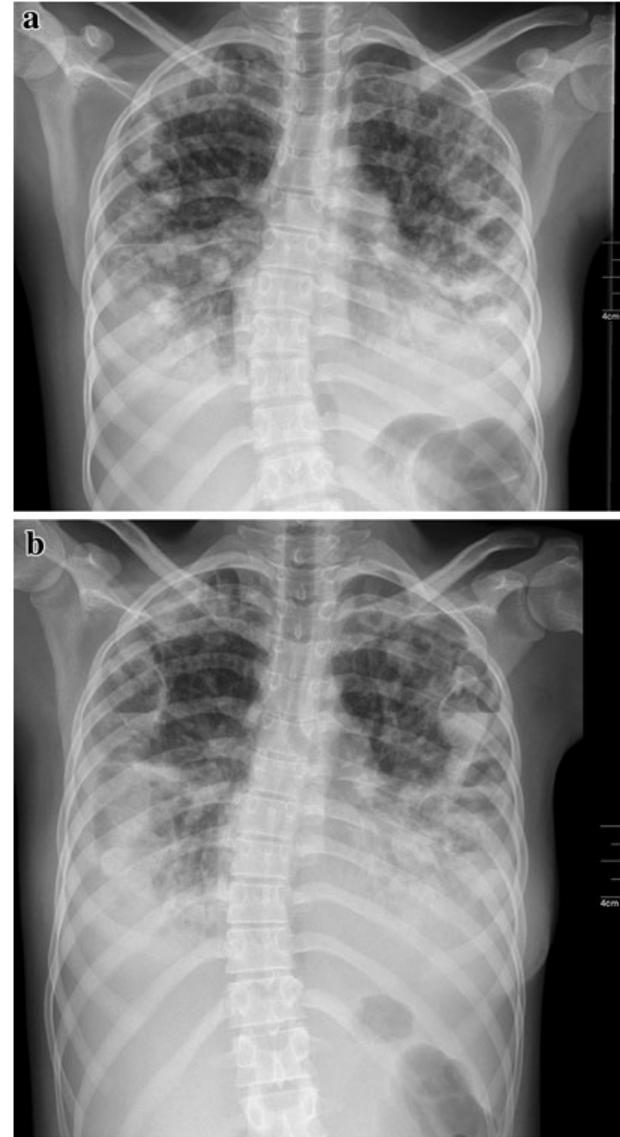


Fig. 1 a Chest X-ray at admission shows bilateral infiltrates and cavitary lesions. b Chest X-ray 6 days after admission demonstrates deterioration of cavitary lesions with air-fluid levels

# Anaerobes and pneumonia

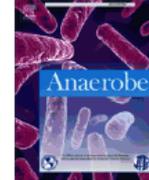
Anaerobe 18 (2012) 235–239



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Clinical microbiology

## Anaerobic bacterial infection of the lung

John G. Bartlett\*

*Johns Hopkins University, School of Medicine, 1830 East Monument Street, Rm 447, Baltimore, MD 21202, United States*

# Anaerobes and pneumonia

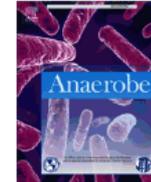
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Clinical microbiology  
Anaerobic infections  
John G. Bartlett  
Johns Hopkins University

**Table 2**  
Clinical features of pulmonary infections involving anaerobic bacteria<sup>a</sup>.

Number <sup>b</sup>	Pneumonitis 79	Abscess 83	Empyema 51	Total 213
Age (yrs)	60	52	49	51
Peak fever (°F)	102.6	102.1	102.4	102.4
WBC ( $\times 1000/\text{mL}$ )	13.7	15.0	21.6	15.0
Duration six (days)	3	14	15	7
Weight loss (Yes)	3%	43%	55%	30%
Putrid discharge	4%	49%	63%	32%
Mortality	4%	4%	6%	4%

<sup>a</sup> Categories are mutually exclusive.

<sup>b</sup> All figures are median values.

# What are the outcomes of MDR in VAP?

American Journal of Infection Control xxx (2014) 1-4



ELSEVIER

Contents lists available at [ScienceDirect](#)

American Journal of Infection Control

journal homepage: [www.ajicjournal.org](http://www.ajicjournal.org)



Brief report

## The impact of multidrug resistance on outcomes in ventilator-associated pneumonia

Rudy Tedja MD<sup>a</sup>, Amy Nowacki PhD<sup>b</sup>, Thomas Fraser MD<sup>a,c</sup>, Cynthia Fatica RN<sup>c</sup>,  
Lori Griffiths RN<sup>c</sup>, Steven Gordon MD<sup>a</sup>, Carlos Isada MD<sup>a</sup>, David van Duin MD, PhD<sup>d,\*</sup>

<sup>a</sup> Department of Infectious Diseases, Medicine Institute, Cleveland Clinic, Cleveland, OH

<sup>b</sup> Department of Quantitative Health Sciences, Cleveland Clinic, Cleveland, OH

<sup>c</sup> Department of Infection Prevention, Quality and Patient Safety Institute, Cleveland Clinic Foundation, Cleveland Clinic, Cleveland, OH

<sup>d</sup> Division of Infectious Diseases, University of North Carolina, Chapel Hill, NC

# What are the outcomes of MDR in VAP?

American Journal of Infection Control xxx (2014) 1-4



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America

jour

Brief report

The impact of multidrug resistant ventilator-associated pneumonia

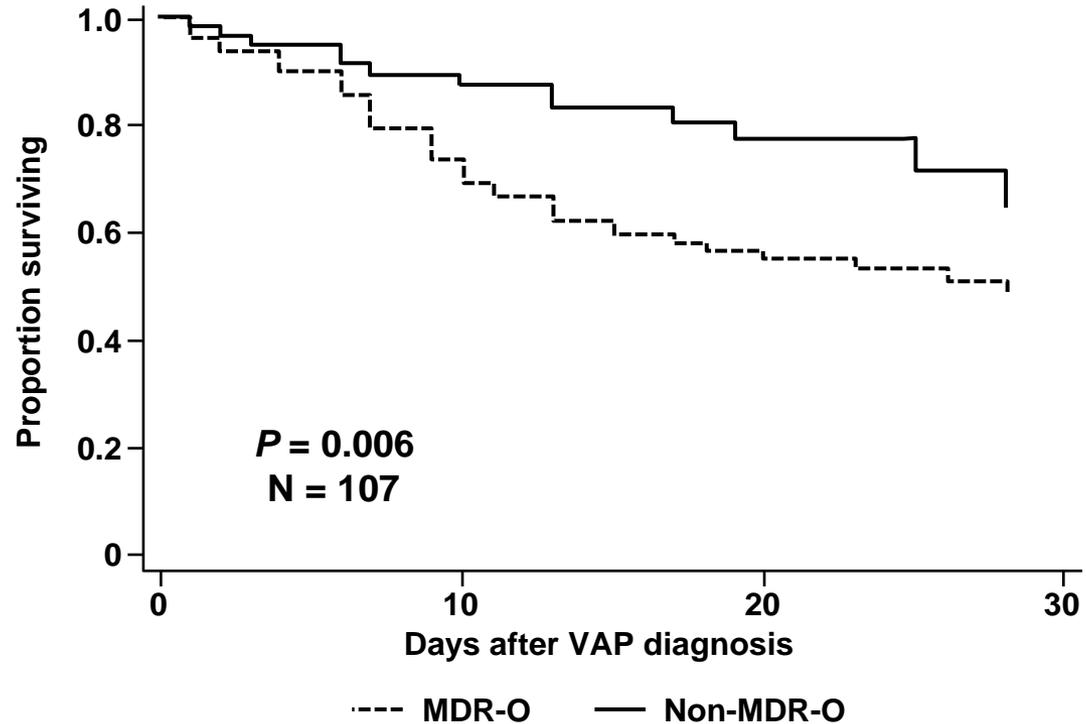
Rudy Tedja MD<sup>a</sup>, Amy Nowacki PhD<sup>b</sup>,  
Lori Griffiths RN<sup>c</sup>, Steven Gordon MD<sup>d</sup>

<sup>a</sup> Department of Infectious Diseases, Medicine Institute, Cleveland

<sup>b</sup> Department of Quantitative Health Sciences, Cleveland Clinic, C

<sup>c</sup> Department of Infection Prevention, Quality and Patient Safety

<sup>d</sup> Division of Infectious Diseases, University of North Carolina, CH



Survival after diagnosis of ventilator-associated pneumonia. Time to death is shown, censored by hospital discharge. NDR-O, multidrug-resistant organism.

Tedja, et al. *Am J Infect Control*. 2014;pii: S0196-6553(13)01428-4.

# What are the outcomes of MDR in VAP?

American Journal of Infection Control xxx (2014) 1-4



ELSEVIER

America

jour

Brief report

The impact of multidrug resistant ventilator-associated pneumonia

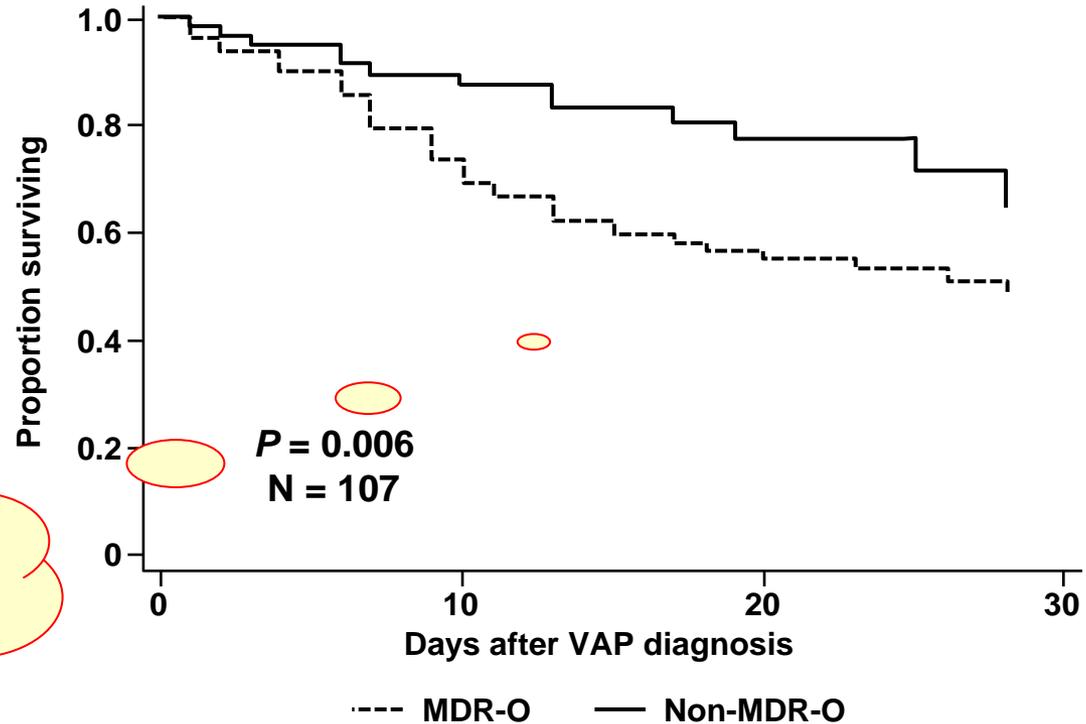
Rudy Tedja MD<sup>a</sup>, Amy Nowacki PhD<sup>b</sup>,  
Lori Griffiths RN<sup>c</sup>, Steven Gordon MD<sup>d</sup>

<sup>a</sup> Department of Infectious Diseases, Medicine Institute, Cleveland

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<sup>d</sup> Division of Infectious Diseases, University of North Carolina, CH



Survival after diagnosis of ventilator-associated pneumonia. Time to death is shown, censored by hospital discharge. NDR-O, multidrug-resistant organism.

I guess such a difference would command a very high price for an anticancer drug...

# What happens if you are inadequate...

Intensive Care Med (2013) 39:682–692  
DOI 10.1007/s00134-013-2828-9

ORIGINAL

## **Clinical outcomes of *Pseudomonas aeruginosa* pneumonia in intensive care unit patients**

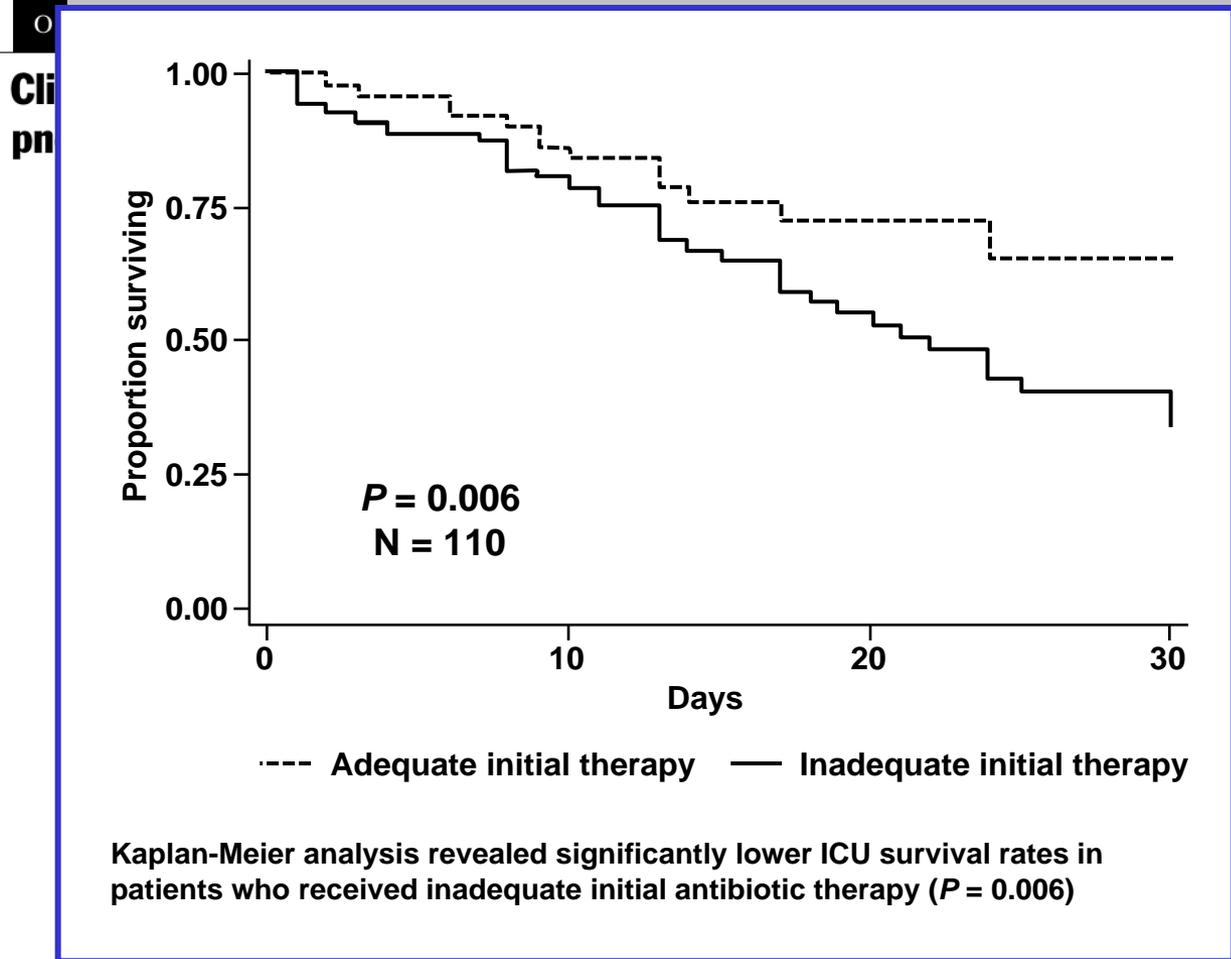
Mario Tumbarello  
Gennaro De Pascale  
Enrico Maria Trecarichi  
Teresa Spanu  
Federica Antonicelli  
Riccardo Maviglia  
Mariano Alberto Pennisi  
Giuseppe Bello  
Massimo Antonelli

Tumbarello, et al *Intensive Care Med.* 2013;39:682-92

# What happens if you are inadequate...

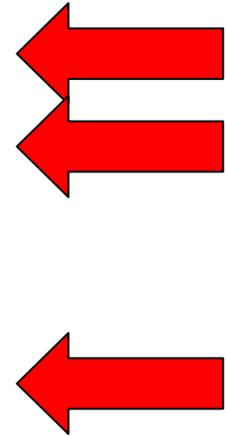
Intensive Care Med (2013) 39:682–692  
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Enrico Maria Treçarichi  
Teresa Spanu  
Federica Antonicelli  
Riccardo Maviglia  
Mariano Alberto Pennisi  
Giuseppe Bello  
Massimo Antonelli



# Key questions to ask when **setting** guidelines in infectious diseases (with application to pneumonia)

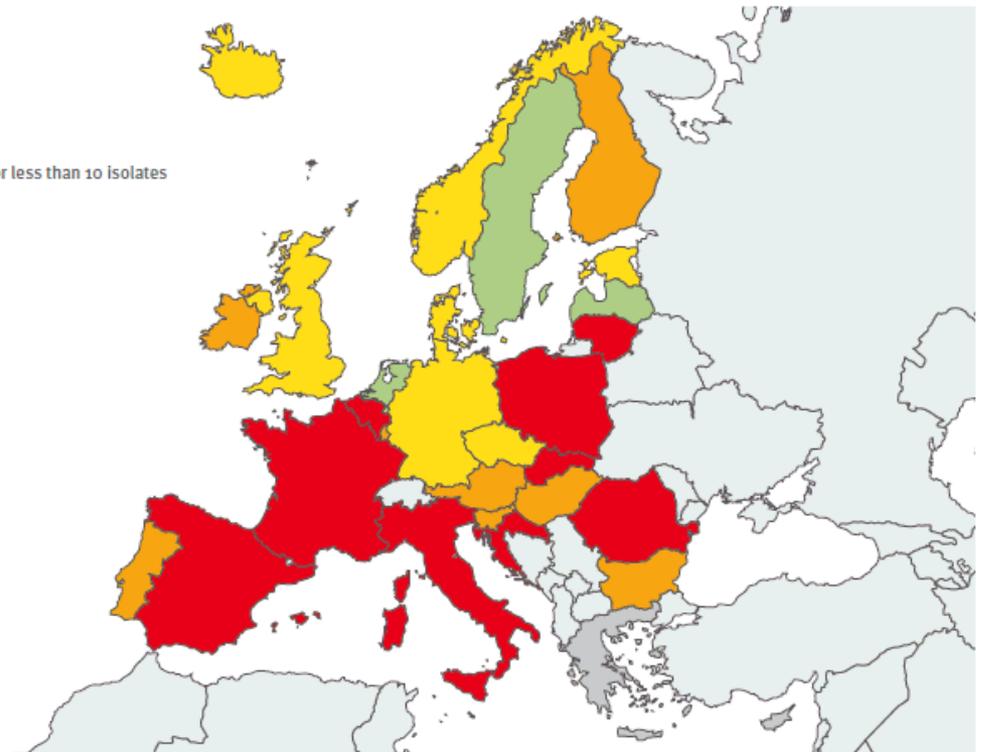
- How sure are you of the diagnosis ?
- **Which are the main pathogens ?**
- **What are their current resistance patterns ?**
- How should the therapy be initiated (empiric vs. directed) ?
- **Which level of adverse effects is acceptable ?**
- Which patients do you mainly treat?
- Does cost matter?
- What are your real choices?



# Some potential approaches

- **Community-acquired pneumonia**
  - local (country, region...) data on resistance (example: macrolides and *S. pneumoniae*)

Figure 3.37. *Streptococcus pneumoniae*. Percentage (%) of invasive isolates non-susceptible to macrolides by country, EU/EEA countries, 2012



European Centre for Disease Prevention and Control. Antimicrobial resistance surveillance in Europe 2012. Annual Report of the European Antimicrobial Resistance Surveillance Network (EARS-Net). Stockholm: ECDC; 2013. p. 52

# Some potential approaches

- **Community-acquired pneumonia**
  - local (country, region...) data on resistance (example: macrolides and *S. pneumoniae*)
  - **stratification for occurrence of resistant pathogens (or with decreased susceptibility [ $\beta$ -lactams])**

# Some potential approaches

- **Community-acquired pneumonia**

- local (country, region...) data on resistance (example: macrolides and *S. pneumoniae*)

- stratification for occurrence of resistant pathogens (or with decreased sensitivity)

**Table 2.** Scoring System to Evaluate Presence of MDR Pathogens in Hospitalized Patients With CAP

Variable	Score
No risk factors for MDR pathogen (including comorbidities)	0
≥ 1 of the following: cerebrovascular disease; diabetes; COPD; antimicrobial therapy in preceding 90 days; immunosuppression; home wound care; home infusion therapy (including antibiotics)	0.5
Residence in a nursing home or extended-care facility	3
Hospitalization for ≥ 2 days in preceding 90 days	4
Chronic renal failure	5

Adapted from Aliberti S, Di Pasquale M, Zanaboni AM, et al. Stratifying risk factors for multidrug-resistant pathogens in hospitalized patients coming from the community with pneumonia. *Clin Infect Dis.* 2012;54(4):470–478.<sup>12</sup> © 2011; Oxford University Press. Used with permission.

**Abbreviations:** CAP, community-acquired pneumonia; COPD, chronic obstructive pulmonary disease; MDR, multidrug-resistant.

# Some potential approaches

- **Community-acquired pneumonia**

- local (country, region...) data on resistance (example: macrolides and *S. pneumoniae*)
- stratification for occurrence of resistant pathogens (or with decreased susceptibility [ $\beta$ -lactams])
- **scoring of severity and potential for Gram-negatives**



Table 4. Recommended community-acquired pneumonia therapy and management from published international guidelines \*

	BTS guidelines [24]	ATS/IDSA guidelines [25]	ERS/ESCMID guidelines [26]
<b>Moderate/high severity patients*</b>	<p><u>CURB65 score 3 or more</u> consider ICU</p> <p>Treat with <math>\beta</math>-lactam plus macrolide iv</p>	<p>Consider ICU for <u>sepsis or &gt;2 minor severity criteria</u></p> <p>Increased Comorbidities or prior antimicrobials (within 3 months) treat with respiratory fluoroquinolone or beta lactam plus macrolide iv</p>	<p>Consider ICU for <u>respiratory failure or sepsis or &gt;2 minor severity criteria</u></p> <p>Stratify by risk for <i>Pseudomonas aeruginosa</i> Non-antipseudomonal treat with cephalosporin III + macrolide Or Moxifloxacin or levofloxacin <math>\pm</math> non-antipseudomonal cephalosporin III</p>

\*These are not necessarily the terms used in the guidelines but give a broad translation of what the guidelines state.

# Some potential approaches

- **Community-acquired pneumonia**

- local (country, region...) data on resistance (example: macrolides and *S. pneumoniae*)
- stratification for occurrence of resistant pathogens (or with decreased susceptibility [ $\beta$ -lactams])
- scoring of severity and potential for Gram-negatives
- **presence of *S. aureus* (MSSA / MRSA)**

Type of infection	MSSA	MRSA	days
acute / non PVL	<ul style="list-style-type: none"> <li>• BLRP (150 mg/kg) <sup>a</sup></li> <li>• amoxyclav (2-6g)</li> <li>• clindamycin <sup>c</sup></li> </ul>	<ul style="list-style-type: none"> <li>• vancomycin (30mg/kg) <sup>b</sup></li> <li>• teicoplanin (400-600 mg/day)</li> <li>• clindamycin <sup>c</sup></li> <li>• pristinamycin</li> </ul>	4-7
necrosis / PVL +	<ul style="list-style-type: none"> <li>• BLRP (150-200 mg/kg) <sup>a</sup></li> <li>+ clindamycin <sup>c</sup></li> </ul>	<ul style="list-style-type: none"> <li>• vancomycin (30-40 mg/kg <sup>b</sup> or continuous infusion)</li> <li>+ clindamycin <sup>c</sup></li> <li>• linezolid (1.2g q12h) *</li> <li>• ceftaroline (1.2g) + clindamycin <sup>c</sup></li> <li>*</li> </ul>	≥ 14
<sup>a</sup> $\beta$ -lactamase-resistant penicillin (in 3 or 4 administrations/day (q8h or Q6h)) ; <sup>b</sup> in 2 administrations per day (q12h) ; <sup>c</sup> 1.8 to 2.4 g/day in 3-4 administration per day (q8h or q6h); * non-validated (off-label)			

Translated from Valour *et al* Rev Pneumol Clin. 2013;69:368-82

# Some potential approaches

- **Hospital acquired pneumonia (including VAP)**
  - preventive measures (VAP)

**Table 1. Effects of the main preventive measures for ventilator-associated pneumonia prevention in randomized controlled studies or last meta-analyses**

Intervention	Year, design	Patients (n)	RRR of VAP (%)
<b>Reducing the time at risk</b>			
NPPV	2005, meta-analysis (12 studies)	3030	↓ 25%
<b>Preventing endotracheal tube colonization and minimizing contaminated microaspirations</b>			
Silver-coated ET	2008, RCT (54 ICUs)	1509	↓ 36%
Saline instillation before tracheal suctioning	2009, RCT (one ICU)	262	↓ 54%
ET with SSD	2010, meta-analysis (13 RCTs)	2442	↓ 48% (four RCTs)
Endotracheal tube with ultrathin membrane and SSD	2007, RCT (one ICU)	280	↓ 64%
Endotracheal tube with an ultrathin and tapered-shape cuff membrane	2008, RCT (one ICU)	134	↓ 45%
Continuous control of tracheal cuff pressure	2011, RCT (one ICU)	122	↓ 63%
Head-of-bed elevation	1999, RCT (two ICUs)	86	↓ 78%
Kinetic beds	2006, meta-analysis (15 RCTs)	1169	↓ 53% (10 RCTs)
Positive end-expiratory pressure	2008, RCT (three ICUs)	131	↓ 63%
<b>Modulation of colonization</b>			
Oral care with chlorhexidine	2010, meta-analysis (12 RCTs)	2341	↓ 24%
Probiotics	2010, RCT (one ICU)	146	↓ 47%

Data from [17]. ET, endotracheal tube; NPPV, noninvasive positive pressure ventilation; RCT, randomized controlled trial; RRR, relative risk reduction; SBT spontaneous breathing trial; SSD, subglottic secretion drainage; VAP, ventilator-associated pneumonia.

# Some potential approaches

- **Hospital acquired pneumonia (including VAP)**
  - guidelines: 1. target organisms

**Table 3. Current guidelines for the empirical treatment of hospital-acquired pneumonia, including ventilator-associated pneumonia**

	Early-onset HAP/VAP ( $\leq$ Day 4 of hospital stay/MV) without risk factors for MDR pathogen <sup>a</sup>	Late-onset HAP/VAP ( $\geq$ Day 5 of hospital stay/MV) OR presence of $\geq 1$ risk factor for MDR pathogen <sup>b</sup>
Most common (i.e. targeted) pathogens	MSSA	MRSA
	<i>Streptococcus pneumoniae</i> and other streptococci <i>Haemophilus</i> sp.	<i>Pseudomonas aeruginosa</i>
	Wild-type <i>Enterobacteriaceae</i>	Drug-resistant <i>Enterobacteriaceae</i> <i>Acinetobacter baumannii</i> <i>Stenotrophomonas maltophilia</i> .

# Some potential approaches

- Hospital acquired pneumonia (including VAP)**

- guidelines: 2. Societies' recommendations for "early onset" HAP/VAP without risk factors for MDR pathogen (\*)

British Society of Antimicrobial Chemotherapy (2008)	American Thoracic Society/Infectious Diseases Society of America (2005)	European Respiratory Society/ European Society of Clinical Microbiology and Infectious Diseases/European Society of Intensive Care Medicine (2009)
<ul style="list-style-type: none"> <li>• aminopenicillin/beta-lactamase inhibitor, <b>or</b></li> <li>• cefuroxime</li> </ul>	<ul style="list-style-type: none"> <li>• ceftriaxone, <b>or</b></li> <li>• levofloxacin, moxifloxacin, or ciprofloxacin, <b>or</b></li> <li>• ampicillin/sulbactam, <b>or</b></li> <li>• ertapenem</li> </ul>	<ul style="list-style-type: none"> <li>• ampicillin/sulbactam or amoxicillin/clavulanate, <b>or</b></li> <li>• cefuroxime, cefotaxime or ceftriaxone, <b>or</b></li> <li>• moxifloxacin or levofloxacin (not ciprofloxacin)</li> </ul>

\* add vancomycin or linezolid if MRSA is suspected

Adapted from Barbier, et al. *Curr Opin Pulm Med.* 2013;19:216-28

See also *Am J Respir Crit Care Med.* 2005;171:388–416 / *JAC.* 2008;62:5–34 / *Intensive Care Med* 2009; 35:9–29

# Some potential approaches

- Hospital acquired pneumonia (including VAP)**

- guidelines: 2. Societies' recommendations for "late onset" HAP/VAP or with  $\geq 1$  risk factors for MDR pathogen

British Society of Antimicrobial Chemotherapy (2008)	American Thoracic Society/Infectious Diseases Society of America (2005)	European Respiratory Society/ European Society of Clinical Microbiology and Infectious Diseases/European Society of Intensive Care Medicine (2009)
<b>early onset with risk MDR</b> <ul style="list-style-type: none"> <li>• ceftaxime or ceftriaxone <b>or</b></li> <li>• fluoroquinolone <b>or</b></li> <li>• piperacillin/tazobactam</li> </ul>	<ul style="list-style-type: none"> <li>• cefepime or ceftazidime, <b>or</b></li> <li>• imipenem or meropenem, <b>or</b></li> <li>• piperacillin/tazobactam, <b>or</b></li> <li>• ciprofloxacin or levofloxacin <b>or</b></li> <li>• amikacin or gentamicin or tobramycin</li> </ul>	<ul style="list-style-type: none"> <li>• ceftazidime <b>or</b></li> <li>• imipenem or meropenem <b>or</b></li> <li>• piperacillin/tazobactam + ciprofloxacin or levofloxacin</li> </ul>
<b>late onset</b> <ul style="list-style-type: none"> <li>• use local epidemiology</li> <li>• if <i>P.aeruginosa</i>: ceftazidime, ciprofloxacin, meropenem or piperacillin/tazobactam</li> </ul>		

\* add vancomycin or linezolid if MRSA is suspected

Adapted from Barbier *et al* Curr Opin Pulm Med. 2013;19:216-28

See also *Am J Respir Crit Care Med* 2005;171:388–416 / *JAC*. 2008;62:5–34 / *Intensive Care Med*. 2009; 35:9–29

# Guidelines: Local vs General

- The empiric algorithm derived from analysis of local microbiologic data predicted significantly better coverage than one defined by an unmodified guideline-driven approach for early HAP/VAP. Our locally-derived TICU algorithm of ceftriaxone+vancomycin for early pneumonia and piperacillin-tazobactam+vancomycin for late pneumonia optimizes the adequacy of initial therapy. Understanding local patterns of pneumonia ensures the creation and maintenance of empiric algorithms that achieve the best clinical outcomes.

**Becher RD, Hoth JJ, Rebo JJ, Kendall JL, Miller PR. Locally derived versus guideline-based approach to treatment of hospital-acquired pneumonia in the trauma intensive care unit. *Surg Infect (Larchmt)*. 2012 Dec;13(6):352-9. doi: 10.1089/sur.2011.056. PubMed PMID: 23268613.**

**1: Dalhoff K, Abele-Horn M, Andreas S, Bauer T, von Baum H, Deja M, Ewig S, Gastmeier P, Gatermann S, Gerlach H, Grabein B, Höffken G, Kern WV, Kramme E, Lange C, Lorenz J, Mayer K, Nachtigall I, Pletz M, Rohde G, Rosseau S, Schaaf B, Schaumann R, Schreiter D, Schütte H, Seifert H, Sitter H, Spies C, Welte T; German Society for Anaesthesiology and Intensive Care Medicine; German Society for Infectious Diseases; German Society for Hygiene and Microbiology; German Respiratory Society; Paul-Ehrlich-Society for Chemotherapy. [Epidemiology, diagnosis and treatment of adult patients with nosocomial pneumonia. S-3 Guideline of the German Society for Anaesthesiology and Intensive Care Medicine, the German Society for Infectious Diseases, the German Society for Hygiene and Microbiology, the German Respiratory Society and the Paul-Ehrlich-Society for Chemotherapy]. *Pneumologie*. 2012 Dec;66(12):707-65. doi: 10.1055/s-0032-1325924. Epub 2012 Dec 6. German. PubMed PMID: 23225407. <https://www.thieme-connect.com/DOI/DOI?10.1055/s-0032-1325924>**

# Haemophilus: is it important ?



<http://www.pathologyoutlines.com/topic/lymphnodeshinfluenzae.html>

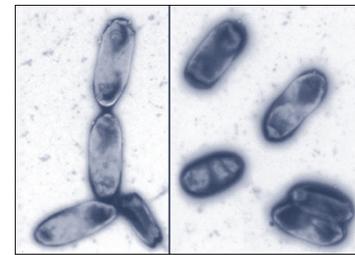
- Haemophilus is often considered as a colonizer of the upper respiratory tract with risks only for patients with COPD
- However, in coinfection with a preceding viral infection, Haemophilus may colonize the lung, leading to lethal secondary bacterial pneumonia.

## Genome-wide fitness profiling reveals adaptations required by *Haemophilus* in coinfection with influenza A virus in the murine lung

Sandy M. Wong<sup>a</sup>, Mariana Bernui<sup>b</sup>, Hao Shen<sup>b,1</sup>, and Brian J. Akerley<sup>a,1</sup>

<sup>a</sup>Department of Microbiology and Physiological Systems, University of Massachusetts Medical School, Worcester, MA 01605; and <sup>b</sup>Department of Microbiology, University of Pennsylvania Perelman School of Medicine, Philadelphia, PA 19104

# Haemophilus: this may be why !



<http://www.pathologyoutlines.com/topic/lymphnodesinfluenzae.html>

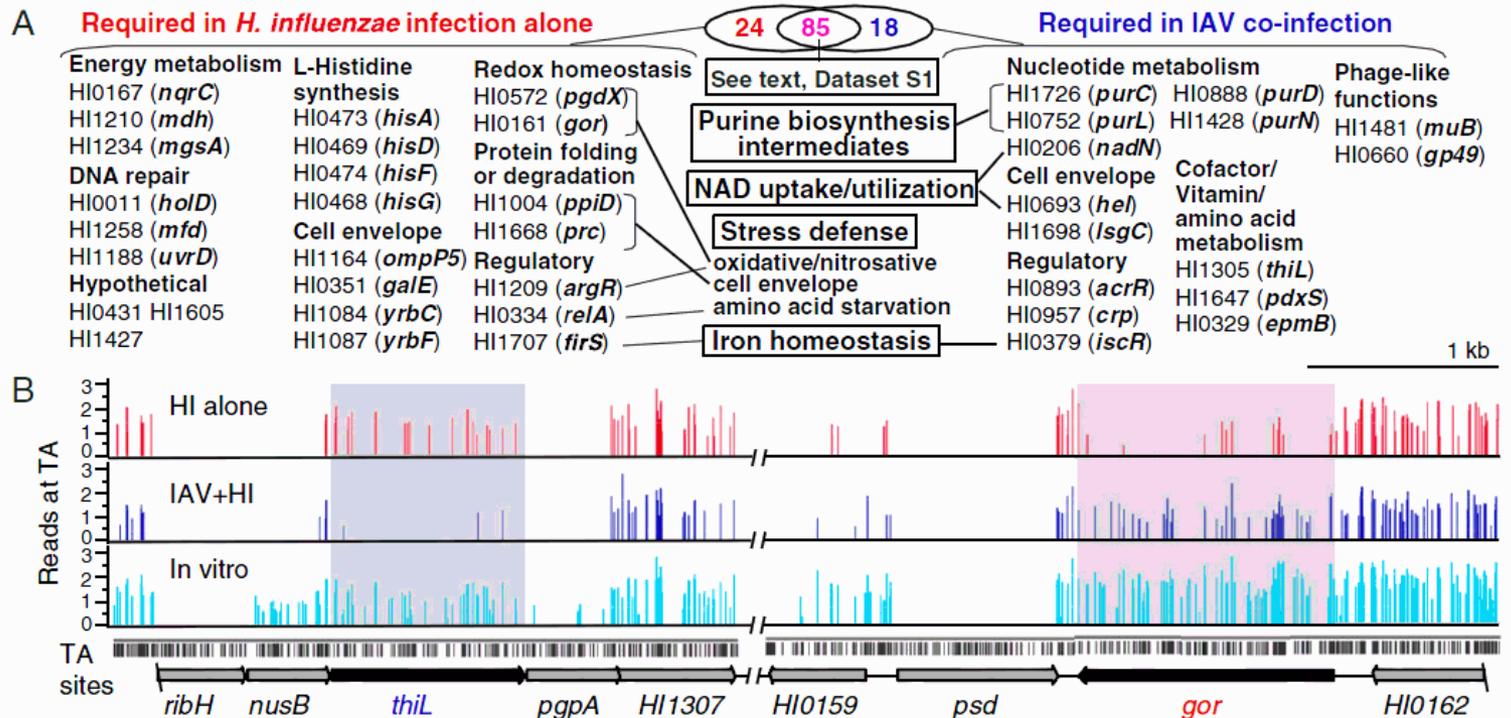
- Haemophilus with risks
- However, colonize

## Genome-wide required for influenza

Sandy M. Wong<sup>a</sup>, Mar

<sup>a</sup>Department of Microbiology, University of P

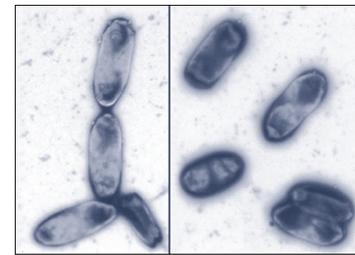
Proc Natl Acad Sci U S



**Fig. 1.** Genome-scale evaluation by HITS of the effect of IAV coinfection on lung colonization by *H. influenzae* mutants. (A) Coinfection alters survival requirements of *H. influenzae* in the lung. Candidate virulence genes with no observed effects on fitness in vitro were sorted into categories based on their roles in fitness detected in vivo. Genes required in both in vivo conditions are listed in Dataset S1 and discussed in the text. (B) Representative HITS data preinfection (in vitro) and postinfection with *H. influenzae* alone (HI alone) or IAV/*H. influenzae* coinfection (IAV+HI) for loci containing *thiL* and *gor*. Colored bars on the x axes designate sites of transposon insertions detected via sequencing and heights indicate relative abundance of insertions detected at each site (y axis:  $\log_{10}$ -transformed reads). Black bars below the x axis represent genomic TA dinucleotide positions.

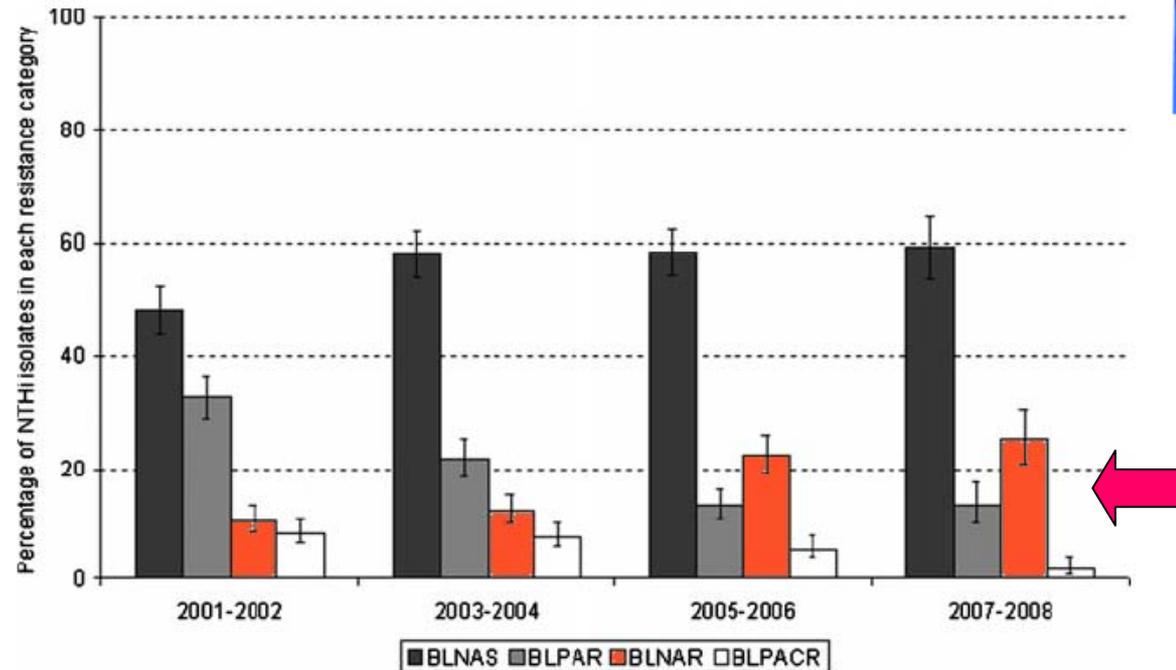
# Haemophilus and resistance: recto

## Are $\beta$ -lactamase-negative ampicillin-resistant (BLNAR) isolates important ?



<http://www.pathologyoutlines.com/topic/lymphnodeshinfluenzae.html>

**Fig. 1** Evolution of  $\beta$ -lactam resistance of all NTHi strains isolated from children aged 5 years or less in France, from 2001 to 2008. NTHi=non-typeable *Haemophilus influenzae*; BLNAS =  $\beta$ -lactamase-negative ampicillin-susceptible; BLPAR =  $\beta$ -lactamase-positive ampicillin-resistant; BLNAR =  $\beta$ -lactamase-negative ampicillin-resistant; BLPACR =  $\beta$ -lactamase-positive amoxicillin-clavulanic acid-resistant



Dabernat et al. *Eur J Clin Microbiol Infect Dis.* 2012;31:2745-53

**Warning:** antibiotic discs may fail to fully separate between BLNAS and BLNAR populations

(Garcia-Cobos, et al. *JAC.* 2013;68: 159-63)

**Good news:** The majority of invasive *H. influenzae* including BLNAR remain susceptible to third-generation cephalosporins

(Garcia-Cobos, et al. *JAC.* 2014;69:111-6)

See also Puig, et al *PLoS One.* 2013;13-8:e82515 for clinical success with ceftriaxone and fluoroquinolones

# Haemophilus and resistance: verso

## But other regions may be spared



<http://www.pathologyoutlines.com/topic/lymphnodeshinfluenzae.html>

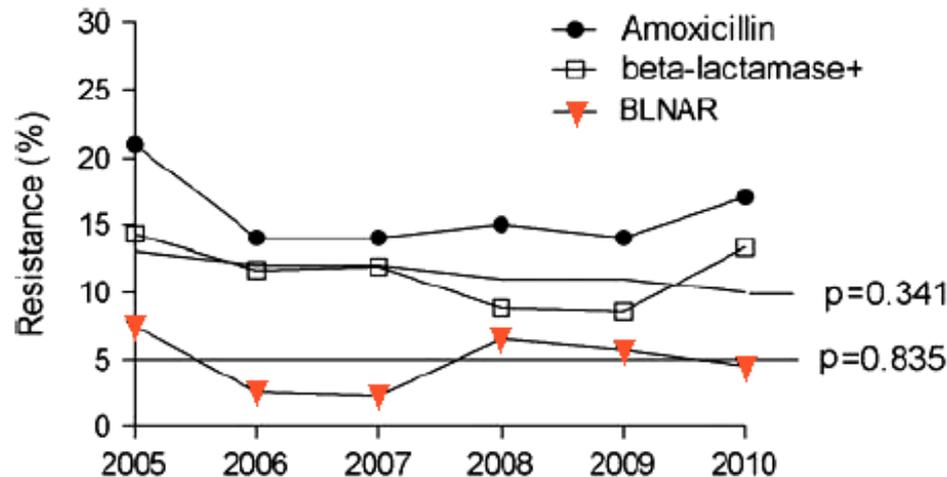
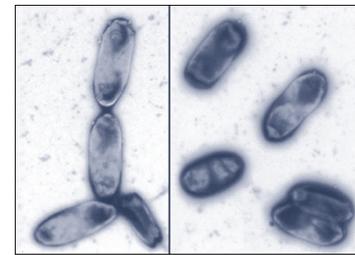


Figure 3. Beta-lactamase-positive and BLNAR isolates over a 6-y period. Division of amoxicillin-resistant strains into BLNAR isolates and beta-lactamase-positive isolates. BLNAR prevalence was approximately 5% over the 6 y and trend analysis showed a stable trend over time. The prevalence of beta-lactamase-positive isolates was approximately 11% over the studied time period. The logistic regression trend line showed a decrease over time, although no statistical difference over time was detected ( $p = 0.341$ ).

# Haemophilus and fluoroquinolones

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Original article

A molecular analysis of quinolone-resistant *Haemophilus influenzae*: Validation of the mutations in Quinolone Resistance-Determining Regions

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Shoji H, et al. *J Infect Chemother.* 2014;20:250-5

We confirmed that these five mutations strongly contribute to quinolone resistance and found that the degree of resistance is related to the number of the mutations. In addition, the three strains of 18 susceptible strains (16.7%) also had a single mutation. These strains may therefore be in the initial stage of quinolone resistance. Currently, the frequency of quinolone-resistant *H. influenzae* is still low. However, as has occurred with  $\beta$ -lactams, an increase in quinolone use may lead to more quinolone-resistant strains.

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